



AF/1646
JFW

In Re Application of: James P. Elia)
Serial No.: 09/836,750) Group Art Unit: 1646
Filed: April 17, 2001) Examiner: Elizabeth C. Kemmerer
For: METHOD FOR REPAIRING) Client Docket No. 1000-10-CO1
A DAMAGED PORTION)
OF A HUMAN HEART)

MAIL STOP APPEAL BRIEF-PATENTS
Commissioner for Patents
P.O. Box 1450
Alexandria, VA 22313-1450

**APPELLANT'S RESPONSE TO
NOTIFICATION OF NON-COMPLIANT APPEAL BRIEF**

Dear Sir:

This paper responds to the October 5, 2007 Notification of Non-Complaint Appeal Brief (37 C.F.R. 41.37) (hereinafter "Notification") in which the Appeal Brief filed February 23, 2007 was deemed to be defective because:

After consultation with SPE Gary Nickol and Deputy Chief Appeals Administrator Patrick Nolan, it was determined that the brief of 2/23/07 is not compliant in view of the improper request to the Board to consider claim 246 as depending on appealed claim 244 when, in fact, claim 246 actually depends from claim 245. Furthermore, the conditional indication that claim 246 would be amended "at an appropriate time" was also deemed to be improper. Finally, it was determined that Appellant should be advised that claims 245, 248, 249, 252, 264-267, 272-279, 286, and 287, which were indicated as withdrawn from Appeal by Appellant, would stand or fall with claims 236, 238, 239, 243, 244, 246, 247, 250, 251, 253, 257-263, 268-271, and 280-295 on the issue of

enablement since they are still officially pending, under examination, and rejected for lack of enablement on the same grounds as the other pending and examined claims.

Appellant has considered the above advisement set forth in the Notification.

In response to the Notification, Appellant has cancelled claims 245, 246, 248, 249, 252, 264-267, 272,-279, 286, and 287 in an Amendment mailed on October 11, 2007. Entry of such Amendment by the Examiner is in order because it conforms with the requirements of 37 CFR 41.33(b)(1) and 37 CFR 1.116 by cancelling claims and removing issues; and, therefore, it is believed to render the concurrently-submitted Amended Appellant's Appeal Brief compliant under 37 CFR 41.37.

Cancellation of the above claims eliminates all issues raised in the advisement in regard to the status of the claims on appeal.

Appellant believes that the Amended Appellant's Appeal Brief eliminates any unintended procedural problems present in Appellant's Appeal Brief filed on February 23, 2007.

Respectfully submitted,

Dated: 10/22/07

Gerald K. White

Gerald K. White
Reg. No. 26,611
Attorney for Appellant

GERALD K. WHITE & ASSOCIATES, P.C.
205 W. Randolph Street
Suite 835
Chicago, IL 60606
Phone: (312) 920-0588
Fax: (312) 920-0580
Email: gkwpatlaw@aol.com



IN THE UNITED STATES PATENT AND TRADEMARK OFFICE
BOARD OF PATENT APPEALS AND INTERFERENCES

In Re Application of: James P. Elia)	Docket No.: 1000-10-CO1
)	
Serial No.: 09/836,750)	Group Art Unit: 1646
)	
Filed: April 17, 2001)	
)	
For: METHOD FOR GROWING)	Examiner: Elizabeth C. Kemmerer, Ph.D.
MUSCLE IN A HUMAN HEART)	

MAIL STOP APPEAL BRIEF
Commissioner for Patents
P.O. Box 1450
Alexandria, VA 22313-1450

APPELLANT'S AMENDED APPEAL BRIEF

CERTIFICATE OF MAILING

I hereby certify that the attached APPELLANT'S AMENDED APPEAL BRIEF was deposited, as First Class Mail, in an envelope addressed to: MAIL STOP APPEAL BRIEF, Commissioner for Patents, P.O. Box 1450, Alexandria, VA 22313-1450 this 22nd day of October, 2007.

Dated: 10/22/07

Gerald K. White
Printed Name: GERALD K. WHITE

GERALD K. WHITE & ASSOCIATES, P.C.
205 W. Randolph Street
Suite 835
Chicago, IL 60606
Phone: (312) 920-0588
Fax: (312) 920-0580
Email: gkwpatl@aol.com

TABLE OF CONTENTS

REAL PARTY IN INTEREST	1
RELATED PROCEEDINGS	2
STATUS OF CLAIMS AND CLAIMS UNDER APPEAL	3
STATUS OF AMENDMENTS	4
SUMMARY OF CLAIMED SUBJECT MATTER	5
GROUND OF REJECTION FOR REVIEW ON APPEAL	8
ARGUMENT	9
Rejection of Claims 236, 238, 239, 243, 244, 246, 247, 250, 251, 253, 257-263, 268-271, and 280-285 under 35 U.S.C. §112, first paragraph	9
CONCLUSION AND RELIEF SOUGHT	32
CLAIMS APPENDIX	33
EVIDENCE APPENDIX	36
RELATED PROCEEDINGS APPENDIX	38

REAL PARTY IN INTEREST

The real parties in interest in the instant appeal are Assignees, Dental Marketing Specialists, Inc., an Arizona corporation, 9377 E. Bell Road, Suite 385 Scottsdale, Arizona 85260, and Jerry W. Bains and Salee C. Bains Irrevocable Trust, 9013 Red Lawrence Drive, Carefree, Arizona 85377. Subsequent to the assignment recordal for the instant application, the address of Dental Marketing Specialists, Inc., changed to 7364 East Crimson Sky Trail, Scottsdale, Arizona 85262. Also, subsequent to the assignment recordal for the instant application, the address of Jerry W. Bains and Salee C. Bains Irrevocable Trust, changed to 39096 N. 102nd Way, Scottsdale, Arizona 85262.

RELATED PROCEEDINGS

There is one appeal proceeding known to Appellant's legal representatives, which may be related to, directly affect, or may have a bearing upon the Board's decision in the pending appeal. Such appeal proceeding is that taken for co-pending Serial No. 09/794,456, filed February 27, 2001, in which Appellant's Brief was filed with the Patent and Trademark Office on February 8, 2007. There are no related interferences or judicial proceedings known to Appellant, Appellants' legal representatives, or Assignee, which may be related to, directly affect, be directly affected by, or may have a bearing on the Board's decision in the pending appeal. The attached Related Proceedings Appendix confirms the above statements.

STATUS OF CLAIMS AND CLAIMS UNDER APPEAL

Claims 1-5 were cancelled in the Preliminary Amendment filed April 17, 2001.

Claims 204, 205, and 237 were cancelled in the Amendment filed February 17, 2004.

Claims 245, 246, 248, 249, 252, 264-267, 272-279, 286, and 287 were cancelled in the Amendment mailed October 11, 2007. Such or similar claims are pending in co-pending applications Serial No. 11/605,153, filed November 28, 2006 and Serial No. 09/794,456, filed February 27, 2001. By cancelling such claims, Appellant has chosen to reduce the number of issues for the instant appeal and does not acquiesce to, or in any way agree with, the correctness of any rejection of these claims in the prosecution of the present application.

Claims 254-256 were cancelled in the Amendment filed November 21, 2005.

Claims 6-235 and 240-242 stand withdrawn, by the Examiner, from consideration as being directed to a non-elected invention. Claims 204 and 205 were cancelled by Appellant and thus were incorrectly identified by the Examiner as being withdrawn.

In view of the cancellation of the above-identified claims, the correctness of the Examiner's Final Rejection of claims 236, 238, 239, 243, 244, 247, 250, 251, 253, 257-263, 268-271, and 280-285 under 35 U.S.C. §112, first paragraph, for lack of enablement constitutes the sole issues on appeal.

STATUS OF AMENDMENTS

An Amendment cancelling claims 245, 246, 248, 249, 252, 264-267, 272-279, 286, and 287 was mailed on October 11, 2007.

SUMMARY OF CLAIMED SUBJECT MATTER

Appellant's invention is directed to a method of using well-known compositions (materials), old and well-known administration techniques for such compositions, and equally old and well-known medical apparatus to produce a novel result, i.e., the use of growth factors, including a cell (stem cell), such as bone marrow stem cells ("BMC's"), to grow a new artery and new cardiac muscle in the heart of a human patient and also to growing such new artery and cardiac muscle and repair a dead or damaged portion of a heart. Antecedent bases in the specification for various claim elements are included below.

Appellant's novel contribution to the medical art is defined in the broadest scope in generic claim 236 on appeal as comprising a method for growing a new portion of a human heart by placing a growth factor in the body of a human patient and forming a new artery and new cardiac muscle thereby and claims 238 and 239 cause repair of dead and damaged portions of the heart with the growth of new arteries and cardiac muscle (page 45, lines 17-22; page 46, lines 3-14). A growth factor, as called for by claim 236, broadly encompasses compositions and living organisms, which promote the growth of soft tissue in the body of a patient (page 20, lines 10-14). Appellant's specification on page 21 broadly recites that, "The growth factor can be administered orally, systemically, in a carrier, by hypodermic needle...or by any other desired method." Appellant's invention specifically describes using patient size, vascularity, simplicity of access, ease of exploitation, or any other desired factors in determining the selected area of the patient for administering said growth factor (page 45, lines 1-16). Appellant describes dosages

of growth factors useful for achieving growth of a new artery and achieving heart repair as defined in claims 236, 238, and 239 (page 53, lines 13-19; page 56, lines 7-19; and page 62, lines 1-10) and describes monitoring heart repair by determining blood flow through the new artery by using any readily available commercial device such as ultrasound, angiogram, etc. (page 56, lines 20-25). The specification on page 47 discloses that booster shots of growth factor may be required to repair an organ that is not operating at a desired capacity.

Appellant's elected invention is defined in claim 243, which directly depends from claim 236, and specifically limits the growth factor to a subgenus comprising a member selected from the group consisting of cells, cellular products, and derivatives of cellular products (page 37, lines 19-26 and page 47, line 22 to page 48, line 15). Claims 244 and 271 further limit the invention by specifying that the growth factor of claims 243 and 259, respectively, comprises a cell (page 37, lines 19-26). Claims 247, 268, and 269 directly depend from claims 236, 262, 263, respectively and further limit the method of said claims 236, 262, and 263, respectively, by reciting that the growth factor is placed in said patient by injection (page 21, line 5; page 45, line 14); and claim 251 depends from and further limits the method of claim 236 by requiring the growth factor be placed in said patient by a carrier (page 21, lines 3-6). Claim 253 depends from claim 236 and requires that the growth factor comprises a gene and a cell (page 46, lines 6-9). Claims 257, 258, 259, and 260 directly depend from and further limit claims 236, 238, 239, and 243, respectively, by requiring that the growth factor is locally placed in the human body (page 21, lines 4-10 and page 46, lines 3-9). Claims 261, 262, and 263 directly depend from and further limit claim 236, 238, and 239, respectively, by requiring that the growth

factor involved in the growth of new arteries and new cardiac muscle (claim 261), the repair of dead portions of the heart (claim 262), and the repair of damaged portions of the heart (claim 263) comprises living stem cells harvested from bone marrow (page 40 lines 27 – page 42, line 30). Claims 270 and 271 directly depend from claims 258 and 259, respectively and require placing a cell into the heart adjacent a dead (claim 270) or damaged (claim 271) portion of the heart causing new arteries and new cardiac muscle to be grown and repair of dead (claim 270) and damaged (claim 271) portions of a human heart to be repaired (pages 45 and 46). Claims 280, 281, and 282 directly depend from claims 236, 238, and 239, respectively and require calculating blood flow through the newly grown artery (page 56, lines 20-25). Such calculation provides an indication of artery growth. Claims 283, 284, and 285 directly depend from claims 236, 238, and 239, respectively, and require observing the newly grown arteries (page 56, lines 20-25). Such observation provides a description of artery growth.

GROUND OF REJECTION FOR REVIEW ON APPEAL

Appealed claims 236, 238, 239, 243, 244, 247, 250, 251, 253, 257-263, 268-271, and 280-285 stand finally rejected under 35 U.S.C. §112, first paragraph, as failing to comply with the enablement requirement.

ARGUMENT

Rejection of Claims 236, 238, 239, 243, 244, 247, 250, 251, 253, 257-263, 268-271, and 280-285 Under 35 U.S.C. §112, first paragraph

As a preliminary matter, Appellant notes that the cancellation of claims 245, 246, 248, 249, 252, 264-267, 272-279, 286, and 287 renders moot the Final Rejection of claims 248, 249, 252, and 274-279 under 35 U.S.C. §112, first paragraph, new matter; the Final Rejection of claim 245 under 35 U.S.C. §112, second paragraph, indefinite; and the Final Rejection of claims 286 and 287 for double patenting.

The Examiner finally rejected appealed claims 236, 238, 239, 243, 244, 247, 250, 251, 253, 257-263, 268-271, and 280-285, “under 35 U.S.C. 112, first paragraph, as failing to comply with the enablement requirement.” Specifically, the Examiner, on pages 7 and 8 of the Final Rejection dated September 22, 2006, states that:

The claim(s) contains subject matter which was not described in the specification in such a way as to enable one skilled the art to which it pertains, or with which it is most nearly connected, to make and/or use the invention.

Appellant disagrees that the scope of protection provided by the appealed claims is not adequately enabled by the application disclosure. Appellant intends to, and does hereinafter, argue the patentability of each claim separately, i.e., the patentability of the claims on appeal do not stand or fall together.

At the outset, Appellant believes that there are three important factors to consider when determining whether the instant specification contains a disclosure that would have enabled a skilled person in the medical art to make and use the claimed invention. When these factors are considered, there can be no doubt that Appellant’s specification provides an enabling disclosure. The three factors are discussed below.

First, as pointed out in the Summary of Claimed Subject Matter portion of the instant Brief, there is a considerable body of disclosure relating to Appellant's generic invention of repairing organs in human patients, including the heart, by forming a new cardiac muscle and artery and of elected and non-elected growth factors suitable for effecting such repair and formation. In this regard, Appellant's specification provides a substantial body of disclosure regarding growing and/or replacing organs and/or forming arteries and tissues using well-known growth factors. The specification describes a class of growth factors that broadly and specifically includes genes, nucleic acids, a patient's own cells, or universal cells, e.g., stem cells, etc., all of which are described to promote tissue growth through differentiation and morphogenesis. The Examiner has only considered the disclosure regarding enablement as it specifically relates to the elected growth factor species, cells. The Examiner's selective reading, which ignores Appellant's broad and specific disclosure relating to non-elected growth factor species disclosure, is clearly erroneous under relevant case law. When an applicant elects to prosecute a species following an election requirement, the Examiner is not permitted to wear blinders and focus solely upon the elected species and ignore the scope of enablement provided by the specification as a whole. There should be no doubt that the specification taken as a whole, when properly read and understood by one skilled in the art, meets the statutory requirement for enablement under current law.

It is clear that the Examiner intended to so erroneously limit her review of the application disclosure from the statement at page 6, lines 1-8 of the February 22, 2006,

Office Action in co-pending patent application Serial No. 09/794,456, filed February 27, 2001 that:

The claims are being examined to the extent that they read on the elected invention, administration of cells, and thus the generic concept of growth factor is not relevant. (emphasis added)

It is evident that the Examiner also erroneously ignored disclosure related to non-elected species, such as genes and proteins, in evaluating enablement. In this regard, for example, please see page 10, lines 4-7 and page 12, lines 14-21 of the Final Rejection of September 22, 2006.

Second, the Examiner has not taken issue with the fact that the administration techniques and administered materials disclosed by Appellant were individually old and well known as of the filing date of the instant patent application. The materials and administration techniques disclosed by Appellant were routinely employed in the medical art, but not in the claimed combination, at the time the instant application was filed. Appellant's contribution to the medical arts resides in the claimed method of treating a heart in a human patient by implanting cells in said patient and growing cardiac muscle and an artery in said heart.

Third, the Examiner stated that the level of skill in the art is high. Appellant agrees that the skill level is high when it is considered that many years of education, training, and experience are required in the medical field.

Once the above-identified relevant materials and administration techniques set forth in Appellant's specification are properly considered in their entirety, Appellant believes that there should be no question that one skilled in the medical art is enabled to make and use the claimed invention. This conclusion is reinforced, as noted above, by

the fact that the materials and administration techniques, but not the inventive results, were well known when the instant application was filed. MPEP Section 2164 states that the purpose of the enablement requirement is to describe the claimed invention in such terms to permit one skilled in the art to make and use the invention. Such Section cautions that detailed procedures for making and using the invention may not be necessary if the description of the invention itself is sufficient to permit those skilled in the art to make and use the invention. MPEP Section 2164.01 states that:

A patent need not teach, and preferably omits, what is well known in the art. *In re Buchner*, 929 F2d. 660, 661, 18 USPQ 2d 1331, 1332 (Fed. Cir. 1991); *Hybritech, Inc. v. Monoclonal Antibodies, Inc.*, 802 F2d. 1367, 1384, 231 USPQ 81, 94 (Fed. Cir. 1986) cert denied, 480 U.S. 947 (1987); and *Lindemann Maschinenfabrik GMBH v. American Hoist and Derrick Co.*, 730 F2d. 1452, 1463, 221 USPQ 481, 489 (Fed. Cir. 1984).

Appellant believes that the above caution is especially relevant to the instant factual situation where the Examiner has conceded that there was a high level of skill in the art at the time the instant application was filed, coupled with the fact that the Examiner has not taken issue with Appellant's position that all the materials, methods, and apparatus needed to practice the invention were well known at the time of the invention. Thus, Appellant submits that the instant disclosure clearly enables one skilled in the medical arts to make and/or use the full scope of the claimed invention without undue experimentation because a reasonable consideration of the three above-delineated factors and the interaction thereof leads to the inevitable conclusion that the disclosure is enabling.

The Examiner has the burden to establish and support by convincing objective evidence a *prima facie* case of lack of enablement. For reasons set forth below, Appellant believes the Examiner has failed to meet such burden.

The Board's attention is respectfully directed to page 11 of the Final Rejection, where the Examiner concluded:

Due to the large quantity of experimentation necessary to determine how to effectively administer cells to achieve *de novo* formation of cardiac muscle and an artery and thereby grow a new portion of a pre-existing heart, the lack of direction/guidance presented in the specification regarding the same, the absence of working examples directed to the same, the complex nature of the invention, the contradictory state of the prior art, the unpredictability of the effects of an agent on a physiological response, and the breadth of the claims which fail to recite limitations regarding cell type or dosage or site of delivery, etc., undue experimentation would be required of the skilled artisan to make and/or use the claimed invention in its full scope (emphasis added).

As plainly evident, the Examiner's conclusion is of a conditional nature, i.e., the rejection depends upon the achievement of *de novo* cardiac muscle and arteries. However, neither the specification nor the claims are directed to *de novo* cardiac muscle and arteries. The condition established by the Examiner to measure enablement of the claims, i.e., the achievement of *de novo* cardiac muscle and arteries, is not present in the claims; and the rejection must be reversed as not being relevant to the claimed invention.

Appellant submits that the construction of the appealed claims accorded by the Examiner on pages 8 and 11 of the Final Rejection of September 22, 2006 as requiring *de novo* cardiac muscle and artery formation is clearly erroneous. The Examiner's statement that, "Applicant has defined a new artery as an organ comprising two or more kinds of tissue..." is incorrect and misleading (emphasis added). The specification on page 44

defines the term “organ” used by Appellant as consisting “of two or more kinds of tissues joined into one structure that has a certain task.” The Examiner’s statement that, “Applicant appears to imply that the ‘new artery’ recited in the claims must be formed *de novo*, and not merely repair, growth or re-direction of an existing artery,” is also incorrect and misleading. Appellant made no such statement or implication. Appellant’s specification at pages 45-50 describes that growth factors are utilized to “grow” and for “forming” new muscles and arteries and to facilitate differentiation into organs or function specific tissues, including muscle tissue and arteries. The specification at pages 54, 56, and 62 clearly defines the claimed term “new artery,” and the scope of the claims is legally determined by this disclosure. It is clear from such disclosure what Appellant intended the term “new artery” to mean. See Philips v. AWH., Corp., 03-1269-1286, decided July 12, 2005.

The Examiner, not Appellant, appears to have coined the term “*de novo* artery” and thus becomes its lexicographer. The Examiner has not explained what is encompassed by such term nor has she provided any evidence in this record which defines or recognizes the scientific relevance of such term. Further, the Examiner has failed to point to any language in the claims or description in the subject application limiting Appellant’s claimed invention to the formation of *de novo* arteries. The Examiner’s attempt to support her position by stating at page 8 of the Final Rejection, “See the discussion regarding fusion versus formation of new cells,” (referring to Appellant’s February 17, 2004 Amendment) does not clearly identify any alleged implication of Appellant that a new artery must be formed *de novo*. Appellant has reviewed the February 17, 2004 Amendment and finds that it used the term “fusion” at

pages 42 and 45. Such usage is unrelated to the term *de novo* arteries, and thus Appellant is at a loss as to how to respond to the Examiner's statement. Appellant stands ready to respond, via a Reply Brief, to any other portion once it is identified by the Examiner. The Examiner has taken Appellant's remarks in the above-mentioned Amendment out of context. Appellant's remarks in that response were directed to distinguishing the present invention from the Murry et al. (hereinafter "Murry" and of record) reference's implantation of skeletal satellite (stem) cells in murine models, which did not result in artery growth. Appellant did not explicitly or implicitly argue that the present invention distinguishes over Murry by *de novo* growth of arteries.

Accordingly, the Examiner's enablement determination is conditional to the extent it is bottomed on the specification's failure to provide enablement for the achievement of *de novo* arteries. The Examiner's conditional rejection is patently erroneous and, perforce, should be summarily reversed.

Even though Appellant believes that the Examiner committed reversible error in misconstruing the scope of the claims and conditionally rejecting the claimed subject matter based on such construction, a full treatment of the Final Rejection as best understood follows.

The first paragraph of the statute requires nothing more than objective enablement, and it is of no importance whether such teaching is set forth by use of illustrative examples or by broad terminology. As a general matter, an application disclosure, which contains a teaching of how to make and use the invention in terms which correspond in scope to those used in describing the invention sought to be patented, is considered to be in compliance with the enabling requirement of the statute. In re Marzocchi, 58 CCPA

1069, 439 F.2d 220, 169 USPQ 367, 369-370 (1971). Further, “Section 112 does not require that a specification convince persons skilled in the art that the assertions therein are correct.” [Emphasis added]. In re Robins, 429 F.2d 452, 166 USPQ 552 (CCPA, 1970).

When evaluating enablement, it is incumbent upon the Examiner to determine what subject matter each claim recites, i.e., the scope of protection sought for each claim. The scope of dependent claims are properly determined with respect to 35 U.S.C. §112, fourth paragraph. See MPEP Section 2164.08. It is clear that the Examiner’s analysis did not treat the subject matter of each claim separately or treat the dependent claims according to statutory mandate. The only claims that the Examiner addressed regarding the specifically claimed subject matter in the Final Rejection were cancelled claims 248, 249, and 274-277, which specifically recite intravenous, intraluminal, and angioplasty modes of delivery. The appealed claims do not specifically require such cancelled subject matter. The Examiner states that the underlying fact at issue is whether or not more than routine experimentation would be required to practice the claimed invention and addresses this issue by reference to the guidelines established in In re Wands, 858 F.2d 731, 737, 8 USPQ 2d 1400, 1404 (Fed. Cir. 1988). As evidence in support of her non-enablement determination, the Examiner relies on the Strauer et al. (hereinafter “Strauer”), Deb et al. (hereinafter “Deb”), and Murry publications (all of record). None of these publications specifically teaches injection of stem cells into a human patient and growing an artery as called for by claims 268 and 269.

Initially, Appellant points out that it is evident the Examiner failed to consider the disclosure provided by Appellant’s specification as a whole in determining compliance

with the enablement requirement of the statute. The appropriate factual determination is whether the instant specification reasonably teaches one skilled in the art how to make and use the claimed subject matter. As demonstrated above, the Examiner erroneously restricted the factual determination to the elected species of growth factor and, thusly, ignored those portions of the specification describing a broader generic invention and also ignored disclosure related to non-elected species. Appellant is entitled to have the entire disclosure considered in determining compliance with 35 U.S.C. §112, first paragraph. See In re Anderson, 471 F.2d. 1237, 176 USPQ 331, (CCPA 1973) and In re Johnson and Farnham, 558 F.2d 1008, 194 USPQ 187, 195 (CCPA 1977). Further, it is well settled that the test for enablement must take into consideration that which is known in the prior art – that a patent should preferably omit that which is well known/understood in the particular art to which the claims are directed. See MPEP Section 2164.01 and the authorities cited therein.

Appellant submits that a careful analysis of the Wands factors in light of the application disclosure compels a conclusion that undue experimentation would not be required to practice the claimed invention. Contrary to the Examiner's assertion, the Strauer publication does not disclose any experimental protocol required for practicing the invention. Rather, Strauer discloses following protocols that were routinely used in the art and employing an off-the-shelf angioplasty balloon catheter, such as, for example, a catheter of the type described in Nabel et al, U.S. Patent No. 5,328,470 (of record), for transplanting autologous bone marrow mononuclear cells to treat acute myocardial infarction (MI) in human patients. The concept of containment to prevent backflow and prolong contact time is clearly taught by Nabel et al. Thus, contrary to the Examiner's

assertions, it is clear that Strauer did not have to perform any experimentation in order to choose an appropriate delivery system or devise a containment system that would prevent backflow of cells and thus provide a prolonged time for cell implantation. Appellant's specification provides direction and guidance in regard to the well known use in the prior art of a containment system for controlling "carry away" and prolonged contact of cells in Examples 18 and 19 in regard to injection of cells. It is important to note that claims 247, 250, 268, and 269 specifically recite injecting cells, and the Examiner has not critically reviewed the enablement provided by Appellant's disclosure *vis-à-vis* the scope of these claims. The Examiner has conceded, at page 22 of the Final Rejection, that the administration of cells is old in the art. Strauer does not describe using any experimental protocol to determine appropriate cell population, i.e., there is no requirement for experimentation described by Strauer for determining or using specific bone marrow stem cell species. Regarding time of treatment, Strauer does not disclose that determining time of treatment required experimentation. Strauer (published in 2002) elected to treat patients following five to nine days after suffering an MI. Note that a later publication of Strauer, the 2005 Strauer publication (of record), discloses treating chronic MI in patients that had transmural MI some 27 months earlier. Again, no experimentation regarding treatment time was noted. It is evident that the time of treatment following an MI is not a critical variable and undue experimentation would not be required. To the extent that the Examiner may be relying on Murry's murine models to establish that the time of administration is critical, Appellant points out that Strauer 2005 is the "best evidence" in regard to whether time of treatment in human patients is critical. Strauer 2005 teaches that stem cells can be used to successfully treat MI in

human patients suffering either acute or chronic disease. Thus, the Examiner's conclusion that "great quantities of experimentation" would be required to practice the claimed invention is not supported on the record and is fatally flawed.

The Examiner's contention that, "The specification provides no guidance along the lines of the details worked out by Strauer," is misplaced. Firstly, none of the claims on appeal require the use of an angioplasty balloon catheter; and, therefore, it is improper for the Examiner to look solely to Strauer for guidance. Secondly, application Example 19 (page 56) describes a detailed regimen for treating a patient with a damaged heart by injecting a growth factor for promoting artery growth, which includes mode, dosage, and means for evaluating success of treatment. Specifically, Example 19 describes a regimen wherein cells are injected slowly over "a five second time period." The application disclosure also teaches, on pages 40-42, 47, and 48, utilizing autologous stem cells harvested from bone marrow and blood of the patient (self-cell therapy) or from cell cultures (allogenic) to grow organs, i.e., arteries, by differentiation and morphogenesis (page 48).

The Examiner states at pages 10 and 13 of the Final Rejection, "The specification contains only prophetic Examples," and that, "There are no working examples directed to administering stem cells to dead or damaged portions of a heart," but fails to explain how this supports a holding of non-enablement and, for good reason, since actual working examples are not a requisite for satisfying the enabling requirement of the statute. Actual working examples are not required if the invention is disclosed in a manner that one skilled in the art would be able to practice it. Section 2164.02 of the MPEP states that:

Compliance with the enablement requirement of 35 U.S.C.
112, first paragraph, does not turn on whether an example

is disclosed. An example may be “working” or “prophetic.” A working example is based on work actually performed. A prophetic example describes an embodiment of the invention based on predicted results rather than work actually conducted or results actually achieved.

The Examiner’s contention on page 10 of the Final Rejection that, “...none of the prophetic examples are directed to administration of cells to grow a new artery...” is not supported on the record. One skilled in the art reading the specification at page 46, lines 3-8 would reasonably understand that Appellant disclosed a method comprising seeding, e.g., injecting, appropriate cells (stem cells) or other growth factors to promote growth of cardiac muscle and blood vessels (arteries) in a damaged portion of a human heart. Furthermore, one reasonably skilled in the art would understand from page 47, line 22 through page 48, line 15 that a patient’s own (autologous) stem cells can be used to grow function specific tissue, such as muscle and arteries, *in vivo* through differentiation and morphogenesis. Those skilled in the art understand that “morphogenesis” is the formation and differentiation of tissues and organs and that an artery is an organ. See pages 33 and 48 of the specification.

Appellants agree with the Examiner that the medical arts in general are complex. However, while the physiological reactions involved may be complex, the practice of the claimed invention is straightforward. The called-for cells, e.g., stem cells, the methods of administering, and the particular apparatus required for administering the cells are old and well known in the medical arts.

The Examiner’s statement at page 10 of the Final Rejection that:

The state of the prior art does not support the specification’s (and claims’) assertion that a new artery can be grown. None of the numerous post-filing date

publications put on the record by Applicant to support enablement of the claimed invention report the *de novo* growth of an artery as defined by Applicant, including Strauer

evinces a misunderstanding of the manner in which the Court applied “the state of the art” factor in In re Wands. The Court applied this factor in determining whether all the materials and methods needed to practice the Wands invention were known in the art. Appellant relies upon the instant specification and not upon any “post-filing date publications” to support enablement. As discussed above, Appellant’s specification does not need to support enablement for *de novo* growth of arteries because the claims in issue do not recite or require such a limitation. Moreover, Applicant never defined a new artery as a *de novo* artery, as erroneously alleged by the Examiner.

The Examiner admits that the level of skill in the art was high at the time the instant application was filed.

The Examiner states that the claimed invention is unpredictable because of its use of physiological agents, citing supporting authority. However, the Examiner does not provide any succinct reasoning or evidence as to why one skilled in the art would doubt that the asserted scope of objective enablement in Appellant’s specification is not in fact commensurate with the scope of the claims. The specification provides multiple embodiments using multiple well-known species of growth factors for promoting artery growth employing multiple well-known administration modes carried out with multiple well-known apparatus.

The Examiner states at page 11, first paragraph, of the Final Rejection that:

The breadth of the claims is quite large. The elected invention is directed to a method of administering any type

of cell to an undefined area of a human body to grow new cardiac muscle and a new artery (of any type or location) to achieve growth of a new portion of a pre-existing heart.

Firstly, the claims in issue require cells, stem cells, and bone marrow stem cells. One of ordinary skill in the art appraised of the specification disclosure would readily comprehend the type of cell, i.e., stem cell, required for promoting morphogenesis, e.g., artery growth. Secondly, the claims require growing new cardiac muscle and a new artery which limits the selected placement area to those locations that result in cardiac muscle growth and artery growth in the patient's heart. Regarding the degree of repair, Examples 18 and 19 describe using readily available commercial devices to assess blood flow through the new artery and thus determine the degree of success. There is no requirement that an applicant's claims set forth the practical limits of operation for the invention. One must look to the specification, not the claims, in determining compliance with the first paragraph of the statute. It is clear that the Examiner, in determining compliance with the enablement requirement of the statute, has ignored the scope of enablement provided by the specification as a whole. The Examiner's limited evaluation of Appellant's specification constitutes reversible error. See In re Johnson and Farnham, supra., and In re Fuetterer, 319 F. 2d 259, 265, 138 USPQ 217, 223 (CCPA 1963), cited therein. Moreover, the Examiner's rejection fails to specifically address the inventions of claims 270 and 271 requiring placing cells adjacent dead or damaged portions of the human patient's heart and the inventions of claims 268 and 269 requiring injecting stem cells into a human patient to grow an artery.

As demonstrated earlier, none of the claims in issue require *de novo* formation of an artery; and thus the Examiner has conditioned the enablement rejection on a lack of

enablement regarding the unclaimed and undisclosed formation of *de novo* arteries. Being that the Examiner's stated condition has not been met, the rejection must fail.

Except for the unidentified "contradictory state of the prior art" and the failure of the claims to recite "dosages," the factors listed by the Examiner in the above conclusory statement correspond to the Wands factors, which were fully addressed and rebutted earlier. Again, the Examiner has provided no evidence to support her conclusion. Regarding the alleged "contradictory state of the prior art," the Examiner has failed to identify any pre- or post-filing publications presenting evidence in regard to the subject matter of the claims on appeal, which contradicts the objective enablement provided by the instant application. The references cited by the Examiner on page 13 of the Final Rejection to "show the state of the prior art" were specifically directed to the inventions defined in cancelled claims 248, 249, and 274-277, which claims are directed to subject matter that is not at issue in the present appeal. The Examiner failed to articulate how such prior art is relevant to establishing a *prima facie* case of lack of enablement of the subject matter called for by the appealed claims *vis-à-vis* the objective enablement provided by the instant specification. To the extent the Examiner is challenging the predictability of Appellant's described cardiac muscle and artery growth, the Perin et al. publication (hereinafter "Perin" and of record), which describes implanting BMC's to provide heart repair suffices to ally such challenge. Perin provides autopsy proof that such heart repair involves both cardiac muscle and artery growth. Regarding "prior art," none has been identified or cited by the Examiner against Appellant's claims and for good reason. Appellant was the first to disclose and claim a method for treating a human

heart by implanting cells, such as stem cells, and growing new cardiac muscle and arteries.

Regarding dosages, it is axiomatic that claims do not have to recite dosage levels where dosage levels would be understood by those skilled in the art. As succinctly stated in MPEP Section 2164.01(c):

It is not necessary to specify the dosage or method of use if it is known to one skilled in the art that such information could be obtained without undue experimentation. If one skilled in the art, based on knowledge of compounds having similar physiological or biological activity, would be able to discern an appropriate dosage or method of use without undue experimentation, this would be sufficient to satisfy 35 U.S.C. 112, first paragraph.

Prophetic Examples 18, 19, and 36 describe methods for carrying out the invention including dosage amounts for promoting artery growth and heart repair. Appellant's specification describes new artery growth and heart repair by direct injection of growth factor cells in dosage ranging from approximately 6.25×10^6 (Example 18 & 36) to approximately 12.5×10^6 (Example 19). Available off-the-shelf cDNA clones (nucleic acids) are directly injected into either the cardiac muscle (Example 19) or the coronary artery (Example 36). Each example describes repairing a damaged heart by forming a new artery which results in increased coronary blood flow. Each example also discloses slowly injecting the growth factor to avoid any carry away. Example 18 discloses that a containment system may be used. While these examples employ nucleic acids, one skilled in the art reading the specification, which teaches that cells, i.e., stem cells, possess equivalent activity to genes (nucleic acids) and other genetic material, in forming a new artery and repairing a dead or damaged portion of a heart, would be able to easily extrapolate the number on a weight basis of mononuclear cells required to obtain

equivalent results. Note in this regard that Strauer discloses injecting six (6) to seven (7) times with 1.5 to 4×10^6 cells without disclosing any difference in results over the entire dosage range. Therefore, there is no significant clinical difference between Appellant's 6.25 to 12.5×10^6 and Strauer's 9 to 28×10^6 dosage ranges. Further, such skilled person would understand that intravenous or intraluminal administration routes would generally require larger doses than the direct injection route of Examples 18, 19 and 36, and, for example, simply doubling the dosage to 12.5 to 25×10^6 cells would essentially encompass Strauer's entire range.¹ It is clear from Strauer that there is no risk for over-dosing, particularly using autologous BMC's, which are contemplated in Appellant's specification. cf. In re Bundy, 642 F. 2d 430, 434, 209 USPQ 48, 51-52 (CCPA 1981). The Examiner has failed to establish why one skilled in the art would not be able to extrapolate those examples across the entire scope of the claims. See MPEP Section 2164.02.

The limited evaluation of Appellant's specification performed by the Examiner in determining compliance with the enablement requirement of the statute fails to comport with current law and constitutes reversible error. In re Johnson and Farnham, supra. That the Examiner intended to so limit her review of the application disclosure was clearly demonstrated above.

When properly considered, there should be no doubt that the application's disclosure, including the above specifically identified passages, fully enables one skilled

¹ The conversion for dosages of nucleic acids to corresponding dosages of cells was conducted as follows. Examples 19 and 36 specified dosages of 500 micrograms (ug) and 250 ug, respectively. The weight of nucleic acids of an average cell was considered to equal 40 picograms (pg). The described dosages of 250 and 500 ug when converted to pg by multiplying by 10^6 equals 250×10^6 pg and 500×10^6 pg. Since nucleic acids of an average cell have an average weight of 40 pg, a conversion is made by dividing 250×10^6 and 500×10^6 by 40 to arrive at the equivalent cell dosages, which are 6.25×10^6 and 12.5×10^6 , respectively.

in the medical art to make and use the invention. The Board's attention is once again respectfully directed to Appellant's specification at page 21, lines 4-15, where it is disclosed that growth factors may be administered by a wide variety of techniques, including injection. In addition, the Board's attention is also directed to Appellant's specification at page 45, line 1-16, where it is disclosed that arteries may be grown in a heart by injecting a gene (non-elected species of growth factor) into muscle at a desired site. Moreover, Appellant's specification at page 46, lines 3-7, indicates that seeding with cells and/or genes adjacent a dead cardiac muscle grows new arteries. Seeding is a generic term that includes injection. Example 19 describes repairing a damaged portion of a human patient's heart by slowly injecting (to avoid leakage) a growth factor solution into the patient's cardiac muscle adjacent a clogged artery and growing a new artery and teaches testing for blood flow through the new artery using available commercial devices. Example 36 describes repeating the injection process of Example 18 for growing a new coronary artery, and both examples disclose dosages for obtaining the desired artery growth. That Examples 19 and 36 describe cDNA clones as growth factor species is not important because the specification is replete with disclosure that if cells, such as stem cells and more specifically a patient's own stem cells, are implanted/reimplanted differentiation and morphogenesis into an organ, i.e., artery, occurs. One reasonably skilled in the art appraised of such disclosure would readily be able to predict and comprehend that stem cell growth factors are equivalent to cDNA clones in providing the desired artery formation.

It is a fact that the disclosed administration techniques were well established in the medical art prior to Appellant's invention and must be considered in any evaluation of

enablement. It is also a fact that cells, including stem cells, were well known and characterized prior to Appellant's claimed invention. The Board is respectfully requested to take Official Notice of the fact that processing bone marrow and peripheral blood for recovering mononuclear stem cells was routine in the medical arts prior to Appellant's invention. Typically, bone marrow transplant procedures involved removing bone marrow from the patient which is filtered, treated, and transplanted immediately or frozen and stored for later use. Another established fact—that stem cell banks were created as early as the 1950's—indicates that those skilled in the medical art were familiar with harvesting, handling, culturing, preserving, separating, and storing, etc. such stem cells. The handling and treatment of cells, as well as stem cells, has been long known and practiced in the medical art prior to Appellant's filing date. In this regard, see the Trigg 2002 publication in Pediatric Transplantation entitled, "Milestones in the Development of Pediatric Hematopoietic Stem Cell transplantation – 50 Years of Progress" (of record). As mentioned in this publication, stem cell handling and preparation techniques have been known for decades. The publication provides evidence of the fact that stem cells harvested from bone marrow and from blood of patients has long been known and isolated as part of national blood bank programs. Further, the Board is referred to three publications (all of record); namely: (1) Areman, et al. 1990 publication in Prog. Clin. Biol. Res., entitled, "Automated Isolation of Mononuclear cells using the Fenwal CS3000 blood cell separator;" (2) Angelini, et al. 1990 publication in Haematologica, entitled, "Human bone marrow processing using Cobe 2991 and CS 3000 blood cell separators for further ex vivo manipulation;" and (3) Janssen, et al. 1992 publication in J Hematother, entitled, "Use of the Terumo SteriCell for the processing of bone marrow and peripheral

blood stem cells,” which confirm the above-mentioned facts. In addition, the Caplan 1991 publication in Journal of Orthopaedic Research entitled, “Mesenchymal Stem Cells” (of record) reported culturing human bone marrow and isolating mesenchymal stem cells for growing bone in murine models. Appellant believes that such evidence confirms the fact that cells, including stem cells, were well known and characterized prior to Appellant’s claimed invention.

Once the relevant materials and administration techniques set forth in Appellant’s specification are properly considered in their entirety, Appellant believes that there should be no question that one skilled in the medical art is enabled to make and use the claimed invention. This conclusion is reinforced by the fact that the materials and administration techniques, but not the inventive result, were well known when the instant application was filed.

Appellant submits that the Examiner has apparently failed to comprehend that Appellant has used old and routine administration techniques and old materials to achieve a remarkable new result, which is repair of dead or damaged heart tissue. Inasmuch as the claimed administration techniques and materials were well known to those skilled in the art, a person skilled in the medical art would not require an extensive, detailed description of such old elements of the invention and thus would be enabled to make and use the claimed invention once guided and directed to the administration techniques and materials by Appellant’s specification. It is noted that the Examiner has not challenged the fact that these elements were known as of the filing date of the instant application, and for good reason.

In summary, Appellant believes that the Examiner's evidence of lack of enablement, which comprises the Examiner's erroneous assessment of Strauer and Deb, as discussed above, when considered *vis-à-vis* the evidence of enablement provided by Appellant's specification combined with a fair and reasonable reading of Strauer and Deb, coupled with Wollert et al. (hereinafter "Wollert" and of record), fails to establish a *prima facie* case of lack of enablement under current law. Note that Wollert is cited as evidence that the high pressure technique of Strauer is not necessary, and thus the Examiner's contention that the instant disclosure's lack of including the post-filing administration technique of Strauer is nonenabling is patently erroneous. It has been further demonstrated above that the Examiner's conclusion is not supported by sound, objective evidence. Rather, the conclusion is speculative and thereby amounts to no more than the Examiner's opinion. Thus, this rejection should be reversed.

Assuming, *arguendo*, that the Examiner somehow met the burden of establishing a *prima facie* case of lack of enablement, Appellant believes that any such case has been rebutted by the submission of the multiple Declarations of two experts in the field, Dr. Richard Heuser and Dr. Andrew E. Lorincz (all of record). Such Declarations are set forth in the Evidence Appendix (Items 14-22). The conclusions set forth in the respective Declarations establish an objective fact that is highly material to a determination of enablement. These two highly skilled medical experts read relevant portions, including generic and non-elected species portions, and reached the determination that one skilled in the medical arts, armed with the knowledge in the disclosures, would be enabled to practice the claimed method and to predictably anticipate the results defined therein without need for resorting to undue experimentation. See paragraphs 5-7 of the Third

Supplemental Declaration of Dr. Lorincz and paragraphs 5-7 of the Fourth Supplemental Declaration of Dr. Heuser.

The Examiner attempted to diminish the weight to be accorded to such Declarations by essentially rearguing that more than routine experimentation would be required. Moreover, the Examiner's arguments regarding the Declarations do not appear to be directly related to the claims on appeal. Instead, the Examiner's arguments relate to subject matter contained in cancelled claims. Other than restating her opinion, the Examiner has not met her burden of addressing the probative value of the objective evidence in Appellant's Declarations. By failing to articulate adequate reasons to rebut the Heuser and Lorincz Declarations, the Examiner "failed to consider the totality of the record for the purpose of issuing a final rejection and thus erred as a matter of law." In re Alton, 76 F.3d 1168, 37 USPQ2d 1578 (Fed.Cir. 1996). It is trite law that the Examiner must consider the probative value of such evidence *vis-à-vis* any asserted *prima facie* case. See In re Oetiker, at 1445, 24 USPQ 2d at 1444. In re Keller, 642 F.2d 413, 208 USPQ 871, (CCPA 1981). In the absence of critical analysis, the Examiner appears to be relying solely upon her opinion rather than assessing weight to the objective evidence proffered in the Declarations. The Examiner, not being a skilled person in the medical art, must give weight to these expert opinions rather than substitute her own opinion. See In re Neave, 370 F.2d 961, 152 USPQ 274, (CCPA 1967).

In the Final Rejection, the Examiner has questioned the presence or absence of factual support for the expert opinions. In doing so, the Examiner has erroneously implied that the experts relied upon "some publications" in addition to Appellant's disclosure in reaching their opinions. A concise reading of the Declarations reveals that

the experts relied solely upon the guidance and direction in the application's generic and specific disclosures pertaining to the claims coupled with their skills in the medical art in rendering their conclusions. Appellant, likewise, relies upon such disclosure.

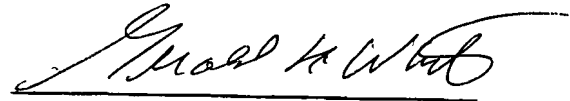
As a final point, the Board's attention is again respectfully directed to the In re Wands decision, which led to the grant of a patent. The Court found that the Patent and Trademark Office's determination of nonenablement was unsupported by the evidence in the record. The Court further noted that the skill level in the art was high and that known materials were utilized in the practice of the invention in weighing the evidence. The instant fact situation is similar to that of In re Wands because the skill level is also high and known administration techniques and known materials are also utilized in the practice of the invention. In addition to such factual parallelism, Appellant provided expert objective evidence in the form of the Declarations of Drs. Heuser and Lorincz. These medical experts read portions of the specification setting forth the generic invention and elected and non-elected species of such generic invention and determined that one skilled in the medical arts, armed with the guidance and direction in the specification disclosures, would be enabled to practice the methods defined in the claims on appeal and to predictably anticipate the results defined therein without need for resorting to undue experimentation. When the guidance and direction provided by Appellant's specification disclosure, the level of knowledge and the content of the prior art at the time of the invention as established in the record and Appellant's declaration evidence are interpreted in a reasonable manner, analysis considering the factors in In re Wands compels a conclusion that undue experimentation would not be required to practice the invention called for in the appealed claims.

CONCLUSION AND RELIEF SOUGHT

In view of the foregoing, Appellant urges the Board to reverse the outstanding rejection of claims 236, 238, 239, 243, 244, 247, 250, 251, 253, 257-263, 268-271, and 280-285 under 35 U.S.C. §112, first paragraph, and respectfully requests that the instant application be passed to issue.

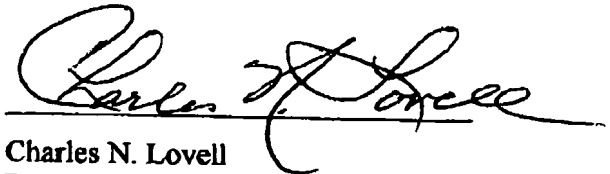
Respectfully submitted,

Dated: 10/23/07



Gerald K. White
Reg. No. 26,611
Attorney for Appellant

Dated: 10/22/07



Charles N. Lovell
Reg. No. 38,012
Attorney for Appellant

GERALD K. WHITE & ASSOCIATES, P.C.
205 W. Randolph Street
Suite 835
Chicago, IL 60606
Phone: (312) 920-0588
Fax: (312) 920-0580
Email: gkwpatlaw@aol.com

CLAIMS APPENDIX

Claims 236, 238, 239, 243, 244, 247, 250, 251, 253, 257-263, 268-271, and 280-285 are pending in the application, are under final rejection, are being appealed, and are listed below.

- | | |
|-----------|---|
| Claim 236 | A method of growing a new portion of a pre-existing heart comprising the steps of placing a growth factor in a body of a human patient and growing new cardiac muscle and growing a new artery in said heart. |
| Claim 238 | The method of claim 236, further comprising repairing a dead portion of said heart. |
| Claim 239 | The method of claim 236, further comprising repairing a damaged portion of said heart. |
| Claim 243 | The method of claim 236, wherein said growth factor comprises a member selected from the group consisting of cells, cellular products, and derivatives of cellular products. |
| Claim 244 | The method of claim 243, wherein said growth factor comprises a cell. |
| Claim 247 | The method of claim 236, wherein said growth factor is placed in said patient by injection. |
| Claim 250 | The method of claim 247, wherein said injection is intramuscular. |
| Claim 251 | The method of claim 236, wherein said growth factor is placed in said patient by a carrier. |

- Claim 253 The method of claim 236, wherein said growth factor comprises a gene and a cell.
- Claim 257 The method of claim 236, wherein said growth factor is locally placed in said body.
- Claim 258 The method of claim 238, wherein said growth factor is locally placed in said body.
- Claim 259 The method of claim 239, wherein said growth factor is locally placed in said body.
- Claim 260 The method of claim 243, wherein said growth factor is locally placed in said body.
- Claim 261 The method of claim 236, wherein said growth factor comprises living stem cells harvested from bone marrow.
- Claim 262 The method of claim 238, wherein said growth factor comprises living stem cells harvested from bone marrow.
- Claim 263 The method of claim 239, wherein said growth factor comprises living stem cells harvested from bone marrow.
- Claim 268 The method of claim 262, wherein said stem cell is placed in said patient by injection.
- Claim 269 The method of claim 263, wherein said stem cell is placed in said patient by injection.
- Claim 270 The method of claim 258, wherein said growth factor comprises a cell and said cell is placed adjacent to said dead portion of said heart.

- Claim 271 The method of claim 259, wherein said growth factor comprises a cell and said cell is placed adjacent to said damaged portion of said heart.
- Claim 280 The method of claim 236 further comprising calculating blood flow through said newly grown artery.
- Claim 281 The method of claim 238 further comprising calculating blood flow through said newly grown artery.
- Claim 282 The method of claim 239 further comprising calculating blood flow through said newly grown artery.
- Claim 283 The method of claim 236 further comprising observing said newly grown artery.
- Claim 284 The method of claim 238 further comprising observing said newly grown artery.
- Claim 285 The method of claim 239 further comprising observing said newly grown artery.

EVIDENCE APPENDIX

1. Office Action dated February 22, 2006 (page 6, lines 1-8) in co-pending application Serial No. 09/794,456 filed February 27, 2001.
2. Strauer et al. publication in Circulation entitled, "Repair of Infarcted Myocardium by Autologous Intracoronary Mononuclear Bone Marrow Cell Transplanattion in Humans" cited by Applicant as Exhibit E in Declaration of Dr. Richard Heuser filed June 17, 2003 (in connection with concurrently-filed Amendment).
3. Deb et al. publication in Circulation entitled, "Bone Marrow-Derived Cardiomyocytes Are Present in Adult Human Heart" cited by Applicant as Exhibit II in the Amendment filed February 17, 2004.
4. Murry et al. 1996 publication in J. Clin. Invest. entitled, "Skeletal Myoblast Transplantation for Repair of Myocardial Necrosis" cited by the Examiner in the November 28, 2003 Office Action.
5. Nabel et al. U.S. Patent No. 5,328,470 cited by the Examiner in the November 28, 2003 Office Action.
6. Strauer et al. 2005 publication in Circulation entitled, "Regeneration of Human Infarcted Heart Muscle by Intracoronary Autologous Bone Marrow Cell Transplantation in Chronic Coronary Artery Disease" cited by Applicant as Exhibit D in the Amendment filed November 21, 2005.
7. Perin et al. 2005 publication in Circulation entitled, "Transendocardial, Autologous Bone Marrow Mononuclear Cell Injection in Ischemic Heart Failure" cited by Applicant as Exhibit E in the Amendment filed November 21, 2005.
8. Trigg 2002 publication in Pediatric Transplantation entitled, "Milestones in the Development of Pediatric Hematopoietic Stem Cell transplantation – 50 Years of Progress" cited by Appellant as Reference AO in the 3rd Supplemental Information Disclosure Statement filed July 30, 2004.
9. Areman et al. 1990 publication in Prog. Clin. Biol. Res., entitled "Automated Isolation of Mononuclear cells using the Fenwal CS3000 blood cell separator" (as found on the website www.ncbi.nlm.nih.gov) cited by Appellant as Exhibit F to Appellant's Appeal Brief filed June 13, 2005.

10. Angelini et al. 1990 publication in Haematologica, entitled, "Human bone marrow processing using Cobe 2991 and CS 3000 blood cell separators for further ex vivo manipulation" (as found on the website www.ncbi.nlm.nih.gov) cited by Appellant as Exhibit G to Appellant's Appeal Brief filed June 13, 2005.
11. Janssen et al. 1992 publication in J Hematother, entitled, "Use of the Terumo SteriCell for the processing of bone marrow and peripheral blood stem cells" (as found on the website www.ncbi.nlm.nih.gov) cited by Appellant as Exhibit H to Appellant's Appeal Brief filed June 13, 2005.
12. Caplan 1991 publication in Journal of Orthopaedic Research, entitled, "Mesenchymal Stem Cells" cited by Applicant as Exhibit E in the Amendment filed June 26, 2006.
13. Wollert 2004 publication in The Lancet entitled, "Intracoronary autologous bone-marrow cell transfer after myocardial infarction: the BOOST randomized controlled clinical trial" cited by Applicant as Reference AP in the 3rd Supplemental Information Disclosure Statement filed July 30, 2004.
14. Declaration of Dr. Richard Heuser filed on June 17, 2003.
15. Supplemental Declaration of Dr. Heuser filed on February 17, 2004.
16. 2nd Supplemental Declaration of Dr. Heuser filed on dated July 30, 2004.
17. 3rd Supplemental Declaration of Dr. Heuser cited by Appellant as Exhibit I in the Appeal Brief filed June 13, 2005.
18. 4th Supplemental Declaration of Dr. Heuser cited by Applicant as Exhibit A in the Amendment dated June 26, 2006.
19. Declaration of Dr. Andrew E. Lorincz filed on June 17, 2003.
20. Supplemental Declaration of Dr. Andrew E. Lorincz filed on February 17, 2004.
21. 2nd Supplemental Declaration of Dr. Andrew E. Lorincz filed on July 30, 2004.
22. 3rd Supplemental Declaration of Dr. Andrew E. Lorincz cited by Applicant as Exhibit B in the Amendment dated June 26, 2006.

RELATED PROCEEDINGS APPENDIX

1. Appeal related to Serial No. 09/794,456, filed February 27, 2001, Appellant's Brief filed February 8, 2007.

EVIDENCE APPENDIX

Item 1



UNITED STATES PATENT AND TRADEMARK OFFICE

UNITED STATES DEPARTMENT OF COMMERCE
United States Patent and Trademark Office
Address: COMMISSIONER FOR PATENTS
P.O. Box 1450
Alexandria, Virginia 22313-1450
www.uspto.gov

APPLICATION NO.	FILING DATE	FIRST NAMED INVENTOR	ATTORNEY DOCKET NO.	CONFIRMATION NO.
09/794,456	02/27/2001	James P. Elia	1000-10-CO	4530

7590 02/22/2006

Gerald K. White
GERALD K. WHITE & ASSOCIATES, P.C.
Suite 835
205 W. Randolph Street
Chicago, IL 60606

EXAMINER

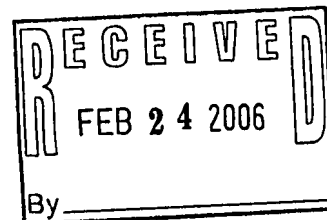
KEMMERER, ELIZABETH

ART UNIT	PAPER NUMBER
----------	--------------

1646

DATE MAILED: 02/22/2006

Please find below and/or attached an Office communication concerning this application or proceeding.



Office Action Summary

Application No.

09/794,456

Applicant(s)

ELIA, JAMES P.

Examiner

Elizabeth C. Kemmerer, Ph.D.

Art Unit

1646

-- The MAILING DATE of this communication appears on the cover sheet with the correspondence address --

Period for Reply

A SHORTENED STATUTORY PERIOD FOR REPLY IS SET TO EXPIRE 3 MONTH(S) OR THIRTY (30) DAYS, WHICHEVER IS LONGER, FROM THE MAILING DATE OF THIS COMMUNICATION.

- Extensions of time may be available under the provisions of 37 CFR 1.136(a). In no event, however, may a reply be timely filed after SIX (6) MONTHS from the mailing date of this communication.
- If NO period for reply is specified above, the maximum statutory period will apply and will expire SIX (6) MONTHS from the mailing date of this communication.
- Failure to reply within the set or extended period for reply will, by statute, cause the application to become ABANDONED (35 U.S.C. § 133). Any reply received by the Office later than three months after the mailing date of this communication, even if timely filed, may reduce any earned patent term adjustment. See 37 CFR 1.704(b).

Status

- 1) ☒ Responsive to communication(s) filed on 09 January 2006.
- 2a) ☐ This action is FINAL. 2b) ☒ This action is non-final.
- 3) ☐ Since this application is in condition for allowance except for formal matters, prosecution as to the merits is closed in accordance with the practice under *Ex parte Quayle*, 1935 C.D. 11, 453 O.G. 213.

Disposition of Claims

- 4) ☒ Claim(s) 7-40 is/are pending in the application.
- 4a) Of the above claim(s) 8-10 and 22-24 is/are withdrawn from consideration.
- 5) ☐ Claim(s) _____ is/are allowed.
- 6) ☒ Claim(s) 7, 11-21 and 25-40 is/are rejected.
- 7) ☐ Claim(s) _____ is/are objected to.
- 8) ☐ Claim(s) _____ are subject to restriction and/or election requirement.

Application Papers

- 9) ☐ The specification is objected to by the Examiner.
- 10) ☐ The drawing(s) filed on _____ is/are: a) ☐ accepted or b) ☐ objected to by the Examiner.
Applicant may not request that any objection to the drawing(s) be held in abeyance. See 37 CFR 1.85(a).
Replacement drawing sheet(s) including the correction is required if the drawing(s) is objected to. See 37 CFR 1.121(d).
- 11) ☐ The oath or declaration is objected to by the Examiner. Note the attached Office Action or form PTO-152.

Priority under 35 U.S.C. § 119

- 12) ☐ Acknowledgment is made of a claim for foreign priority under 35 U.S.C. § 119(a)-(d) or (f).
- a) ☐ All b) ☐ Some * c) ☐ None of:
1. ☐ Certified copies of the priority documents have been received.
 2. ☐ Certified copies of the priority documents have been received in Application No. _____.
 3. ☐ Copies of the certified copies of the priority documents have been received in this National Stage application from the International Bureau (PCT Rule 17.2(a)).
- * See the attached detailed Office action for a list of the certified copies not received.

Attachment(s)

- 1) ☒ Notice of References Cited (PTO-892)
- 2) ☐ Notice of Draftsperson's Patent Drawing Review (PTO-948)
- 3) ☒ Information Disclosure Statement(s) (PTO-1449 or PTO/SB/08)
Paper No(s)/Mail Date 1/9/06.
- 4) ☐ Interview Summary (PTO-413)
Paper No(s)/Mail Date. _____.
- 5) ☐ Notice of Informal Patent Application (PTO-152)
- 6) ☐ Other: _____.

DETAILED ACTION

Status of Application, Amendments, And/Or Claims

A request for continued examination under 37 CFR 1.114 was filed in this application after appeal to the Board of Patent Appeals and Interferences, but prior to a decision on the appeal. Since this application is eligible for continued examination under 37 CFR 1.114 and the fee set forth in 37 CFR 1.17(e) has been timely paid, the appeal has been withdrawn pursuant to 37 CFR 1.114 and prosecution in this application has been reopened pursuant to 37 CFR 1.114. Applicant's submission filed on 21 November 2005 has been entered.

Claims 1-6 are canceled. Claims 10-12 and 22-24 remain withdrawn from consideration as being directed to non-elected invention, for reasons of record. Claims 7, 11-21, and 25-40 are under examination.

The text of those sections of Title 35, U.S. Code not included in this action can be found in a prior Office action.

Withdrawn Objections And/Or Rejections

The rejection of claims 7, 11-15, 18, 19, 21, 25-29, 32, and 33 as unpatentable over Murry et al. is *withdrawn* upon further consideration.

The rejection of claims 20 and 34 as unpatentable over Murry et al. in view of Nabel et al. is *withdrawn* upon further consideration.

35 U.S.C. § 112, First Paragraph – New Matter

Claims 16, 17, 20, 30, 31, and 34 remain rejected under 35 U.S.C. 112, first paragraph, as failing to comply with the written description requirement. The claim(s) contains subject matter which was not described in the specification in such a way as to reasonably convey to one skilled in the relevant art that the inventor(s), at the time the application was filed, had possession of the claimed invention. The basis for this rejection is of record.

Applicant's arguments (pp. 11-13, amendment received 21 November 2005) have been fully considered but are not found to be persuasive for the following reasons.

As an initial matter, it is noted that Applicant incorrectly characterizes the instant rejection as an enablement rejection. The instant rejection is, and always has been, based on the written description component of 35 U.S.C. § 112, first paragraph, and not enablement. See p. 4 of the Office Action mailed 28 November 2003; p. 3 of the Office Action mailed 01 June 2004, and p. 2 of the Advisory Action mailed 26 November 2004. Applicant is reminded that *Vas-Cath Inc. v. Mahurkar*, 19USPQ2d 1111 (CAFC 1991) makes clear that the written description provision of 35 U.S.C. §112 is severable from its enablement provision (see page 1115).

Applicant points to p. 45 of the specification as describing injecting growth factors into a patient intravenously, intraluminally, or intramuscularly to promote growth of an artery, and applying genes or other genetic material with an

Art Unit: 1646

angioplasty balloon. This is not found to be persuasive because page 45, lines 13-16, of the specification reads as follows:

"**VEGF proteins** can be made in a lab and injected into a patient intravenously, intraluminally or intramuscularly to promote the growth of a new artery. Or, the **genes (or other genetic material)** can be applied with an angioplasty balloon, with the assistance of a vector, or by any other method." (emphases added)

Clearly, this section of the specification is limited to use of proteins or nucleic acids (genes or genetic material). Regarding "intravenous" and "intraluminal" delivery, this section of the specification is limited to the suggestion of administering a protein. Nowhere else in the specification is it suggested that cells should be administered intravenously or intraluminally. Regarding angioplasty delivery, the second sentence quoted above is limited to the suggestion of administering genes or other genetic material by angioplasty balloon. The specification defines "growth factors" as comprising cells, but does not define "genetic material" as comprising cells. For example, p. 31, lines 11-13, of the specification states "...the genetic material comprises comparable artificially produced genes, or genes harvested from other human beings or animals." Page 32, lines 8-9 state "genetic material can comprise comparable artificially produced genes or genes removed from another animal or otherwise generated." Page 35, line 4 clearly distinguished between growth factors (defined as encompassing cells) and genetic material: "genetic material plus growth factor(s) are implanted..." Page 35, lines 12-14 states "Genetic material is well conserved in nature. The Drosophila eyeless gene (ey), the mouse small ey gene (pax-6), and the Aniridia gene in humans are all homologous." Page 36,

Art Unit: 1646

lines 25-26 state "Genes control structure and function. A gene or a bit of genetic material may act as a master control gene..." Clearly, the specification uses "genetic material" as pertaining to nucleic acids such as genes. It is also noted that one skilled in the art would only interpret "assistance of a vector," recited in the same sentence that uses "genetic material," as only applying to nucleic acids (genes or RNA or cDNA, etc.).

Applicant points to Capon v. Eshhar v. Dudas, 03-1480-1481 (CAFC 2005) as controlling precedent that 112 does not require recitation in the specification of features already known by workers in the technological field to which the invention is directed. Applicant urges that the examiner mistakenly posited that generic inventions involving biochemical processes require a higher threshold for compliance with 112 because of a perception that success is not assured. Applicant argues that the Court in Capon observed that the USPTO must determine the sufficiency of support on a case-by-case basis given the state of the art at the time of the invention and in light of evidence of record. Applicant argues that the examiner has failed to address the generic concept that Applicant described – the concept of selecting a growth factor (herein the elected subgenus cells) and administering same into the body of a human patient using conventional methods and apparatus to achieve the goals of the claims. This is not found to be persuasive because the instant fact pattern is distinct from that in the case law cited by Applicant. Capon v. Eshhar, 76 USPQ2d 1078 (CAFC 2005) concerns whether or not claims to chimeric DNA molecules are adequately described by a generic description. The issue concerned written description of

Art Unit: 1646

products, not method steps. The issue here is not whether or not workers in this technology already knew the features of the cells recited in the claims; rather, the issue is that the instant specification did not set forth contemplation of a method step wherein cells were administered intravenously, intraluminally, or via angioplasty. As discussed in the previous paragraph, the instant specification did not set forth contemplation of such method steps. The claims are being examined to the extent they read on the elected invention, administration of cells, and thus the generic concept of growth factor is not relevant. Furthermore,

MPEP § 2163.02 reads:

"An Applicant shows possession of the claimed invention by describing the claimed invention with all of its limitations using such descriptive means as words, structures, figures, diagrams, and formulas that fully set forth the claimed invention. See Lockwood v. American Airlines, Inc., 107 F.3d 1565, 1572, 41 USPQ2d 1961, 1966 (Fed. Cir. 1997). Possession may be shown in a variety of ways including description of an actual reduction to practice, or by showing that the invention was "ready for patenting" such as by the disclosure of drawings or structural chemical formulas that show that the invention was complete, or by describing distinguishing identifying characteristics sufficient to show that the applicant was in possession of the claimed invention."

In the instant case, none of these criteria have been met. There was no reduction to practice, and the specification only refers to method steps involving proteins, genes and "genetic material," *but not cells*, as being useful in intravenous, intraluminal and angioplasty delivery. Therefore, the rejection is maintained.

35 U.S.C. § 112, First Paragraph – Enablement

Claims 7, 11-21, and 25-40 are rejected under 35 U.S.C. 112, first paragraph, as failing to comply with the enablement requirement. The claim(s) contains subject matter which was not described in the specification in such a way as to enable one skilled in the art to which it pertains, or with which it is most nearly connected, to make and/or use the invention. The basis of this rejection is set forth at pp. 4-20 AND 29-41 of the Advisory Action mailed 26 November 2004, as well as at pp. 5-17 AND 24-28 of the Final Office Action mailed 01 June 2004.

Applicant's arguments (pp. 12-14, amendment received 21 November 2005) have been fully considered but are not found to be persuasive for the following reasons.

Applicant argues that evidence has been submitted to support enablement of the methods claimed in claims 16, 17, 30, and 31. Applicant points to Strauer 2002 and Strauer 2005 as repotting successful heart repair. Applicant indicates that there can be no "timing issue" that would require more than routine experimentation. This has been fully considered but is not found to be persuasive. Strauer (2002, Circulation 106:913, 1918) disclose balloon catheter injection of bone marrow cells to repair a dead portion of a heart. Claims 16, 17, 30, and 31 pertain to intravenous and intraluminal injection only, and thus Strauer is not particularly relevant. Although balloon catheter administration can be considered a species of the genus of intraluminal delivery methods, it is not commensurate in scope with the claims which read on delivery to the lumen of arteries, veins, intestines, heart chambers, lung, peritoneum, etc. Furthermore,

Art Unit: 1646

Strauer's balloon catheter administration involved infusion of the cells by high-pressure injection directly into the necrotic area, to avoid the "wash-away" effect of standard intraluminal administration (Strauer, p. 1917, third paragraph of left column). It is noted that the specification as originally filed provides no guidance regarding high-pressure injection. Thus the post-filing date publication of Strauer cannot be relied upon to support enablement of the claims, as it uses methods which were not disclosed in the specification as originally filed. Additionally, Strauer specifically points out the shortcomings of intravenous administration of cells for heart therapy at p. 1917, second paragraph of the left column, where they state that "only a very small fraction of infused cells can reach the infarct region," "intravenous application would require many circulation passages to enable infused cells to come into contact with the infarct-related artery," and "homing of cells to other organs could considerably reduce the numbers of cells dedicated to cell repair in the infarcted zone." Thus, Strauer specifically provides evidence of non-enablement of the instant claims reciting intravenous administration of cells. "Strauer 2005" was not attached to the amendment. However, it was cited in a related application, and thus is made of record on the accompanying Notice of References cited. Strauer et al. (2005, J. Am. Coll. Cardiol. 46:1651-1658) used the same procedure as that in Strauer 2002, and thus also does not support enablement of claims 16, 17, 30, and 31 for the same reasons as discussed regarding Strauer 2002.

Regarding claims 7, 11-15, 18-21, 25-29, and 32-34, Applicant argues that the rejection must fail considering the totality of the evidence. Applicant urges

that the specification describes standard systems of identification as well as known procedures for selecting and isolating known cells (bone marrow stem cells) and known apparatus and methods for administering such cells to achieve the desired therapeutic result. Applicant indicates that specification describes specific materials and administration routes. This has been fully considered but is not found to be persuasive. Applicant does not point to any specific section of the specification as supporting these statements. The previous Office Actions have reviewed the specification's teachings and found that they do not provide enablement for the instant claims.

Applicant argues that the examiner has failed to consider the generic concept of selecting well-known appropriate cells and administering such cells using well-known methods and apparatus to grow muscles and arteries in a human patient's heart that do not occur in nature. Applicant urges that the examiner has failed to cite any evidence in the record showing this concept in the prior art. This has been considered but is not found to be persuasive in that it is a confusing argument. Is Applicant admitting that the claimed methods were so well known in the prior art that the specification need not have disclosed anything in addition to the prior art? Also, if the examiner had found evidence that the concept occurred in the prior art, then no enablement rejection would have been made. However, additional prior art rejections under 35 U.S.C. §§ 102 and 103 may have been made. Finally, 35 U.S.C. § 112, first paragraph, requires that the specification set forth what Applicant contemplates as the invention and provide sufficient disclosure so that the skilled artisan can make and use the claimed

invention without undue experimentation. It is improper to pick and choose among unconnected sections of a specification in an attempt to capture another research group's post-filing date discoveries.

Finally, Applicant points to the Perin et al. trials as evidence that nothing more than routine experimentation was required to carry out the technique. Applicant characterizes Perin et al. as following Applicant's basic regimen. This has been fully considered but is not found to be persuasive. The Perin et al. evidence has already been considered. Perin refers to "transendocardial" injections of cells and "intramyocardial" injections of cells, both of which appear to be intramuscular forms of administration. Thus, Perin also cannot be used to support enablement of the rejected claims which recite intravenous or intraluminal administration of cells. Furthermore, the Perin et al. evidence submitted by Applicant is merely an Abstract which does not set forth any experimental details such that it can be ascertained whether or not Perin et al. used the same approach as that set forth in the claims corresponding to the same scope, or whether or not methods or materials other than those contemplated by Applicant were used. In other words, it cannot be ascertained whether or not Perin et al. constitutes evidence commensurate in scope with the claims.

Applicant submitted an article in an IDS dated 09 January 2006 (McDougall, 2005, Men's Health, pp. 164-171 and 194) to support enablement of the claimed invention. The reported information is based on an interview with Dr. Perin regarding his stem cell therapy approach for treating heart disease.

Art Unit: 1646

Applicant quotes from the article that "it's like Miracle-Gro for your heart."

Applicant characterizes the article as describing the procedure of injecting autologous bone marrow stem cells by catheter into the patient's heart as "astonishingly easy." Applicant argues that this submission is consistent with the expert evidence proffered by Drs. Heuser and Lorincz and supports Applicant's arguments that one skilled in the medical arts would have to resort to nothing more than routine experimentation to practice the described and claimed invention. Applicant urges that such submission further evinces the novel/pioneering nature of applicant's invention which uses old materials, old apparatus and old and well known methods to achieve a truly novel and miraculous result. This has been fully considered but is not found to be persuasive. The article submitted by Applicant is from "Men's Health" magazine, which is written for lay people, not those of ordinary skill in the art. Thus, any statements in the article regarding the ease or simplicity of the procedure are misleading. They may appear simple to the patient receiving the therapy; however, this has no bearing on how much research was involved in developing the techniques used. The research referred to in the article was that of Dr. Perin. So far, only a lay person's magazine article and an abstract have been submitted regarding Dr. Perin's trials. Such do not set forth the details relevant to the question of whether or not Dr. Perin's work supports enablement of the instant invention.

In view of the preponderance of the totality of the evidence, the enablement rejection is properly maintained.

35 U.S.C. § 112, Second Paragraph

Claims 13 and 27 remain rejected under 35 U.S.C. 112, second paragraph, as being indefinite for failing to particularly point out and distinctly claim the subject matter which applicant regards as the invention. The basis of this rejection is of record.

Applicant's arguments (pp. 7-11 of the amendment received 21 November 2005) have been fully considered but are not found to be persuasive for the following reasons.

Applicant reviews the recent finding in Phillips v. AWH Corporation (75 USPQ2d 1321) that claims are generally given their ordinary and customary meaning in the art, and that claims should be read in the context of the disclosure. Applicant argues that Phillips states that extrinsic evidence is less significant than the intrinsic record. Applicant points to the finding in Phillips that dictionary evidence can be useful, but such evidence is less reliable than specifications and prosecution histories. Applicant argues that the examiner should interpret the words "multifactorial and non-specific" in light of the specification, giving the words their ordinary meaning. Applicant argues that the examiner's interpretation is based on non-contextual sources places the terms out of context and do not enjoy the same weight of evidence as the specification. This has been fully considered but is not found to be persuasive. The examiner takes no issue with the general principles discussed in Phillips. The specification was the first place consulted by the examiner to breathe life and meaning into the

Art Unit: 1646

term "multifactorial and non-specific" as applied to cells. As explained previously on the record, neither the specification nor the art provides an unambiguous definition for the term. Page 37 of the specification states, "Multifactorial and nonspecific cells (such as stem cells and germinal cells) can provide the necessary in vivo and in vitro cascade of genetic material once an implanted master control gene's transcription has been activated." The use of "such as" clearly implies that the term "multifactorial and non-specific cells" is intended to encompass cells other than stem cells and germinal cells. However, neither the specification nor the art disclose what these other cells are. In the absence of this information, the skilled artisan cannot determine the metes and bounds of the claims at issue. The functional portion of the definition, "...provide[s] the necessary in vivo and in vitro cascade of genetic material ..." makes no sense. What is a cascade of genetic material? Thus, the specification does not define these terms, and the metes and bounds of the claimed invention cannot be determined. A search of the prior art indicated that the relevant art also does not use the terms "multifactorial and non-specific" in connection with cells. See Appendix A, submitted as evidence, regarding a search done in the database Medline. The first result uses "multifactorial" to describe diseases. The second result uses "multifactorial" to describe a process. The third result uses "multifactorial" to describe a process. The fourth result uses "multifactorial" to describe analyses. The fifth result uses "multifactorial" to describe a process. The sixth result uses "multifactorial" to describe a study. Each of these usages is consistent with the examiner's position that the term "multifactorial," given its

ordinary and customary usage in the art, is used to describe causes, effects and processes, not cells.

Applicant argues that the examiner's position is supported by a lack of search results regarding the terms followed by a series of suppositions and speculations regarding the meaning of the terms. Applicant characterizes the examiner's position as amounting to nothing more than opinion due to lack of evidence. Applicant indicates that while a chemist may interpret "multifactorial" to be limited to describing a process, one in the medical arts would not. Applicant urges that the term "factor" is well known in the medical art, and that "multifactorial" would be understood by one in the medical art to mean more than one factor. This has been fully considered but is not found to be persuasive. The rejection is supported by evidence. See discussion of the specification and attached search results. Applicant has provided no evidence that chemists and medical artisans would interpret "multifactorial" in different ways. Finally, Applicant's definition of "multifactorial" as meaning "more than one factor" makes no sense when applied to cells. What is a "more than one factor cell?"

Applicant refers to the fifth supplemental IDS as providing definitions. Applicant argues that the definitions are confirming evidence that the disputed terms are known and used properly in the specification, and that the IDS identifies the terms as adjectives. This has been fully considered but is not found to be persuasive. Regarding the dictionary definitions provided by the fifth supplemental IDS, the dictionary.net's definition of multifactorial is "involving or depending on several factors or causes (especially pertaining to a condition or

Art Unit: 1646

disease resulting from the interaction of many genes)." This supports the rejection in that the term "multifactorial" is not used to describe cells. It is used to describe a cause (for example, of the disease) or an effect (for example, of the genes). Similarly, the dictionary.net's definition of nonspecific is "not caused by a specific agent; used also of staining in making microscope slides; 'nonspecific enteritis'" supports the rejection. "Nonspecific" is not used to describe cells. How can cells be "not caused by a specific agent?" The definition uses the term to describe causes (i.e., nonspecific enteritis is a disease caused by undefined factors). The selection of thesaurus words quoted by Appellant ("...undecided, undetermined, undifferentiated...") is also problematic. Cells can be in various stages of differentiation. For example, an embryonic stem cell would clearly be completely undifferentiated, as it can differentiate into any cell type. However, a promyelocyte is "undifferentiated" to an extent in that it can differentiate into a basophil, eosinophil, or neutrophil, whereas it cannot differentiate into any other cell type (e.g., keratinocytes, neural cells, muscle cells). The instant specification does not clarify whether such intermediate cells are encompassed by the term "multifactorial and non-specific."

Applicant provides definitions from Merriam Webster's Medline Plus

Medical Dictionary, namely:

Factor: (noun) a substance that functions in or promotes the function of a particular physiological process or bodily system.
Multifactorial: (adjective) having, involving, or produced by a variety of elements or causes.

Art Unit: 1646

Applicant argues that "factor" means a substance such as a cell that promotes a particular physiological process, such as growth of an artery. Applicant argues that "multifactorial" is used to denote the quality of a cell when a variety of elements (factors) promote the growth of an artery. This has been fully considered but is not found to be persuasive. Applicant's definitions support the rejection. Applicant equates "factor" with "cell." Thus, substituting "cells" for "factors" in Applicant's second sentence, "multifactorial" is used to denote the quality of a cell when a variety of elements [cells] promote the growth of an artery. This simply makes no sense. Regarding the Merriam Webster's Medline Plus Medical Dictionary definition of multifactorial, what types of cells have, involve, or are produced by a variety of elements or causes?

Applicant argues that the terms were understood by those skilled in the art, pointing to the second supplemental declarations of Drs. Heuser and Lorincz. The second supplemental declarations of Drs. Heuser and Lorincz submitted under 37 CFR 1.132 are insufficient to overcome the rejection of claims 13 and 27 based upon 35 U.S.C. § 112, second paragraph because, although the declarations use the term "multifactorial and non-specific cells," they do not explain what cells are encompassed by the term. See section 7 of each of the Heuser and Lorincz second supplemental declarations. In view of the totality of the evidence of record, which includes the specification, prior art of record, and declarations submitted under 37 CFR 1.132, an unambiguous definition of the term "multifactorial and non-specific cells" has not been provided.

Applicant points to the two Strauer et al. publications. Specifically, Applicant points to p. 1656 of Strauer 2003 as stating that cardiac lesions are multifactorial. This has been fully considered but is not found to be persuasive because it supports the instant rejection. Strauer et al. 2003 was not attached to the response ; however, as it was cited in a related application, it has been cited on a Notice of References Cited and attached to this Office Action. Strauer et al. (2003, Circulation 107:929) uses the term "multifactorial" to describe a disease, not cells.

Applicant points to Strauer 2005 as stating that the regenerative potential of bone marrow derived stem cells may be explained by any of four mechanisms, and that "mechanisms" are further referred to as "factors." Applicant argues that the cells can be described as four-factor cells, i.e., multifactorial. Applicant concludes that the totality of the evidence indicates that the rejection should be withdrawn. Applicant also argues that "non-specific" is synonymous with "non-specialized." This has been fully considered but is not found to be persuasive. Strauer 2005 uses "four mechanisms" to describe "regenerative potential," not the cells *per se*. Even if Strauer 2005 could be tortuously construed as describing bone marrow stem cells as multifactorial, Strauer 2005 only discusses bone marrow stem cells. The specification already indicates that stem cells are exemplary of "multifactorial and non-specific" cells. The issue is what cells other than stem cells and germinal cells can be considered multifactorial and non-specific, given that the art does not apply these terms to cells.

Art Unit: 1646

In view of the preponderance of the totality of the evidence, the rejection is maintained.

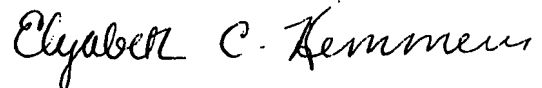
Conclusion

No claims are allowed.

Any inquiry concerning this communication or earlier communications from the examiner should be directed to Elizabeth C. Kemmerer, Ph.D. whose telephone number is (571) 272-0874. The examiner can normally be reached on Monday through Thursday, 7:00 a.m. to 5:30 p.m.

If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Janet Andres, Ph.D. can be reached on (571) 272-0867. The fax phone number for the organization where this application or proceeding is assigned is 571-273-8300.

Information regarding the status of an application may be obtained from the Patent Application Information Retrieval (PAIR) system. Status information for published applications may be obtained from either Private PAIR or Public PAIR. Status information for unpublished applications is available through Private PAIR only. For more information about the PAIR system, see <http://pair-direct.uspto.gov>. Should you have questions on access to the Private PAIR system, contact the Electronic Business Center (EBC) at 866-217-9197 (toll-free).



ECK

ELIZABETH KEMMERER
PRIMARY EXAMINER

APPENDIX

Dialog level 05.10.03D

Logon file001 14feb06 16:17:11

*File 155: Medline has resumed updating.

Set Items Description

? s (multifactorial (2N) cell?) and stem

12971 MULTIFACTORIAL

2761033 CELL?

66 MULTIFACTORIAL(2N)CELL?

143636 STEM

S1 6 (MULTIFACTORIAL (2N) CELL?) AND STEM

? t s1/7/1-6

1/7/1

DIALOG(R)File 155:MEDLINE(R)

(c) format only 2006 Dialog. All rts. reserv.

18357667 PMID: 16020348

Current concepts in ocular surface reconstruction.

Dogru Murat; Tsubota Kazuo

Tokyo Dental College, Chiba, Japan.

Seminars in ophthalmology (United States) Apr-Jun 2005, 20 (2)

p75-93, ISSN 0882-0538 Journal Code: 8610759

Publishing Model Print

Document type: Journal Article; Review; Review, Tutorial

Languages: ENGLISH

Main Citation Owner: NLM

Record type: MEDLINE; Completed

Diseases that affect the limbal %%%stem%%% %%%cells%%% are %%%multifactorial%%% and present with different stages of severity. The most important features to be considered in evaluating these patients include the degree of limbal %%%stem%%% cell loss, the extent of conjunctival disease, and the presence and etiology of ocular surface inflammation. Other important factors are tear film and eyelid abnormalities, keratinization of the ocular surface, laterality of the disease process, health and age of the patient. Careful consideration of all of these factors help tremendously in tailoring the most suitable method of treatment for each patient. The management of severe ocular surface disease has benefited from numerous advances in recent years. At one time, available techniques for visual rehabilitation consisted of superficial keratectomy, use of artificial tears, tarsorrhaphy as well as lamellar and penetrating keratoplasty. A lamellar or penetrating keratoplasty procedure resulted in a stable surface only for as long as the donor epithelium was present and once the epithelium sloughed off, the ocular surface failed due to conjunctivalization. The last few decades enjoyed the development and, especially, progress of new ocular surface reconstruction techniques such as amniotic membrane transplantation, limbal %%%stem%%% cell transplant procedures, transplantation of cultivated oral mucosal or limbal %%%stem%%% cell sheets. This review will briefly focus on the indications and methodology of each procedure and the currently available clinical data on the results of these procedures. (66 Refs.)

Record Date Created: 20050715

Record Date Completed: 20050818

1/7/2

DIALOG(R)File 155:MEDLINE(R)

(c) format only 2006 Dialog. All rts. reserv.

15341984 PMID: 15145210

Robust conversion of marrow cells to skeletal muscle with formation of marrow-derived muscle %%%cell%%% colonies: a %%%multifactorial%%% process.

Abedi Mehrdad; Greer Deborah A; Colvin Gerald A; Demers Delia A; Dooner Mark S; Harpel Jasha A; Weier Heinz-Ulrich; Lambert Jean-Francois; Quesenberry P J

Roger Williams Medical Center, Department of Research, Providence, RI 02864, USA. mabedi@rwmc.org

Experimental hematology (Netherlands) May 2004, 32 (5) p426-34, ISSN 0301-472X Journal Code: 0402313

Contract/Grant No.: 1P22-RR-18757-01; RR; NCRR; P01-DK-5022; DK; NIDDK; P01-HL-56920; HL; NHLBI; R01-DK-2742; DK; NIDDK; R01-DK-49650; DK; NIDDK

Publishing Model Print

Document type: Journal Article

Languages: ENGLISH

Main Citation Owner: NLM

Record type: MEDLINE; Completed

OBJECTIVE: Murine marrow cells are capable of repopulating skeletal muscle fibers. A point of concern has been the "robustness" of such conversions. We have investigated the impact of type of cell delivery, muscle injury, nature of delivered cell, and %%%stem%%% cell mobilizations on marrow-to-muscle conversion. METHODS: We transplanted green fluorescence protein (GFP)-transgenic marrow into irradiated C57BL/6 mice and then injured anterior tibialis muscle by cardiotoxin. One month after injury, sections were analyzed by standard and deconvolutional microscopy for expression of muscle and hematopoietic markers. RESULTS: Irradiation was essential to conversion, although whether by injury or induction of chimerism is not clear. Cardiotoxin- and, to a lesser extent, PBS-injected muscles showed significant number of GFP(+) muscle fibers, while uninjected muscles showed only rare GFP(+) cells. Marrow conversion to muscle was increased by two cycles of G-CSF mobilization and to a lesser extent by G-CSF and steel or GM-CSF. Transplantation of female GFP to male C57BL/6 and GFP to ROSA26 mice showed fusion of donor cells to recipient muscle. High numbers of donor-derived muscle colonies and up to 12% GFP(+) muscle cells were seen after mobilization or direct injection. These levels of donor muscle chimerism approach levels that could be clinically significant in developing strategies for the treatment of muscular dystrophies. CONCLUSION: In summary, the conversion of marrow to skeletal muscle cells is based on cell fusion and is critically dependent on injury. This conversion is also numerically significant and increases with mobilization.

Record Date Created: 20040517

Record Date Completed: 20040624

1/7/3

DIALOG(R)File 155:MEDLINE(R)

(c) format only 2006 Dialog. All rts. reserv.

13036731 PMID: 11000981

Mobilization of peripheral blood progenitor cells for autografting: chemotherapy and G-CSF or GM-CSF.

Siena S; Bregni M; Gianni A M

The Falck Division of Medical Oncology, Ospedale Niguarda, Cai Granda, Milan, Italy.

Bailliere's best practice & research. Clinical haematology (ENGLAND)
Mar-Jun 1999, 12 (1-2) p27-39, ISSN 1521-6926 Journal Code: 100900679
Publishing Model Print

Document type: Journal Article; Review; Review, Tutorial

Languages: ENGLISH

Main Citation Owner: NLM

Record type: MEDLINE; Completed

The mobilization of haematopoietic progenitor %%%cells%%% is a %%%multifactorial%%% process, still poorly understood at the molecular level. Mobilized haematopoietic progenitors, as defined by the expression of CD34 cell surface molecule, comprise heterogeneous subpopulations of cells committed to different haematopoietic lineages. Haematopoietic progenitors may be mobilized by chemotherapy alone, haematopoietic growth factors alone, or by chemotherapy plus haematopoietic growth factors. The choice of a mobilization regimen that allows an optimal yield of progenitors with a minimum number of leukaphereses should incorporate, in most patients, a disease-specific chemotherapeutic agent(s) plus a haematopoietic growth factor, to be continued until completion of harvest.
(76 Refs.)

Record Date Created: 20001019

Record Date Completed: 20001019

1/7/4

DIALOG(R)File 155:MEDLINE(R)

(c) format only 2006 Dialog. All rts. reserv.

12278080 PMID: 9588003

Advances in hematopoietic %%%stem%%% cell culture.

Audet J; Zandstra P W; Eaves C J; Piret J M

Biotechnology Laboratory, University of British Columbia, Vancouver, Canada.

Current opinion in biotechnology (ENGLAND) Apr 1998, 9 (2) p146-51, ISSN 0958-1669 Journal Code: 9100492

Publishing Model Print

Document type: Journal Article; Review; Review, Tutorial

Languages: ENGLISH

Main Citation Owner: NLM

Record type: MEDLINE; Completed

Recent advances in our understanding of the earliest stages of hematopoietic cell differentiation, and how these may be manipulated under defined conditions in vitro, have set the stage for the development of robust bioprocess technology applicable to hematopoietic cells. Sensitive and specific assays now exist for measuring the frequency of hematopoietic %%%stem%%% cells with long-term in vivo repopulating activity from human as well as murine sources. The production of natural or engineered ligands through recombinant DNA and/or combinatorial chemistry strategies is providing new reagents for enhancing the productivity of hematopoietic %%%cell%%% cultures. %%%Multifactorial%%% and dose-response analyses have yielded new insight into the different types and concentrations of factors required to optimize the rate and the extent of amplification of specific subpopulations of primitive hematopoietic cells. In addition, the rate of cytokine depletion from the medium has also been found to be dependent on the types of cell present. The discovery of these cell-type-specific

parameters affecting cytokine concentrations and responses has introduced a new level of complexity into the design of optimized hematopoietic bioprocess systems. (49 Refs.)

Record Date Created: 19980608

Record Date Completed: 19980608

1/7/5

DIALOG(R)File 155:MEDLINE(R)

(c) format only 2006 Dialog. All rts. reserv.

11563022 PMID: 8875201

Role of the basal cells in premalignant changes of the human prostate: a %%%stem%%% cell concept for the development of prostate cancer.

Bonkhoff H

Institute of Pathology, University of the Saarland, Homburg/Saar, Germany.

European urology (SWITZERLAND) 1996, 30 (2) p201-5, ISSN 0302-2838

Journal Code: 7512719

Publishing Model Print

Document type: Journal Article

Languages: ENGLISH

Main Citation Owner: NLM

Record type: MEDLINE; Completed

OBJECTIVES: Prostatic intraepithelial neoplasias (PIN) result from abnormal differentiation and proliferation processes within the prostatic epithelial cell system. Recent data indicate that basal cells are essentially involved in normal and abnormal growth patterns of the human prostate. **RESULTS:** The basal cell layer represents the proliferative compartment and most probably houses the prostatic %%%stem%%% cell population. Basal cells are targets of several regulatory factors including estrogens, androgens, epidermal growth factor and other nonsteroidal growth factors. During the malignant transformation of the prostatic epithelium (PIN), the basal cell layer loses its proliferative function which is transferred to secretory luminal cell types. These proliferative abnormalities are attended by severe regulatory disorders of the programmed cell death within the prostatic epithelial cell system. The Bcl-2 oncoprotein which blocks the programmed cell death in the proliferative compartment (basal cell layer) in normal conditions, extends to the secretory luminal cell types in high-grade PIN lesions. This, in turn, may increase the genetic instability of the dysplastic epithelium. During the process of tumor invasion, the transformed cells lose their basal cell-specific phenotype and acquire features of exocrine cell types which represent the major phenotype in common prostate cancer. At the point of stromal invasion, the transformed cells produce neoplastic basement membrane material which allows them to penetrate the extracellular matrix. **CONCLUSION:** These data provide theoretical bases for a %%%stem%%% cell concept in the development of prostate cancer and highlights the importance of basal %%%cells%%% in this %%%multifactorial%%% process.

Record Date Created: 19970116

Record Date Completed: 19970116

1/7/6

DIALOG(R)File 155:MEDLINE(R)

(c) format only 2006 Dialog. All rts. reserv.

11537242 PMID: 8850230

[T-cell-rich B-cell lymphoma: multifactorial study of 4 cases]

Linfoma B rico en células T: estudio multifactorial de cuatro casos.

Moreno M M; Fernandez-Flores A; Paradelo A; Rodriguez J M; Ageitos A; Gonzalo I; Marcos B; Robledo M; Rivas C

Departamento de Anatomía Patológica, Fundación Jiménez Díaz, Madrid.

Sangre (SPAIN) Dec 1995, 40 (6) p471-7, ISSN 0036-4355

Journal Code: 0404373

Publishing Model Print

Document type: Journal Article ; English Abstract

Languages: SPANISH

Main Citation Owner: NLM

Record type: MEDLINE; Completed

PURPOSE: With the correlational study of four cases in several areas (clinic, morphoimmunological, ultrastructural and genetic) we try to valorate the still controversial entity known as T-cell rich B-cell lymphoma (TRBL), and establish some useful clues in order to settle down the differential diagnosis between TRBL, Hodgkin's disease (HD), and T-cell non-Hodgkin's lymphomas (TNHL). **PATIENTS AND METHODS:** Cases proceeded from Oncology Department, and had been firstly misdiagnosed either as HD (3 cases) or as TNHL (1 case). Biopsies were processed and stained in routine way, H&E, Giemsa and Wilder. Immunohistological study, using monoclonal antibodies against B-cells, T-cells, histiocytes, activation and proliferation markers, was also performed with avidin biotin peroxidase (ABC) method. Ultrastructural study was performed in three of the cases; two patients were studied by PCR and Southern blot. **RESULTS:** All of the cases showed a diffuse histological pattern, with variable fibrosis, and proliferation of venules and capillaries. Small lymphoid cells, being positive for CD3, were dominant. Large blastic cells, positive for CD20, some of them with a Sternberg-like appearance, could be found, in a spotty pattern. Histiocytes were abundant and positive to CD68. Proliferation index (Ki-67) ranged between 13 and 24.5% being the stain mainly positive for B-cells and in a certain extent, also for T-cells. Ultrastructural features were closer to those of the NHL than to the ones found in HD. Molecular study failed to prove any rearrangement. **CONCLUSIONS:** TRBL is a rare entity between B-cell NHL group. Diagnosis and differential diagnosis (mostly with HD and T-cell NHL) have to be properly made, because of the very distinct prognosis and therapy.

Record Date Created: 19961217

Record Date Completed: 19961217

**List of Patents and Publications For
Applicant's 6th Supplemental
Information Disclosure Statement
Page 1 of 1**

Form PTO-1449 (Modified)
(Use several sheets if necessary)



Docket No: 1000-10-CO

Serial No.: 09/794,456

Group No.: 1646

Filing Date: February 27, 2001

Examiner: Elizabeth Kemmerer

Applicant: James P. Elia

Reference Designation U.S. PATENT DOCUMENTS							
Examiner Initial		Document Number	Date	Name	Class	Sub-Class	Filing Date (if appropriate)

Foreign Patent Documents							
		Document Number	Date	Country	Class	Sub-Class	Translation Yes No

Other Art (including author, title, date, pertinent pages, etc.)		
<div style="font-family: cursive; font-size: 1.2em;">EOK</div>	AU	Men's Health, December 2005, McDougall, "The Greatest medical Revolution of the Century is about to begin"

Examiner <div style="font-family: cursive; font-size: 1.2em;">E. Kemmerer</div>	Date Considered <div style="font-family: cursive; font-size: 1.2em;">2/16/06</div>
---	--

Examiner: Initial if reference considered, whether or not citation is in conformance with MPEP 609. Draw line through citation if not in conformance and not considered. Include copy of this form with next communication to applicant.

Notice of References Cited

Application/Control No.

09/794,456

Applicant(s)/Patent Under

Reexamination

ELIA, JAMES P.

Examiner

Elizabeth C. Kemmerer, Ph.D.

Art Unit

1646

Page 1 of 1

U.S. PATENT DOCUMENTS

*		Document Number Country Code-Number-Kind Code	Date MM-YYYY	Name	Classification
	A	US-			
	B	US-			
	C	US-			
	D	US-			
	E	US-			
	F	US-			
	G	US-			
	H	US-			
	I	US-			
	J	US-			
	K	US-			
	L	US-			
	M	US-			

FOREIGN PATENT DOCUMENTS

*		Document Number Country Code-Number-Kind Code	Date MM-YYYY	Country	Name	Classification
	N					
	O					
	P					
	Q					
	R					
	S					
	T					

NON-PATENT DOCUMENTS

*		Include as applicable: Author, Title Date, Publisher, Edition or Volume, Pertinent Pages)
*	U	Strauer et al., 2005, J. Am. Coll. Cardiol. 46:1651-1658.
*	V	Strauer et al., 2003, Circulation 107:929.
	W	
	X	

*A copy of this reference is not being furnished with this Office action. (See MPEP § 707.05(a).)
Dates in MM-YYYY format are publication dates. Classifications may be US or foreign.

EVIDENCE APPENDIX

Item 2

Repair of Infarcted Myocardium by Autologous Intracoronary Mononuclear Bone Marrow Cell Transplantation in Humans

Bodo E. Strauer, MD; Michael Brehm, MD; Tobias Zeus, MD; Matthias Köstering, MD; Anna Hernandez, PhD; Rüdiger V. Sorg, PhD; Gesine Kögler, PhD; Peter Wernet, MD

Background—Experimental data suggest that bone marrow–derived cells may contribute to the healing of myocardial infarction (MI). For this reason, we analyzed 10 patients who were treated by intracoronary transplantation of autologous, mononuclear bone marrow cells (BMCs) in addition to standard therapy after MI.

Methods and Results—After standard therapy for acute MI, 10 patients were transplanted with autologous mononuclear BMCs via a balloon catheter placed into the infarct-related artery during balloon dilatation (percutaneous transluminal coronary angioplasty). Another 10 patients with acute MI were treated by standard therapy alone. After 3 months of follow-up, the infarct region (determined by left ventriculography) had decreased significantly within the cell therapy group (from 30 ± 13 to $12 \pm 7\%$, $P=0.005$) and was also significantly smaller compared with the standard therapy group ($P=0.04$). Likewise, infarction wall movement velocity increased significantly only in the cell therapy group (from 2.0 ± 1.1 to 4.0 ± 2.6 cm/s, $P=0.028$). Further cardiac examinations (dobutamine stress echocardiography, radionuclide ventriculography, and catheterization of the right heart) were performed for the cell therapy group and showed significant improvement in stroke volume index, left ventricular end-systolic volume and contractility (ratio of systolic pressure and end-systolic volume), and myocardial perfusion of the infarct region.

Conclusions—These results demonstrate for the first time that selective intracoronary transplantation of autologous, mononuclear BMCs is safe and seems to be effective under clinical conditions. The marked therapeutic effect may be attributed to BMC-associated myocardial regeneration and neovascularization. (*Circulation*. 2002;106:1913-1918.)

Key Words: myocardial infarction ■ cell transplantation, intracoronary ■ angiogenesis ■ bone marrow ■ myogenesis

Remodeling of the left ventricle after myocardial infarction (MI) represents a major cause of infarct-related heart failure and death. This process depends on acute and chronic transformation of both the necrotic infarct region and the non-necrotic, peri-infarct tissue.^{1,2} Despite application of pharmacotherapeutics and mechanical interventions, the cardiomyocytes lost during MI cannot be regenerated. The recent finding that a small population of cardiac muscle cells is able to replicate itself is encouraging but is still consistent with the concept that such regeneration is restricted to viable myocardium.³

In animal experiments, attempts to replace the necrotic zone by transplanting other cells (eg, fetal cardiomyocytes or skeletal myoblasts) have invariably succeeded in reconstituting heart muscle structures, ie, myocardium and coronary vessels. However, these cells fail to integrate structurally and do not display characteristic physiological functions.⁴⁻⁷ Another approach to reverse myocardial remodeling is to repair myocardial tissue by using bone marrow–derived cells. Bone

marrow contains multipotent adult stem cells that show a high capacity for differentiation.⁸⁻¹⁰ Experimental studies have shown that bone marrow cells (BMCs) are capable of regenerating infarcted myocardium and inducing myogenesis and angiogenesis; this leads in turn to amelioration of cardiac function in mice and pigs.¹¹⁻¹⁴ However, procedures based on this phenomenon remain largely uninvestigated in a human clinical setting.

An investigation of one patient receiving autologous skeletal myoblasts into a postinfarction scar during coronary artery bypass grafting revealed improvement of contraction and viability 5 months afterward.¹⁵ Autologous mononuclear BMCs transplanted in a similar surgical setting showed long-term improvement of myocardial perfusion in 3 of 5 patients and no change in 2 patients.¹⁶ However, such studies entail a surgical approach and are therefore associated with well-known perioperative risks. Moreover, this surgical procedure cannot be used with MI. We therefore looked for a nonsurgical, safer mode for transplanting autologous cells

Received August 2, 2002; accepted August 2, 2002.

From the Department of Medicine, Division of Cardiology (B.E.S., M.B., T.Z., M.K.) and Institute for Transplantation Diagnostics and Cell Therapeutics (A.H., R.V.S., G.K., P.W.), Heinrich-Heine-University of Düsseldorf, Germany.

Correspondence to Professor Dr Bodo E. Strauer, Department of Medicine, Division of Cardiology, Heinrich-Heine-University, Moorenstr 5, 40225 Düsseldorf, Germany. E-mail: Strauer@med.uni-duesseldorf.de

© 2002 American Heart Association, Inc.

Circulation is available at <http://www.circulationaha.org>

DOI: 10.1161/01.CIR.0000034046.87607.1C

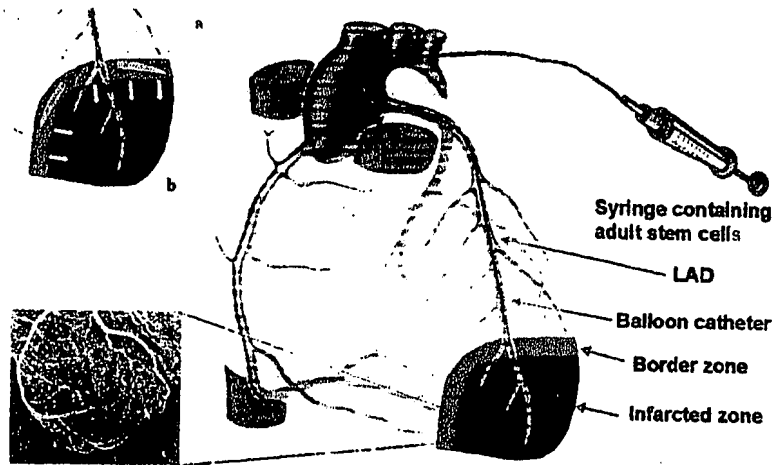


Figure 1. Procedure of cell transplantation into infarcted myocardium in humans. *a*, The balloon catheter enters the infarct-related artery and is placed above the border zone of the infarction. It is then inflated and the cell suspension is infused at high pressure under stop-flow conditions. *b*, In this way, cells are transplanted into the infarcted zone via the infarct-related vasculature (red dots). Cells infiltrate the infarcted zone. Blue and white arrows suggest the possible route of migration. *c*, A supply of blood flow exists within the infarcted zone.³⁸ The cells are therefore able to reach both the border and the infarcted zone.

into postinfarction tissue. A pilot study from our group demonstrated that intracoronary transplantation of autologous mononuclear BMCs 6 days after MI was associated with a marked decrease in infarct area and an increase in left ventricular (LV) function after 3 and 6 months of follow-up.¹⁷ To confirm these results and validate this promising new therapy for MI, we established a clinical trial involving 20 patients for comparing the safety and bioefficacy of autologous BMC transplantation. All 20 patients underwent standard therapy, and 10 patients received additional intracoronary cell transplantation. All 20 patients were followed up for 3 months.

Methods

Patient Population

All 20 patients had suffered transmural infarction according to World Health Organization criteria with the involvement of the left anterior descending coronary artery ($n=4$), left circumflex coronary artery ($n=3$), or right coronary artery ($n=13$). Mean duration of infarct pain was 12 ± 10 hours before invasive diagnostics and therapy. Patients had to be <70 years old and were excluded if one of the following criteria were met: screening >72 hours after infarction, cardiac shock, severe comorbidity, alcohol or drug dependency, or excessive travel distance to the study center.

After right and left heart catheterization, coronary angiography, and left ventriculography, mechanical treatment was initiated with recanalization of the infarct-related artery by balloon angioplasty ($n=20$) and subsequent stent implantation ($n=19$). All patients were monitored in our intensive care unit, and no arrhythmogenic events or hemodynamic impairments were recorded in either patient group.

All 20 patients were briefed in detail about the procedure of BMC transplantation. Informed consent was obtained from 10 patients, who formed the cell therapy group, whereas 10 patients who refused additional cell therapy served as controls. The local ethics committee of the Heinrich-Heine-University, Düsseldorf, approved the study protocol. All procedures conformed to institutional guidelines.

Before taking part in rehabilitation programs, all patients left the hospital with standard medication consisting of acetylsalicylic acid, an ACE inhibitor, a β -blocker, and a statin.

Bone Marrow Aspiration, Isolation, and Cultivation

Seven (± 2) days after acute coronary angiography, bone marrow (~ 40 mL) was aspirated under local anesthesia from ilium of cell therapy patients ($n=10$). Mononuclear BMCs were isolated by Ficoll density separation on Lymphocyte Separation Medium (BioWhittaker) before the erythrocytes were lysed with H_2O . For overnight

cultivation, 1×10^6 BMCs/mL were placed in Teflon bags (Vuelife, Cell Genix) and cultivated in X-Vivo 15 Medium (BioWhittaker) supplemented with 2% heat-inactivated autologous plasma. The next day, BMCs were harvested and washed 3 times with heparinized saline before final resuspension in heparinized saline. Viability was $93 \pm 3\%$. Heparinization and filtration (cell strainer, FALCON) was carried out to prevent cell clotting and microembolization during intracoronary transplantation. The mean number of mononuclear cells harvested after overnight culture was 2.8×10^7 ; this consisted of $0.65 \pm 0.4\%$ AC133-positive cells and $2.1 \pm 0.28\%$ CD34-positive cells. All microbiological tests of the clinically used cell preparations proved negative. As a viability and quality ex vivo control, 1×10^5 cells grown in H5100 medium (Stem Cell Technology) were found to be able to generate mesenchymal cells in culture.

Intracoronary Transplantation of BMCs

Five to nine days after onset of acute infarction, cells were directly transplanted into the infarcted zone (Figure 1). This was accomplished with the use of a balloon catheter, which was placed within the infarct-related artery. After exact positioning of the balloon at the site of the former infarct-vessel occlusion, percutaneous transluminal coronary angioplasty (PTCA) was performed 6 to 7 times for 2 to 4 minutes each. During this time, intracoronary cell transplantation via the balloon catheter was performed, using 6 to 7 fractional high-pressure infusions of 2 to 3 mL cell suspension, each of which contained 1.5 to 4×10^6 mononuclear cells. PTCA thoroughly prevented the backflow of cells and at the same time produced a stop-flow beyond the site of the balloon inflation to facilitate high-pressure infusion of cells into the infarcted zone. Thus, prolonged contact time for cellular migration was allowed.¹⁸

Functional Assessment of Hemodynamics

After 3 months, all 20 patients were followed up by left heart catheterization, left ventriculography, and coronary angiography. Ejection fraction, infarct region, and regional wall movement of the infarcted zone during ejection were determined by left ventriculography. Ejection fraction was measured with Quantcor software (Siemens). To quantify infarction wall movement velocity, 5 axes were placed perpendicular to the long axis in the main akinetic or dyskinetic segment of the ventricular wall. Relative systolic and diastolic lengths were measured, and the mean difference was divided by the systolic duration (in seconds). To quantify the infarct region, the centerline method according to Sheehan was used.¹⁹ All hemodynamic investigations were obtained by two independent observers.

In the cell therapy group before and 3 months after cell transplantation, additional examinations for measuring hemodynamics and myocardial perfusion included dobutamine stress echocardiography, radionuclide ventriculography, catheterization of the right heart, and

TABLE 1. Baseline Characteristics of the Patients

Clinical Data	Cell Therapy	Standard Therapy	P
Characteristics			
No. of patients	10	10	...
Age, y	49±10	50±6	NS
Sex	Male	Male	...
Onset of infarction before angioplasty, h	10±8	13±11	NS
Coronary angiography			
No. of diseased vessels	1.7±0.9	2.1±0.7	NS
No. of patients with LAD/LCX/RCA as the affected vessel	4/1/5	0/2/8	...
No. of patients with stent implantation	9	10	...
Laboratory parameters			
Creatinine kinase, U/L	1138±1170	1308±1187	NS
Creatinine kinase-MB, U/L	106±72	124±92	NS
Bone marrow puncture after angioplasty, d	7±2
Mononuclear bone marrow cells, n (×10 ⁷)	2.8±2.2

Values are mean±SD or number of patients. NS indicates not significant; LAD, left anterior descending coronary artery; LCX, left circumflex coronary artery; and RCA, right coronary artery.

stress-redistribution-reinjection ²⁰¹thallium scintigraphy. The contractility index P_{my}/ESV was calculated by dividing LV systolic pressure (P_{my}) by end-systolic volume (ESV). Perfusion defect was calculated by scintigraphic bull's-eye technique. Each examination was performed according to standard protocols.

There were no complications or side effects determined in any patient throughout the diagnostic or therapeutic procedure or within the 3-month follow-up period.

Statistical Analysis

All data are presented as mean±SD. Statistical significance was accepted when P was <0.05. Discrete variables were compared as rates, and comparisons were made by χ^2 analysis. Intra-individual comparison of baseline versus follow-up continuous variables was performed with a paired t test. Comparison of nonparametric data between the two groups was performed with Wilcoxon test and Mann-Whitney test. Statistical analysis was performed with SPSS for Windows (version 10.1).

Results

Clinical data between the two groups did not differ significantly. The range of creatinine kinase levels was slightly but not significantly higher in the standard therapy group than it was in the cell therapy group (Table 1).

Comparison of the 2 groups 3 months after cell or standard therapy showed several significant differences in LV dynamics, according to the global and regional analysis of left ventriculogram. The infarct region as a percentage of hypokinetic, akinetic, or dyskinetic segments of the circumference of the left ventricle decreased significantly in the cell therapy group (from 30±13 to 12±7%, $P=0.005$). It was also significantly smaller compared with the standard therapy group after 3 months ($P=0.04$). Within the standard therapy group, only a statistically nonsignificant decrease from 25±8 to 20±11% could be seen. Wall movement velocity over the infarct region rose significantly in the cell therapy group (from 2.0±1.1 to 4.0±2.6 cm/s, $P=0.028$) but not in the standard therapy group (from 1.8±1.3 to 2.3±1.6 cm/s, $P=NS$). No significant difference was observed between the

two groups. Ejection fraction increased in both groups, albeit nonsignificantly (from 57±8 to 62±10% in the cell therapy group and from 60±7 to 64±7% in the standard therapy group) (Table 2).

Further significant improvement could also be seen on additional analysis of the cell therapy group alone. Perfusion defect was considerably decreased by 26% in the cell therapy group (from 174±99 to 128±71 cm², $P=0.016$, assessed by ²⁰¹thallium scintigraphy) (Figure 2). Parallel to the reduction in perfusion defect, improvement (Table 3) could also be seen in:

- (1) Cardiac function, as revealed by increase in stroke volume index (from 49±7 to 56±7 mL/m², $P=0.010$) and ejection fraction (from 51±14 to 53±13%, $P=NS$).
- (2) Cardiac geometry, as shown by decreases in both end-diastolic (from 158±20 to 143±30 mL, $P=NS$) and end-systolic volume (from 82±26 to 67±21 mL, $P=0.011$). Radionuclide ventriculography was used to acquire the data.
- (3) Contractility as evaluated by an increase in the velocity of circumferential fiber shortening (from 20.5±4.2 to 24.4±7.7 mm/s, $P=NS$, assessed by stress echocardiography) and by a marked increase in the ratio of systolic pressure to end-systolic volume (from 1.81±1.44 to 2.27±1.72 mm Hg/mL, $P=0.005$).

Discussion

The present report describes the first clinical trial of intracoronary, autologous, mononuclear BMC transplantation for improving heart function and myocardial perfusion in patients after acute MI. The results demonstrate that transplanted autologous BMCs may lead to repair of infarcted tissue when applied during the immediate postinfarction period. These results also show that the intracoronary approach of BMC transplantation seems to represent a novel

TABLE 2. Comparison of Cell Therapy and Standard Therapy Groups

	Cell Therapy	Standard Therapy	P
No. of patients	10	10	...
Infarct region as functional defect			
Hypokinetic, akinetic, or dyskinetic region at 0 mo, %	30±13	25±8	NS
Hypokinetic, akinetic, or dyskinetic region at 3 mo, %	12±7	20±11	0.04
P	0.005	NS	...
Contractility indices			
Infarction wall movement velocity at 0 mo, cm/s	2.0±1.1	1.8±1.3	NS
Infarction wall movement velocity at 3 mo, cm/s	4.0±2.6	2.3±1.6	NS
P	0.028	NS	...
Hemodynamic data			
LV ejection fraction at 0 mo, %	57±8	60±7	NS
LV ejection fraction at 3 mo, %	62±10	64±7	NS
P	NS	NS	...

NS indicates not significant; 0 mo, zero months, which means the time of infarction; 3 mo, 3 months, which means the time of the follow-up examinations. All data were obtained according to analysis of left ventriculogram.

and effective therapeutic procedure for concentrating and/or depositing infused cells within the region of interest.

Neogenesis of both cardiomyocytes and coronary capillaries with some functional improvement has been shown recently by several investigators using bone marrow-derived cells in experimental infarction.^{11–14,18,20–23} Moreover, trans-endothelial migration from the coronary capillaries and incorporation of cells into heart muscle has been observed experimentally.^{3,12,24–26} Until now, clinical data only existed for the cell therapy of surgically treated chronic ischemic heart disease.^{15,16} Our aim was to transform the encouraging results from animal models to a safe clinical setting. The most crucial questions we had to address while designing and

realizing this trial were: (1) What cell population should we deliver? (2) Which application method is the most efficient? (3) When should the cells be transplanted?

In recent years, several laboratories have shown that environmentally dictated changes of fate (transdetermination) are not restricted to stem cells but may also involve progenitor cells at different steps of a given differentiation pathway (transdifferentiation). Moreover, mesenchymal stem cells may represent an ideal cell source for treating different diseases.²⁷ Adult, mononuclear BMCs contain such stem and progenitor cells (≤1%), eg, mesodermal progenitor cells, hematopoietic progenitor cells, and endothelial progenitor cells. In several animal infarction models it has been shown that: (1) Bone marrow hemangioblasts contribute to the formation of new vessels; (2) bone marrow hematopoietic stem cells differentiate into cardiomyocytes, endothelium,

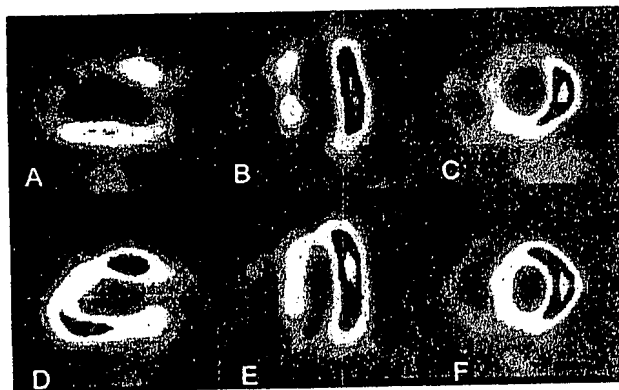


Figure 2. Improved myocardial perfusion of infarcted anterior wall 3 months after intracoronary cell transplantation subsequent to an acute anterior wall infarction detected by ²⁰¹thallium scintigraphy. The images on the left (A, D, sagittal) and in the middle (B, E) show the long axis, whereas those on the right (C, F, frontal) show the short axis of the heart. Initially the anterior wall, with green-colored apical and anterior regions, had reduced myocardial perfusion (A, B, C). Three months after cell transplantation the same anterior wall, now yellow in color, revealed a significant improvement in myocardial perfusion (D, E, F). All illustrations depict the exercise phase.

TABLE 3. Cardiac Function Analysis at 3-Month Follow-Up

	Before Cell Therapy	3 Months After Cell Therapy	P
No. of patients	10	10	...
Hemodynamic data			
LV ejection fraction, %	51±14	53±13	NS
Stroke volume index, mL/m ²	49±7	56±7	0.010
Cardiac geometry			
LV end-diastolic volume, mL	158±20	143±30	NS
LV end-systolic volume, mL	82±26	67±21	0.011
Contractility indices			
Circumferential fiber shortening, mm/s	20.5±4.2	24.4±7.7	NS
P _{sys} /ESV, mm Hg/mL	1.81±1.44	2.27±1.72	0.005
Infarct region as perfusion defect			
²⁰¹ Thallium scintigraphy, cm ²	174±99	128±71	0.016

NS indicates not significant.

and smooth muscle cells⁹⁻¹³; (3) BMCs give rise to mesodermal progenitor cells that differentiate to endothelial cells²⁸; and (4) endothelial progenitors can transdifferentiate into beating cardiomyocytes.²⁹ Thus, several different fractions of mononuclear BMCs may contribute to the regeneration of necrotic myocardium and vessels. In order to utilize this large and perhaps heterogeneous regenerative potential, we decided to use all mononuclear cells from the bone marrow aspirate as a whole, rather than a subpopulation. No further expansion was performed because experimental data have revealed a dramatic decline in the homing capacity of *in vitro* amplified hematopoietic stem or progenitor cells.³⁰

The second question was how to deliver the cells most efficiently. When given intravenously, only a very small fraction of infused cells can reach the infarct region after the following injection: assuming a normal coronary blood flow of 80 mL/min per 100 g of LV weight, a quantity of 160 mL per left ventricle (assuming a regular LV mass of ≈ 200 g) will flow per minute.^{31,32} This corresponds to only about 3% of cardiac output (assuming a cardiac output of 5000 mL/min).³¹ Therefore, intravenous application would require many circulation passages to enable infused cells to come into contact with the infarct-related artery. Throughout this long circulation and recirculation time, homing of cells to other organs could considerably reduce the numbers of cells dedicated to cell repair in the infarcted zone. Thus, supplying the entire complement of cells by intracoronary administration obviously seems to be advantageous for the tissue repair of infarcted heart muscle and may also be superior to intraventricular injection,³³ because all cells are able to flow through the infarcted and peri-infarcted tissue during the immediate first passage. Accordingly, by this intracoronary procedure the infarct tissue and the peri-infarct zone can be enriched with the maximum available amount of cells at all times.

As stem cells differentiate into more mature types of progenitor cells, it is thought that a special microenvironment in so-called niches regulates cell activity by providing specific combinations of cytokines and by establishing direct cellular contact. For successful long-term engraftment, at least some stem cells have to reach their niches, a process referred to as homing. Mouse experiments have shown that significant numbers of BMCs appear in liver, spleen, and bone marrow after intravenous injection.³⁴ To offer the BMCs the best chance of finding their niche within the myocardium, a selective intracoronary delivery route was chosen. Presumably, therefore, fewer cells were lost by extraction toward organs of secondary interest by this first pass-like effect. To facilitate transendothelial passage and migration into the infarcted zone, cells were infused by high-pressure injection directly into the necrotic area, and the balloon was kept inflated for 2 to 3 minutes; the cells were not washed away immediately under these conditions.

The time point for delivery was chosen as 7 to 8 days after infarction onset for the following reasons:

- (1) In dogs, infarcted territory becomes rich in capillaries and contains enlarged, pericyte-poor "mother vessels" and endothelial bridges 7 days after myocardial ischemia and reperfusion. Twenty-eight days later, a significant muscular vessel wall has already formed.³⁵ Thus, with such timing, cells may be able to reach the worst

damaged parts and at the same time salvage tissue. Transendothelial cell migration may also be enhanced because an adequate muscular coat is not yet formed.

- (2) Until now, only one animal study has attempted to determine the optimum time for cardiomyocyte transplantation to maximize myocardial function after LV injury. Adult rat hearts were cryoinjured and fetal rat cardiomyocytes were transplanted immediately, 2 weeks later, and 4 weeks later. The authors discussed the inflammatory process, which is strongest in the first days after infarction, as being responsible for the negative results after immediate cell transplantation, and they assumed that the best results seen after 2 weeks may have been due to transplantation before scar expansion.³⁶ Until now, however, no systematic experiments have been performed with BMCs to correlate the results of transplantation with the length of such a time delay.
- (3) Another important variable is the inflammatory response in MI, which seems to be a superbly orchestrated interaction of cells, cytokines, growth factors and extracellular matrix proteins mediating myocardial repair. In the first 48 hours, debridement and formation of a fibrin-based provisional matrix predominates before a healing phase ensues.³⁷⁻⁴⁰ Moreover, vascular endothelial growth factor is at its peak concentration 7 days after MI, and the decline of adhesion molecules (intercellular adhesion molecules, vascular cell adhesion molecules) does not take place before days 3 to 4 after MI. We assumed that transplantation of mononuclear BMCs within the "hot" phase of post-MI inflammation might lead them to take part in the inflammation cascade rather than the formation of functional myocardium and vessels.

Taking all of this into account, we can conclude that cell transplantation within the first 5 days after acute infarction is not possible for logistical reasons and is not advisable because of the inflammatory process. On the other hand, transplantation 2 weeks after infarction scar formation seems to reduce the benefit of cell transplantation. Although the ideal time point for transplantation remains to be defined, it is most likely between days 7 and 14 after the onset of MI, as in the present study.

This trial was designed as a phase I safety and feasibility trial, meaning that no control group is necessarily required. However, to validate the results, we correlated them with those obtained from 10 patients who refused to get additional cell therapy and thus received standard therapy alone. We are aware of the fact that such a comparison does not reach the power of a randomly allocated, blinded control group. However, the significant improvement with regard to infarct region, hemodynamics (stroke volume index), cardiac geometry (LV end-systolic volume), and contractility ($P_{1/2}/ESV$ and infarction wall movement velocity) did confirm a positive effect of the additional cell therapy because the changes observed in the standard therapy group failed to reach significance.

Another important factor for interpreting the results is time interval between onset of symptoms and revascularization of the infarct-related artery by angioplasty; this represents a crucial determinant of LV recovery. For patients with acute MI, it has

been shown that if the time interval is >4 hours, no significant changes in ejection fraction, regional wall motion, or ESV are observed after 6-month follow-up by echocardiography and angiography.⁴¹ None of our 20 patients was treated by angioplasty within 4 hours after onset of symptoms. Our average time interval was 12 ± 10 hours. Thus, PTCA-induced improvement of LV function can be nearly excluded; indeed, the only mild and nonsignificant changes within the standard therapy group are consistent with the above-mentioned data.⁴¹ In contrast, the cell therapy group showed considerable and significant improvement in the same parameters, which may be attributed to BMC-mediated coronary angiogenesis and cardiomyoneogenesis.

These results show that transplantation of autologous BMCs, as well as the intracoronary approach, represent a novel and effective therapeutic procedure for the repair of infarcted myocardium. For this method of therapy, no ethical problems exist, and no side effects were observed at any point of time. The therapeutic benefit for the patient's heart seems to prevail. However, further experimental studies, controlled prospective clinical trials, and variations of cell preparations are required to define the role of this new approach for the therapy of acute MI in humans.

References

- Pfeffer MA, Braunwald E. Ventricular remodeling after myocardial infarction: experimental observations and clinical implications. *Circulation*. 1990;81:1161-1172.
- Ertl G, Gaudron P, Hu K. Ventricular remodeling after myocardial infarction: experimental and clinical studies. *Basic Res Cardiol*. 1993;88:125-137.
- Quaini F, Urbanek K, Beltrami AP, et al. Chimerism of the transplanted heart. *N Engl J Med*. 2002;346:5-15.
- Leor J, Patterson M, Quinones MJ, et al. Transplantation of fetal myocardial tissue into infarcted myocardium of rat: a potential method for repair of infarcted myocardium? *Circulation*. 1996;94(suppl II):332-336.
- Murry CE, Wiseman RW, Schwartz SM, et al. Skeletal myoblast transplantation for repair of myocardial necrosis. *J Clin Invest*. 1996;98:2512-2523.
- Taylor DA, Atkins BZ, Hungspreugs P, et al. Regenerating functional myocardium: improved performance after skeletal myoblast transplantation. *Nat Med*. 1998;4:929-933.
- Tomita S, Li RK, Weisel RD, et al. Autologous transplantation of bone marrow cells improves damaged heart function. *Circulation*. 1999;100(suppl II):247-256.
- Blau HM, Brazelton TR, Weimann JM. The evolving concept of a stem cell: entity or function? *Cell*. 2001;105:829-841.
- Krause DS, Theise ND, Collector MI, et al. Multi-organ, multi-lineage engraftment by a single bone marrow-derived stem cell. *Cell*. 2001;105:369-377.
- Goodell MA, Jackson KA, Majka SM, et al. Stem cell plasticity in muscle and bone marrow. *Ann N Y Acad Sci*. 2001;938:208-218.
- Orlic D, Kajstura J, Chimenti S, et al. Bone marrow cells regenerate infarcted myocardium. *Nature*. 2001;410:701-705.
- Orlic D, Kajstura J, Chimenti S, et al. Mobilized bone marrow cells repair the infarcted heart, improving function and survival. *Proc Natl Acad Sci U S A*. 2001;98:10344-10349.
- Kocher AA, Schuster MD, Szabolcs MJ, et al. Neovascularization of ischemic myocardium by human bone-marrow-derived angioblasts prevents cardiomyocyte apoptosis, reduces remodeling and improves cardiac function. *Nat Med*. 2001;7:430-436.
- Tomita S, Mickle DA, Weisel RD, et al. Improved heart function with myogenesis and angiogenesis after autologous porcine bone marrow stromal cell transplantation. *J Thorac Cardiovasc Surg*. 2002;123:1132-1135.
- Menasche P, Hagege AA, Scorsin M, et al. Myoblast transplantation for heart failure. *Lancet*. 2001;357:279-280.
- Hamano K, Nishida M, Hirata K, et al. Local implantation of autologous bone marrow cells for therapeutic angiogenesis in patients with ischemic heart disease: clinical trial and preliminary results. *Jpn Circ J*. 2001;65:845-847.
- Strauer BE, Brehm M, Zeus T, et al. Myocardial regeneration after intracoronary transplantation of human autologous stem cells following acute myocardial infarction. *Disch med Wschr*. 2001;126:932-938.
- Wang JS, Shum-Tim D, Chedrawy E, et al. The coronary delivery of marrow stromal cells for myocardial regeneration: pathophysiologic and therapeutic implications. *J Thorac Cardiovasc Surg*. 2001;122:699-705.
- Sheehan FH, Bolson EL, Dodge HT, et al. Advantages and applications of the Centerline method for characterizing regional ventricular function. *Circulation*. 1986;74:293-305.
- Sussman M. Cardiovascular biology: hearts and bones. *Nature*. 2001;410:640-641.
- Toma C, Pittenger MF, Cahill KS, et al. Human mesenchymal stem cells differentiate to a cardiomyocyte phenotype in the adult murine heart. *Circulation*. 2002;105:93-98.
- Kamihata H, Matsubara H, Nishiue T, et al. Implantation of bone marrow mononuclear cells into ischemic myocardium enhances collateral perfusion and regional function via side supply of angioblasts, angiogenic ligands, and cytokines. *Circulation*. 2001;104:1046-1052.
- Ferrari G, Cusella-De Angelis G, Coletta M, et al. Muscle regeneration by bone marrow-derived myogenic progenitors. *Science*. 1998;279:1528-1530.
- Kawamoto A, Gwon HC, Iwaguro H, et al. Therapeutic potential of ex vivo expanded endothelial progenitor cells for myocardial ischemia. *Circulation*. 2001;103:634-637.
- Robinson SW, Cho PW, Levitsky HI, et al. Arterial delivery of genetically labelled skeletal myoblasts to the murine heart: long-term survival and phenotypic modification of implanted myoblasts. *Cell Transplant*. 1996;5:77-91.
- Bitner RE, Schofer C, Weipoltshammer K, et al. Recruitment of bone-marrow-derived cells by skeletal and cardiac muscle in adult dystrophic mdx mice. *Anat Embryol (Berl)*. 1999;199:391-396.
- Jiang Y, Jahagirdar B, Reinhardt RL, et al. Pluripotency of mesenchymal stem cells derived from adult marrow. *Nature*. 2002;20:1-12.
- Reyes M, Lund T, Lenvik T, et al. Purification and ex vivo expansion of postnatal human marrow mesodermal progenitor cells. *Blood*. 2001;98:2615-2625.
- Condorelli G, Borello U, De Angelis L, et al. Cardiomyocytes induce endothelial cells to trans-differentiate into cardiac muscle: implications for myocardium regeneration. *Proc Natl Acad Sci U S A*. 2001;98:10733-10738.
- Szilvassy SJ, Bass MJ, Van Zant G, et al. Organ-selective homing defines engraftment kinetics of murine hematopoietic stem cells and is compromised by ex vivo expansion. *Blood*. 1999;93:1557-1566.
- Gregg DE, Fisher LC. Blood supply to the heart. In: *Handbook of Physiology*. Vol 2. Washington, DC: American Physiological Society; 1963:chap 44, 1517-1584.
- Strauer BE. Myocardial oxygen consumption in chronic heart disease: role of wall stress, hypertrophy and coronary reserve. *Am J Cardiol*. 1979;43:730-740.
- Toma C, Pittenger MF, Byrne BJ, et al. Adult human mesenchymal stem cells differentiate to a striated muscle phenotype following arterial delivery to the murine heart. *Circulation*. 2000;102(suppl II):II-683. Abstract.
- Hendrikx PJ, Martens CM, Hagenbeek A, et al. Homing of fluorescently labeled murine hematopoietic stem cells. *Exp Hematol*. 1996;24:129-140.
- Ren G, Michael LH, Entman ML, et al. Morphological characteristics of the microvasculature in healing myocardial infarcts. *J Histochem Cytochem*. 2002;50:71-79.
- Li RK, Mickle DA, Weisel RD, et al. Optimal time for cardiomyocyte transplantation to maximize myocardial function after left ventricular injury. *Ann Thorac Surg*. 2001;72:1957-1963.
- Frangogiannis NG, Smith CW, Entman ML. The inflammatory response in myocardial infarction. *Cardiovasc Res*. 2002;53:31-47.
- Allgöwer M. *The Cellular Basis of Wound Repair*. Springfield, Ill: Charles C. Thomas; 1956.
- Xie Y, Zhou T, Shen W, et al. Soluble cell adhesion molecules in patients with acute coronary syndrome. *Chin Med J*. 2000;113:286-288.
- Soeki T, Tamura Y, Shinohara H, et al. Serial changes in serum VEGF and HGF in patients with acute myocardial infarction. *Cardiology*. 2000;93:168-174.
- Sheiban I, Frangasso G, Rosano GMC, et al. Time course and determinants of left ventricular function recovery after primary angioplasty in patients with acute myocardial infarction. *J Am Coll Cardiol*. 2001;38:464-471.

EVIDENCE APPENDIX

Item 3

SEARCH

Feedback | Subscriptions | Archives | Search | Table of Contents

Circulation

Access via PAY PER ARTICLE

Published online before print February 24, 2003, doi:10.1161/01.CIR.0000061910.39145.F0
(Circulation. 2003;107:1247.)

© 2003 American Heart Association, Inc.

Brief Rapid Communications

Bone Marrow-Derived Cardiomyocytes Are Present in Adult Human Heart

A Study of Gender-Mismatched Bone Marrow Transplantation Patients

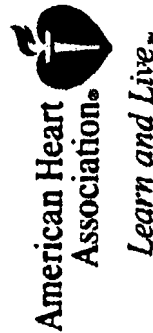
Arjun Deb, MD; Shaohua Wang, MD; Kimberly A. Skelding, MD; Dylan Miller, MD; David Simper, MD; Noel M. Caplice, MD, PhD

From the Division of Cardiovascular Diseases and Molecular Medicine Program, Mayo Clinic, Rochester, Minn.

Correspondence to Noel M. Caplice, MD, PhD, Mayo Clinic, GU 1801, 200 First St SW, Rochester, MN 55905.
E-mail caplice.noel@mayo.edu

► Abstract

Background—Recent studies have identified cardiomyocytes of extracardiac origin in transplanted human hearts, but the exact origin of these myocyte progenitors is currently unknown.



► [Abstract of this Article \(FREE\)](#)

► [Reprint \(PDF\) Version of this](#)

► [Citation Map](#)

► [Email this article to a friend](#)

► [Similar articles found in:](#)

► [Circulation Online](#)

► [PubMed](#)

► [PubMed Citation](#)

► [This Article has been cited by:](#)
other online articles

► [Search Medline for articles by:](#)

► [Deb, A. || Caplice, N. M.](#)

► [Alert me when:](#)

new articles cite this article

► [Download to Citation Manager](#)

► [Collections under which this article appears:](#)

► [Other myocardial biology](#)

► [Myogenesis](#)

► [All Versions of this Article:](#)

107/9/1247 (most recent)

01.CIR.0000061910.39145.F0v1

► [Top](#)
• [Abstract](#)
▼ [Introduction](#)

Methods and Results—Hearts of female subjects (n=4) who had undergone sex-mismatched bone marrow transplantation (BMT) were recovered at autopsy and analyzed for the presence of Y chromosome-positive cardiomyocytes. Four female gender-matched BMT subjects served as controls. Fluorescence in situ hybridization (FISH) for the Y chromosome was performed on paraffin-embedded sections to identify cells of bone marrow origin with concomitant immunofluorescent labeling for -sarcomeric actin to identify cardiomyocytes. A total of 160 000 cardiomyocyte nuclei were analyzed approximating 20 000 nuclei per patient. The mean percentage of Y chromosome-positive cardiomyocytes in patients with sex-mismatched BMT was 0.23±0.06%. Not a single Y chromosome-positive cardiomyocyte was identified in any of the control patients. Immunofluorescent staining for laminin and chromosomal ploidy analysis with FISH showed no evidence of either pseudonuclei or cell fusion in any of the chimeric cardiac myocytes identified.

Conclusions—These data establish for the first time human bone marrow as a source of extracardiac progenitor cells capable of de novo cardiomyocyte formation.

Key Words: chimera • stem cells • myocytes, cardiac • transplantation, bone marrow

Introduction

The concept of the human heart as an organ incapable of self-renewal has recently been challenged by identification of cardiac myocytes of probable extracardiac origin in hearts of patients undergoing sex-mismatched cardiac transplantation.¹⁻⁴ The exact source of these cells is currently unclear, but data from experiments in animals support a bone marrow origin.⁵ It is important to note that a marked discrepancy in the level of cardiac chimerism has been observed in the gender-mismatched cardiac transplantation setting.^{1-4 6,7} Moreover, controversy has arisen with regard to the methodologies used to define chimeric cardiac myocytes in these human studies. Specifically, concerns have recently been raised about the most appropriate techniques required to differentiate (1) true cardiac myocyte nuclei from pseudonuclei,⁶ and (2) diploid nuclei from epigenetic phenomena, such as spontaneous cell fusion.⁸

To address the above issues, we used a specific study design and experimental approach. An ideal method to answer the question of bone marrow origin of chimeric myocytes is to analyze hearts of patients who have undergone gender-mismatched bone marrow transplantation (BMT). The presence of Y chromosome-positive cardiomyocytes in the hearts of female patients would conclusively suggest a bone marrow origin for these cells. By using fluorescence in situ hybridization (FISH) combined with

[Methods](#)
[Results](#)
[Discussion](#)
[References](#)

[Top](#)
[Abstract](#)
[Introduction](#)
[Methods](#)
[Results](#)
[Discussion](#)
[References](#)

immunohistochemistry, we determined the percentage of male cardiomyocytes in autopsy hearts of female patients who had undergone gender-mismatched BMT. To exclude the possibility of false identification of pseudo or fusion nuclei as chimeric cardiomyocytes, additional analyses were performed with the use of basement membrane laminin containing and chromosome 18 multiploidy analysis with FISH, respectively. Gender-matched BMT patients served as controls.

Methods

Patients and Autopsy Tissue

A review of the Mayo Clinic BMT database and autopsy records identified 4 female patients who had received male donor bone marrow. Female patients who had gender-matched BMT were examined as controls. The Mayo Clinic institutional review board granted approval for the study of human tissue samples.

Combined Immunohistochemical and FISH Analysis

Immunohistochemical analysis of cardiac tissue sections was performed by using a monoclonal antibody against -sarcomeric actin (Sigma clone 5c5) and a rabbit antibody against laminin (Sigma, St Louis, Mo). The secondary detection used was respectively an anti-mouse antibody conjugated to Cy-3 (Molecular Probes; red) and an anti-rabbit antibody conjugated to Alexa Fluor (Molecular Probes; green). In separate experiments, liver and skeletal muscle tissue from the same subjects was stained with antibodies to human hepatocyte and skeletal muscle actin with the use of monoclonal antibodies (both from Dako). Hepatocytes and skeletal myocytes were visualized using a secondary anti-mouse antibody conjugated to Cy-3.

After immunostaining, FISH was immediately performed as previously described.³ The X and Y chromosome (CEP X, Y; Vysis Inc; B7322, B-6927) DNA probes used were specific for the satellite region of each chromosome and labeled with Cy-3 and fluorescein isothiocyanate, respectively. For combined analysis, sarcomeric actin and laminin staining and FISH for Y-chromosome were used. In separate experiments a probe to the centromere of human chromosome 18 (CEP 18 Aqua; light blue dot; Vysis) was combined with X (red dot) and Y chromosome (green dot) analysis to evaluate cell ploidy and exclude cell fusion in the chimeric nuclei identified.

In all cases, FISH signals were enumerated using a Zeiss Axioplan microscope equipped with a triple-pass filter (Vysis). Rigorous criteria were used to identify Y chromosome-positive cardiac myocytes as previously described.² Counting of the nuclei and Y chromosome was performed by two independent blinded observers.

Top
Abstract
Introduction
Methods
Results
Discussion
References

► **Results**

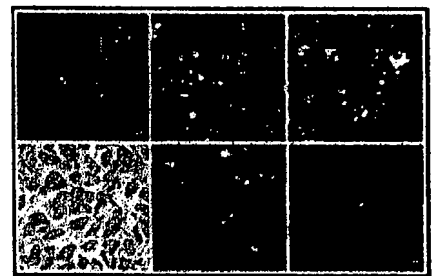
Patient Characteristics

The clinical profiles of the 4 female patients who underwent sex-mismatched BMT are shown in the [Table](#). Subjects had a range of hematologic diseases requiring BMT ($2.8 \pm 0.5 \times 10^8$ infused cells/kg body weight) and received the same pretransplantation conditioning regimen, which consisted of total body irradiation and cyclophosphamide. All patients were maintained on prednisone, and 2 subjects were maintained on additional cyclosporine and azathioprine after transplantation. Autopsy examination showed no macroscopic or microscopic evidence of inflammation in any of the hearts studied ([Figure, A](#)).

▲ Top
▲ Abstract
▲ Introduction
▲ Methods
• Results
▼ Discussion
▼ References

View this table: Clinical Data and Cardiomyocyte Chimerism Analysis of Gender-Mismatched BMT Patients

[\[in this window\]](#)
[\[in a new window\]](#)



View larger version
(88K):
[\[in this window\]](#)
[\[in a new window\]](#)

A, Hematoxylin-and-eosin staining of normal left ventricular myocytes showing no evidence of inflammatory cell infiltrate. B, Cardiomyocyte of female gender-mismatched BMT patient staining positive for α -sarcomeric actin (red) possessing nuclei (blue) positive for Y chromosome (green dot). B, inset, Diploid bone marrow–derived cardiomyocyte nucleus of female gender-mismatched BMT patient showing X chromosome (open arrowhead, red dot), Y chromosome (green dot), and a pair of chromosome 18 (filled arrows, light blue dots) signals; note overlying and surrounding red staining for α -sarcomeric actin. C, Y chromosome–positive true nucleus (blue, green dot; arrow) of bone marrow–derived cardiomyocyte cytoplasm (sarcomeric actin, red) surrounded by basement membrane laminin (green, arrowhead). D, Y chromosome–positive pseudonucleus (open arrowhead) separated from cardiomyocyte (sarcomeric actin, red) by laminin (green-filled arrowheads). E and F, Combined immunofluorescence staining and FISH for Y chromosome in female gender-mismatched BMT subjects showing (E) male skeletal muscle cell (red cytoplasm and blue nucleus with green dot-arrow) and (F) male hepatocyte (red cytoplasm and blue nucleus with green dot-arrow). Note a male cell (open arrowhead) that does not contain with hepatocyte antibody.

Immunofluorescence and FISH Analysis

Histological sections of the left ventricle in gender-mismatched subjects revealed a mean percentage of Y chromosome-positive cardiac myocytes of 0.23±0.06% (Table). The Y chromosome was located eccentrically within the nuclei of chimeric cardiomyocytes (Figure, B and C), and chromosomal ploidy analysis excluded cell fusion (Figure, B, inset). Four female control patients who had undergone sex-matched BMT showed no evidence of Y chromosome positivity in any of the 80 000 cardiomyocyte nuclei analyzed. Basement membrane laminin and sarcomeric actin containing distinguished true chimeric nuclei with surrounding myocyte cytoplasm from pseudonuclei (Figure, C and D). Male bone marrow-derived hepatocytes and skeletal myocytes were also found in the liver and muscle of female gender-mismatched BMT recipients (Figure, E and F), and mean donor cell chimerism in these tissues was 0.4% and 0.2%, respectively (3000 nuclei analyzed). The detection sensitivity of FISH for Y chromosome in this study was 45%, similar to that cited in previous FISH analysis of tissue sections.^{2,4}

Discussion

These data suggest that adult human bone marrow acts as a source of extracardiac progenitor cells contributing to cardiomyocyte formation. The additional use of laminin containing and chromosomal ploidy analysis in this study makes the possibility of confusing pseudonuclei or cell fusion events for chimeric myocytes unlikely. The potential origin and phenotype of marrow myocyte precursors in our subjects includes lineage-restricted mesenchymal,² hematopoietic,¹⁰ and multipotent adult progenitors⁹ and cells of angioblastic lineage.¹¹

Top
Abstract
Introduction
Methods
Results
Discussion
References

Physiological stress and tissue injury are known to release cytokines and chemokines, which may promote mobilization of progenitor cells from the bone marrow to the peripheral circulation.¹² Although no patients in our study group had histological evidence of myocardial inflammation, 3 of 4 patients had respiratory complications such as adult respiratory distress syndrome and bronchiolitis obliterans. It is possible that severe tissue injury occurring in these conditions resulted in high levels of circulating cytokines with consequent mobilization of circulating progenitor cells. Interestingly, prior animal experiments showed no detectable engraftment of marrow-derived cells in the absence of myocardial injury.⁵ The difference between these animal data and our study may reflect differences in species, duration of study, use of "side population" cells exclusively versus unfractionated bone marrow, or other poorly understood phenomena associated with clinical disease and its treatment.

The consistent levels of chimerism seen at 5 weeks and 20 months after marrow transplantation in our present study suggest a steady-state recruitment of marrow progenitors rather than an initial seeding event early after transplantation. It is noteworthy that a similar

recruitment of bone marrow cells occurred in the liver and skeletal muscle as well as the heart, which validates previous animal and human data suggesting multipotent differentiation potential for bone marrow–derived cells.^{11,13} It is well known that marrow-derived mesenchymal and hematopoietic stem cells circulate for long periods after transplantation, allowing an equilibrium to be established between circulating and tissue-specific seeding compartments. It is therefore conceivable that low-level recruitment of blood-borne precursors into the myocardium occurs in response to local events in the tissue microenvironment.

Another possibility is that myocardial injury secondary to the pretransplantation conditioning regimen leads by a repair response to recruitment of marrow precursors into the myocardium. This scenario seems less likely, however, as the degree of chimerism would be expected to decrease over time and a concurrent "response to injury" would be expected from other blood-borne cells such as leukocytes, neither of which was seen in our study. Furthermore, because all our patients had established hematologic disease before BMT, we cannot automatically infer that chimeric events seen in our study occur under normal healthy conditions, nor can we exclude the possibility that pretransplantation disease may have altered posttransplantation seeding of circulating cells. Finally, we can only speculate on the additional modulating effects of immunosuppressive therapy on bone marrow cell recruitment in our subjects.

The mean percentage of bone marrow–derived cardiac myocytes observed in our subjects was low. It is difficult if not impossible to compare our data with previous chimerism studies both from a clinical and methodological perspective^{1–4,7} because it is likely that variables such as chimeric cell detection method, time of study after transplantation, and the presence or absence of inflammation influence the level of myocyte chimerism observed. Finally, while this manuscript was under review, Thiele et al¹⁴ reported 6.4% cardiomyocyte chimerism in a group of male bone marrow transplantation patients, a level more than an order of magnitude greater than our findings. However, the small number of nuclei analyzed and the use of morphology instead of myocyte-specific staining make the identification of chimeric nuclei as true cardiomyocytes less certain in this study.

In conclusion, the present study establishes bone marrow as a contributor to low-level de novo cardiac myocyte formation. The clinical significance of this finding in terms of myocardial regeneration will depend on the success of future efforts to understand and augment the mobilization, homing, and differentiation properties of these cells. Further investigation may also determine whether these cells can be engineered or targeted to diseased myocardium for therapeutic effect.

Acknowledgments

This work was supported in part by grants from the National Institutes of Health (HL66958-NMC) and Mayo Foundation.

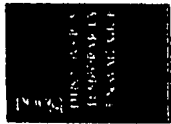
Received November 13, 2002; revision received January 23, 2002; accepted January 23, 2002.

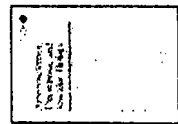
References


1. Hruban RH, Long PP, Perlman EJ, et al. Fluorescence in situ hybridization for the Y-chromosome can be used to detect cells of recipient origin in allografted hearts following cardiac transplantation. *Am J Pathol.* 1993; 142: 975–980. [Abstract]
2. Laflamme MA, Myerson D, Saffitz JE, et al. Evidence for cardiomyocyte repopulation by extracardiac progenitors in transplanted human hearts. *Circ Res.* 2002; 90: 634–640. [Abstract/Free Full Text]
3. Muller P, Pfeiffer P, Koglin J, et al. Cardiomyocytes of noncardiac origin in myocardial biopsies of human transplanted hearts. *Circulation.* 2002; 106: 31–35. [Abstract/Free Full Text]
4. Quaini F, Urbanek K, Beltrami AP, et al. Chimerism of the transplanted heart. *N Engl J Med.* 2002; 346: 5–15. [Abstract/Free Full Text]
5. Jackson KA, Majka SM, Wang H, et al. Regeneration of ischemic cardiac muscle and vascular endothelium by adult stem cells. *J Clin Invest.* 2001; 107: 1395–1402. [Abstract/Free Full Text]
6. Taylor DA, Hruban R, Rodriguez ER, et al. Cardiac chimerism as a mechanism for self-repair: does it happen and if so to what degree? *Circulation.* 2002; 106: 2–4. [Free Full Text]
7. Glaser R, Lu MM, Narula N, et al. Smooth muscle cells, but not myocytes, of host origin in transplanted human hearts. *Circulation.* 2002; 106: 17–19. [Abstract/Free Full Text]
8. Terada N, Hamazaki T, Oka M, et al. Bone marrow cells adopt the phenotype of other cells by spontaneous cell fusion. *Nature.* 2002; 416: 542–545. [CrossRef][Medline]
9. Jiang Y, Jahagirdar BN, Reinhardt RL, et al. Pluripotency of mesenchymal stem cells derived from adult marrow. *Nature.* 2002; 418: 41–49. [CrossRef][Medline]
10. Orlic D, Kajstura J, Chimenti S, et al. Bone marrow cells regenerate infarcted myocardium. *Nature.* 2001; 410: 701–705. [CrossRef][Medline]
11. Korbling M, Katz RL, Khanna A, et al. Hepatocytes and epithelial cells of donor origin in recipients of peripheral-blood stem cells. *N Engl J Med.* 2002; 346: 738–746. [Abstract/Free Full Text]
12. Dominko T, Takahashi D, Martinovich C, et al. Ischemia- and cytokine-induced mobilization of bone marrow–derived endothelial progenitor cells for neovascularization. *Nat Med.* 1999; 5: 431–433. [CrossRef][Medline]
13. Krause DS, Theise ND, Collector MI, et al. Multi-organ, multi-lineage engraftment by a single bone marrow–derived stem cell. *Cell.* 2001; 105: 369–377. [Medline]
14. Thiele J, Varus E, Wickenhauser C, et al. Chimerism of cardiomyocytes and endothelial cells after allogeneic bone marrow transplantation in chronic myeloid leukemia: an autopsy study. *Pathology.* 2002; 23: 405–410. [CrossRef][Medline]

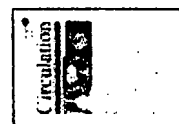
Top
Abstract
Introduction
Methods
Results
Discussion
References

This article has been cited by other articles:

-  **blood** [▶ HOME](#)
V. Meignin, J. Soulier, F. Brau, M. Lemann, E. Gluckman, A. Janin, and G. Socie
Little evidence of donor-derived epithelial cells in early digestive acute graft-versus-host disease
Blood, January 1, 2004; 103(1): 360 - 362.
[\[Abstract\]](#) [\[Full Text\]](#) [\[PDF\]](#)

-  **Arteriosclerosis, Thrombosis, and Vascular Biology** [▶ HOME](#)
J. D. Pearson
Using Endothelial Progenitor Cells for Gene Therapy
Arterioscler. Thromb. Vasc. Biol., December 1, 2003; 23(12): 2117 - 2118.
[\[Full Text\]](#) [\[PDF\]](#)

-  **Proceedings of the National Academy of Sciences** [▶ HOME](#)
D. J. Prockop, C. A. Gregory, and J. L. Spees
One strategy for cell and gene therapy: Harnessing the power of adult stem cells to repair tissues
PNAS, September 30, 2003; 100(90001): 11917 - 11923.
[\[Abstract\]](#) [\[Full Text\]](#) [\[PDF\]](#)

-  **Circulation** [▶ HOME](#)
H. M. Kvasnicka, C. Wickenhauser, J. Thiele, A. Deb, S. Wang, K. A. Skelding, D. Miller, D. Simper, and N. M. Caplice
Quantifying Chimeric Cardiomyocytes * Response
Circulation, August 26, 2003; 108(8): e60 - 60.
[\[Full Text\]](#) [\[PDF\]](#)

Abstract of this Article () Reprint (PDF) Version of this Article Citation Map Email this article to a friend Similar articles found in: Circulation Online PubMed PubMed Citation This Article has been cited by: Search Medline for articles by: Deb, A. Caplice, N. M. Alert me when: new articles cite this article Download to Citation Manager
Collections under which this article appears: Other myocardial biology Myogenesis
All Versions of this Article: 107/9/1247 (most recent) 01.CIR.0000061910.39145.F0v1

HOME HELP FEEDBACK SUBSCRIPTIONS ARCHIVE SEARCH TABLE OF CONTENTS
CIRCULATION ART, THRO, VASC BIO ALL AHA JOURNALS
CIRCULATION RESEARCH HYPERTENSION STROKE

EVIDENCE APPENDIX

Item 4

Skeletal Myoblast Transplantation for Repair of Myocardial Necrosis

Charles E. Murry,* Robert W. Wiseman,* Stephen M. Schwartz,* and Stephen D. Hauschka[‡]

*Department of Pathology, †Department of Radiology, and ‡Department of Biochemistry, University of Washington School of Medicine, Seattle, Washington 98195

Abstract

Myocardial infarcts heal by scarring because myocardium cannot regenerate. To determine if skeletal myoblasts could establish new contractile tissue, hearts of adult inbred rats were injured by freeze-thaw, and $3\text{--}4.5 \times 10^6$ neonatal skeletal muscle cells were transplanted immediately thereafter. At 1 d the graft cells were proliferating and did not express myosin heavy chain (MHC). By 3 d, multinucleated myotubes were present which expressed both embryonic and fast fiber MHCs. At 2 wk, electron microscopy demonstrated possible satellite stem cells. By 7 wk the grafts began expressing β -MHC, a hallmark of the slow fiber phenotype; co-expression of embryonic, fast, and β -MHC continued through 3 mo. Transplanting myoblasts 1 wk after injury yielded comparable results, except that grafts expressed β -MHC sooner (by 2 wk). Grafts never expressed cardiac-specific MHC- α . Wounds containing 2-wk-old myoblast grafts contracted when stimulated *ex vivo*, and high frequency stimulation induced tetanus. Furthermore, the grafts could perform a cardiac-like duty cycle, alternating tetanus and relaxation, for at least 6 min. Thus, skeletal myoblasts can establish new muscle tissue when grafted into injured hearts, and this muscle can contract when stimulated electrically. Because the grafts convert to fatigue-resistant, slow twitch fibers, this new muscle may be suited to a cardiac work load. (*J. Clin. Invest.* 1996. 98:2512–2523.) Key words: myocardial infarction • skeletal myoblast • myosin heavy chain • contractile function • cell transplantation

Introduction

Experimental and clinical therapies for myocardial infarction have focused traditionally on limiting infarct size. Unfortunately, the goal of limiting myocardial injury has been difficult to achieve clinically, because ischemic myocardium dies quite rapidly (1) and most patients wait more than 3 h after coronary occlusion before seeking medical attention. As an alternative approach, we are exploring strategies to induce the injured heart to heal with muscle replacement rather than forming scar tissue.

Address correspondence to Charles E. Murry, M.D., Ph.D., Department of Pathology, Box 357470, Room E-520, HSB, University of Washington School of Medicine, Seattle, WA 98195. Phone: 206-616-8685; FAX: 206-543-3644; E-mail: murry@u.washington.edu

Received for publication 18 April 1996 and accepted in revised form 2 October 1996.

J. Clin. Invest.

© The American Society for Clinical Investigation, Inc.
0021-9738/96/12/2512/12 \$2.00

Volume 98, Number 11, December 1996, 2512–2523

One strategy for muscle regeneration is to transplant either skeletal or cardiac myocytes into the injured heart. Studies from Field's group showed that cardiac myocytes can be transplanted into normal hearts, where they couple with host cardiocytes via intercalated discs (2, 3). However, a major drawback to using cardiocytes is their inability to proliferate in culture. At present it seems unlikely that enough primary cardiocytes could be obtained from the patient or histocompatible donor to repair a myocardial infarct in humans. On the other hand, skeletal muscle satellite cells (muscle stem cells) proliferate well in culture. Satellite cells could be obtained from muscles of infarct patients and rapidly expanded in culture, or stocks of potentially therapeutic myoblasts could be obtained from embryos and frozen for subsequent use (4, 5). Furthermore, physiological studies have shown that when properly conditioned, skeletal muscle can adapt to perform a cardiac-type work load (6). Recent studies have demonstrated the feasibility of grafting skeletal myoblast lines into normal hearts (7) and autologous satellite cells into injured hearts (8, 9). However, to generate significant amounts of functional new muscle the transplanted cells ideally should proliferate and then differentiate into mature myofibers capable of sustaining a cardiac work load. This study was performed to determine the proliferation and differentiation patterns of skeletal myoblasts after engraftment into injured rat hearts and to determine whether this new muscle could support contractile activity.

Methods

Skeletal myoblast isolation and culture. These studies were approved by the University of Washington Animal Care Committee and were conducted in accordance with federal guidelines. Skeletal myoblasts were obtained from the limbs of 1–3-d-old Fischer rats. This inbred strain was used to avoid immune barriers to transplantation. After time of killing, the carcasses were skinned and the limbs were placed into cold tissue culture media. Under a dissecting microscope, the muscles were stripped of surrounding adipose tissue and fascia and bluntly dissected from their tendons. The muscles were minced with iridectomy scissors until a fine slurry was formed. The slurry was then digested in 0.05% trypsin/EDTA (GIBCO-BRL, Gaithersburg, MD) in Ham's saline A at 37°C, with intermittent mechanical agitation to assist dispersal. After 30–45 min the cell suspension was filtered through sterile gauze to remove undispersed tissue fragments and rod shaped mature myofibers. Cells were plated at $\sim 5 \times 10^6$ cells/dish in 100-mm gelatinized plates in 10 ml Ham's F10C media, containing 15% horse serum and 50 $\mu\text{g/ml}$ gentamicin sulfate (Gemini Bioproducts, Inc., Calabasas, CA). Recombinant human basic fibroblast growth factor was added twice daily to a final concentration of 6 ng/ml, and the complete medium was replaced once per day. Approximately 10% of the cells attached and grew with a doubling time of ~ 18 h. The cultures contained a mix of small, oval myoblasts and elongated, spindle-shaped cells consistent with fibroblasts. Subconfluent cultures were passaged every 2–3 d (1:5 split) to minimize the occurrence of myogenic differentiation at higher density. On the day before transplantation, the cultures were tagged for subsequent identification *in vivo*. In some experiments cells were tagged with fluorescent micro-

spheres (1:500 dilution of stock 200 nm yellow-green fluorescent microspheres; Molecular Probes, Eugene, OR). The latex microspheres were endocytosed (typically > 20 spheres/cell) and served as cytoplasmic markers (10). In other experiments, cells were incubated overnight with [³H]thymidine (1 μ Ci/ml) to mark their nuclei after autoradiography. Cultures were trypsinized immediately before transplantation and suspended at a concentration of $\sim 3 \times 10^6$ /ml. Small aliquots of the remaining cell suspension were replated at $\sim 2 \times 10^4$ cells/cm² into gelatinized, multichamber plastic slides, and fixed in methanol after various culture intervals for immunostaining.

Rat cardiac injury models. Inbred male Fischer rats (Simonsen Labs, Gilroy, CA) weighing 350–400 grams were anesthetized with intraperitoneal ketamine-xylazine (68 and 4.4 mg/kg, respectively), intubated, and mechanically ventilated with room air. The heart was exposed aseptically via a left thoracotomy, and a 1-cm-diameter aluminum rod, precooled with liquid nitrogen, was placed in direct contact with the anterior left ventricle for 15 s. Freeze-thaw reproducibly caused a disc-shaped region of coagulation necrosis, ~ 1 cm in diameter, extending ~ 2 mm into the myocardium. It should be noted that while infarcts typically have irregular borders with viable peninsulas of subepicardial myocardium along penetrating vessels, freeze-thaw lesions consist of confluent necrosis in the subepicardium with viable myocardium in the subendocardium. Because they are not transmural, freeze-thaw lesions do not cause cardiac aneurysm formation. Despite these differences, the cellular patterns of coagulation necrosis, inflammation and phagocytosis, granulation tissue formation, and scarring after freeze-thaw injury are indistinguishable from myocardial infarction (11–13), making it a suitable model to study myocardial repair.

In the initial studies, $\sim 3 \times 10^6$ myoblasts in 100 μ l tissue culture media were injected superficially into the center of the injured region immediately after injury, using a 27-gauge needle. Then, the chest was closed and the rats were allowed to recover for timed intervals from 1 d to 3 mo ($n = 4$ /time point). To mimic a clinical situation more closely, a second protocol was used in which the freeze-thaw lesion was allowed to heal for 1 wk before transplanting myoblasts. By 1 wk, most of the necrotic myocardium had been replaced by granulation tissue, but scar formation had not yet begun. The rats ($n = 2$ /time point; no 3 d or 3 mo time points) were reanesthetized and a thoracotomy was repeated. The heart was exposed and a 100- μ l suspension containing $\sim 3 \times 10^6$ myoblasts was injected into the wound as described above. The chest was closed and the animals were allowed to recover for intervals from 1 d to 7 wk.

To detect DNA synthesis in the grafts the rats were treated with the thymidine analogue 5-bromo-2'-deoxyuridine (BrdU) (Boehringer-Mannheim, Indianapolis, IN). 1 d before time of killing, the rats were lightly anesthetized, and a 50-mg tablet of BrdU was implanted subcutaneously for measurement of cell replication. Preliminary studies showed that a subcutaneous 50-mg BrdU tablet gave comparable replication rates to a 24-h continuous infusion with an osmotic mini-pump (not shown). For rats killed 1 d after transplantation, a single 10-mg pulse of BrdU was given intraperitoneally 1 h before time of killing. This avoided incorporation of BrdU into the cells which were cycling at the time of transplantation.

Rats were killed with a pentobarbital overdose and their hearts were excised. In the immediate transplantation groups, the aorta was cannulated and the hearts were perfusion fixed with methyl Carnoy's solution (60% methanol, 30% chloroform, 10% glacial acetic acid), transversely sectioned, and embedded in paraffin by routine methods. In groups transplanted 1 wk after injury, the hearts were transversely sectioned, embedded in OCT (Miles Inc., Kankakee, IL), and frozen in a dry ice-ethanol bath for frozen section analysis. In both protocols, sections of gut were obtained as controls for measurement of cell replication with BrdU.

1. **Abbreviations used in this paper:** BrdU, 5-bromo-2'-deoxyuridine; MHC, myosin heavy chain.

Measurement of contractile function in isolated wound strips. Rat hearts were given 4.5×10^6 myoblasts ($n = 8$) in 100 μ l or a sham injection of saline ($n = 3$) immediately after injury. 2 wk after engrafting, the hearts were excised and transversely sectioned. Under a dissecting microscope, most of the subendocardial myocardium was trimmed away from the injured region, and isolated wound strips ($\sim 1.5 \times 1.5 \times 8$ mm) were prepared. One or two strips were studied from each sham-engrafted heart, and two or three strips were studied from each myoblast-engrafted heart. The strips were ligated at both ends with silk suture and then placed in a bath of physiological saline with the following composition (mmol/liter): 116 NaCl, 4.6 KCl, 1.2 MgSO₄, 2.5 CaCl₂, 26 Mops (pH 7.4), 11 glucose, and 10 mg/liter gentamicin. The buffer was equilibrated with 95% O₂/5% CO₂ and maintained at 20°C via a thermostatically controlled water jacket. Wound strips were mounted between an isometric force transducer (model 60-2995; Harvard Apparatus, Inc., South Natick, MA) and a fixed glass hook. Resting tension was set initially at 0.5 g. Strips were stimulated with 1-ms bipolar pulses delivered via platinum wire electrodes using a Grass model S48 stimulator (Astro-Med, Inc., West Warwick, RI). Voltage was increased in 10-V increments until contractile activity was observed. Force traces were displayed on a digital storage oscilloscope (model 3091; Nicolet Instrument Corp., Madison, WI) and recorded using a General Scanning model RS4-5P strip chart recorder. After determining the force-voltage relationship, the optimal length for force production was determined for each wound strip using test contractions at 2-min intervals, a time sufficient for metabolic recovery in mammalian fast twitch muscles (14). Force-frequency analysis was performed by increasing the stimulation frequency in 1-Hz increments; tetanus was defined as the point where the oscillations of contractile force at the plateau were < 3% of the net force generated (14). Finally, to test fatigability the grafts were subjected to a simulated cardiac-like duty cycle, consisting of 0.33 s of tetanus followed by 0.67 s of relaxation (1:2 cycle), continuing for 6 min. After completion of functional studies the strip's cross-sectional area was determined, and the tissue then was processed for histology or electron microscopy.

Immunocytochemistry. Antibodies used for immunostaining are given in Table I. 6- μ m frozen sections were cut on a cryostat, briefly air dried, and stored at -70°C until use. 5- μ m paraffin sections were deparaffinized in xylene and rehydrated in a graded alcohol series. Cultured cells were fixed and stored in cold PBS until use. For all samples, endogenous peroxidase activity was quenched by incubating with 0.3% H₂O₂ in methanol for 30 min. Immunostaining was carried out at room temperature. Sections were blocked with 1.5% normal horse serum in PBS for 1 h. The sections were then incubated with the primary antibody in 1.5% horse serum for 1 h, followed by incubation with the secondary antibody (rat adsorbed horse anti-mouse, 1:400 dilution; Vector Labs, Inc., Burlingame, CA) for 1 h. Antigens were localized with an avidin-biotin-peroxidase complex (ABC Elite kit; Vector Labs). For staining with a single antibody, diaminobenzidine (Sigma Immunochemicals, St. Louis, MO) was used as a chromogenic substrate. For double immunolabeling with antibodies to myosin and BrdU, sections were first exposed to 1.5 N HCl for 15 min at 37°C to denature the DNA, followed by a rinse in 0.1 mol/liter borax to stabilize the denatured strands. Sections were then stained routinely for myosin heavy chain (MHC) using diaminobenzidine. After a second quenching in 0.3% H₂O₂, sections were blocked with 1.5% normal horse serum, and then incubated with a mouse monoclonal antibody to BrdU for 1 h. After incubation with the secondary antibody (horse anti-mouse), BrdU was localized with an avidin-biotin-peroxidase complex, using True Blue (KPL, Gaithersburg, MD) as substrate. Cross-reactivity between the first primary antibody and the second secondary antibody did not occur, as long as the True Blue substrate was incubated for a short duration (< 1 min). Sections were counterstained either with methyl green, nuclear fast red, or hematoxylin.

Electron microscopy. After measurement of contractile function, one of the tissue strips was immersed in half strength Karnovsky's fix-

Table 1. Antibodies Used for Immunocytochemistry

Antibody	Antigen recognized	Dilution	Source	Reference
MF-20	Sarcomeric MHCs	Hyb. Sup., 1:100	American Type Culture Collection, Rockville, MD	39
MY-32	Skeletal MHC-fast (types IIA and IIB)	Mouse ascites, 1:2000	Sigma Immunochemicals	40
BA-G5	Cardiac MHC- α	Hyb. Sup., 1:5	American Type Culture Collection	41
F1.652	Embryonic MHC	Hyb. Sup., 1:100	Developmental Studies Hybridoma Bank*	42
A4.951	β -MHC	Hyb. Sup., 1:50	American Type Culture Collection	43
Anti-BrdU	BrdU	IgG, 1:50000	Eurodiagnostics, Apeldoorn, The Netherlands	44

IgG, purified IgG monoclonal antibody; *Hyb. Sup.*, hybridoma supernatant. * The monoclonal antibody F1.652, developed in the laboratory of Dr. Helen Blau, was obtained from the Developmental Studies Hybridoma Bank maintained by the Department of Pharmacology and Molecular Sciences, Johns Hopkins University School of Medicine, Baltimore, MD, and the Department of Biological Sciences, University of Iowa, Iowa City, IA, under contract NO1-HD-2-3144 from the National Institute of Child Health and Human Development.

ative and dissected into small cubes < 1 mm in greatest dimension. The tissue was fixed overnight in half strength Karnovsky's fixative, postfixed for 1 h in 1% osmium tetroxide at room temperature, dehydrated through a graded alcohol series followed by propylene oxide, and embedded in Medcast resin (Ted Pella, Inc., Redding, CA). Semithin sections were stained with toluidine blue and examined by light microscopy. Thin sections were cut from selected blocks, stained with lead citrate and uranyl acetate, and examined in a Jeol JEM 1200EXII transmission electron microscope. Representative areas were photographed.

Results

Characteristics of myoblast cultures. The muscle cultures contained a mixed cell population. At least 22% of the cells were skeletal muscle, as indicated by their staining for sarcomeric myosin after switching to a differentiation medium containing 1.5% serum and no FGF for 3 d. This procedure underestimates the true percentage of skeletal muscle cells by several-fold, since the nonmyogenic cells continue to divide after the medium switch while the myoblasts complete their present cell cycle and then terminally differentiate. Approximately 1% of the cells stained with antibodies to smooth muscle α -actin, which can mark either smooth muscle cells or fibroblasts. Virtually none of the cells stained with an antibody for the endothelial marker von Willebrand factor. The remaining cells were presumably fibroblasts.

Histology and differentiation patterns of myoblast grafts. Cultured skeletal myoblasts were transplanted into cardiac freeze-thaw lesions either immediately after injury, or, to mimic a clinical situation more closely, cells were transplanted 1 wk after injury. The two protocols yielded similar results and will be described together; minor differences are noted below. On the first day after transplantation the myoblasts were mononuclear cells (Fig. 1 A). The grafted cells could be distinguished clearly from inflammatory cells within the necrotic tissue by their larger size and characteristic oval shape. (Fibroblast ingrowth from the surrounding tissue had not yet begun at this time.) The identity of the grafted cells was confirmed by their cytoplasmic fluorescent microspheres and radioactive nuclei (not shown). Mitotic figures were common. The grafted cells did not stain with antibodies to skeletal or cardiac MHCs (Fig. 1 B). Thus, muscle differentiation had not yet occurred.

By 3 d after transplantation, many of the grafted cells had fused to form multinucleated myotubes (Fig. 1 C). Myotubes were partially aligned along the short (transverse) axis of the

heart. The myotubes stained with antibodies to sarcomeric MHC, embryonic MHC (Fig. 1 D), and to MHC-fast (not shown). Occasional cross-striations were noted, but these were not frequent at this time (Fig. 1 D). The myotubes did not express cardiac MHC- α . By 1 wk the grafts were easily recognizable as skeletal myofibers and many cells contained cross-striations. As before, the new myofibers stained with antibodies to sarcomeric MHC, embryonic MHC, and MHC-fast, but did not express cardiac MHC- α (not shown). By 2 wk after transplantation the grafts had the appearance of maturing skeletal myofibers (Fig. 1 E). Sarcomeres were well formed, and many cells had peripheral nuclei. The myofibers stained intensely with antibodies to sarcomeric myosin, embryonic MHC (Fig. 1 F), and skeletal MHC-fast (Fig. 1 G). No staining with cardiac MHC- α antibodies was observed at 2 wk.

At 7 wk after transplantation the grafts were islands of mature skeletal muscle within young scar tissue (Fig. 1, H-J). There was a moderate increase in cell diameter compared with 2 wk. None of the muscle grafts were infiltrated or splayed apart by scar tissue, nor was there evidence of fiber atrophy. Vascular density appeared normal for muscle tissue (Fig. 1 J). All of the 7-wk grafts stained strongly with antibodies to sarcomeric myosin and embryonic MHC (Fig. 1 H). The grafts injected immediately after injury stained intensely with antiskeletal MHC-fast, comparable with Fig. 1 F. In contrast, the grafts injected 1 wk after injury stained poorly with antiskeletal MHC-fast (see below). No staining with antibodies to cardiac MHC- α was observed in the grafts, while the adjacent myocardium stained intensely (Fig. 1 I).

At 3 mo after transplantation the grafts again had the appearance of mature skeletal muscle (Fig. 1 K). Most myofibers had peripheral nuclei, and vascular density appeared normal. Fiber diameter was generally larger than in the 7-wk group, indicating that the cells had hypertrophied between 7 wk and 3 mo (compare Fig. 1, J and K). In one heart, however, part of the graft was infiltrated by scar tissue which encircled individual myofibers and was associated with fiber atrophy (Fig. 1 L). The grafts continued to express embryonic MHC and MHC-fast (not shown). Once again, no staining with antibodies to cardiac MHC- α was observed (comparable with Fig. 1 I). At all time points the myofibers were predominantly aligned parallel with the short (transverse) axis of the heart and therefore appeared in longitudinal section. However, some fascicles of muscle appeared obliquely or cross-sectioned in this plane.

In summary, the grafts began to differentiate into myo-

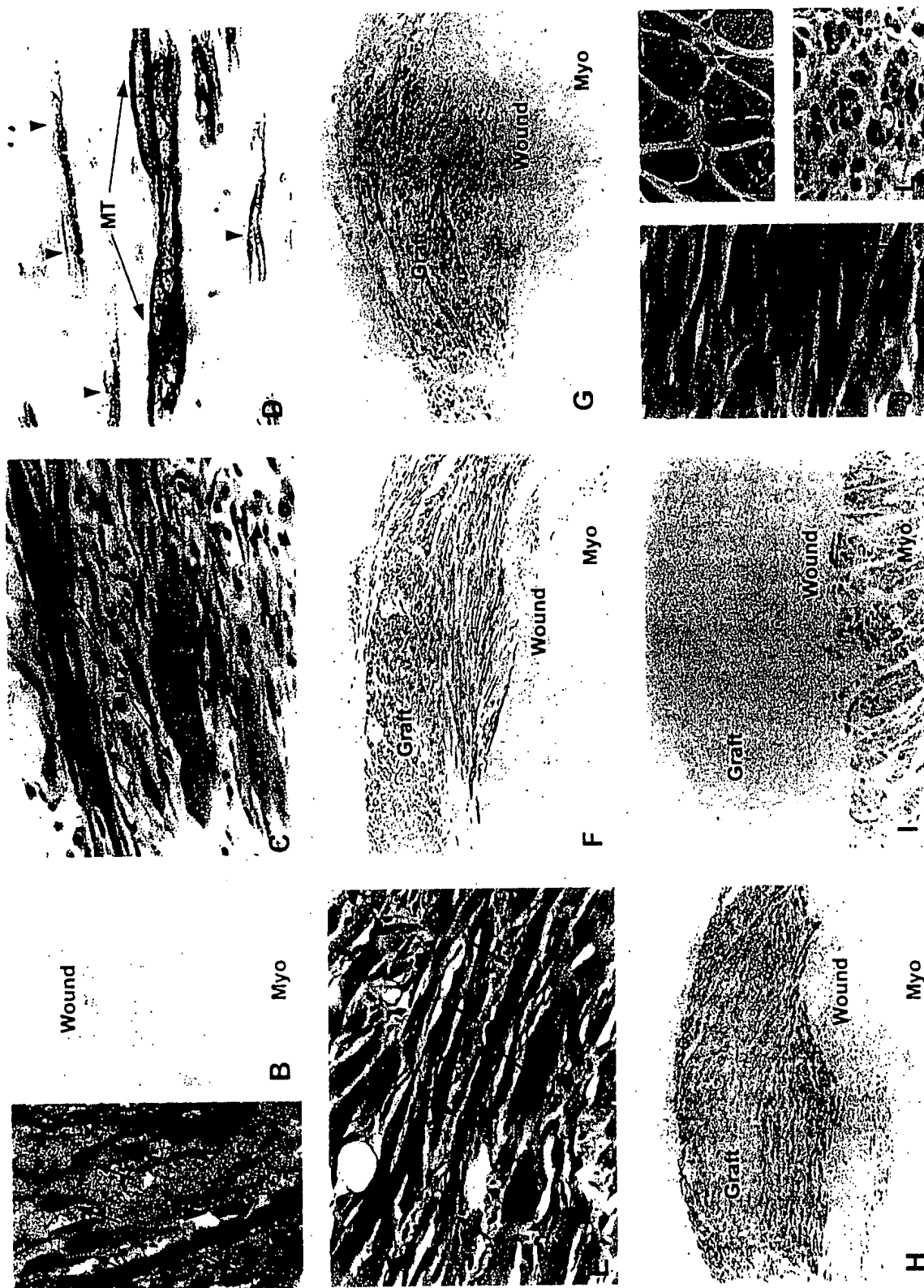


Figure 1. Morphology and MHC expression in skeletal myoblast grafts. Rat hearts were injured by freeze-thaw and syngeneic skeletal muscle cells were grafted into the lesions. All panels in this figure are from hearts which were grafted immediately after injury. (A) 1-d graft. The grafted cells are identifiable as relatively large, oval shaped cells (arrows) within the necrotic myocardium. One graft cell is in mitosis (arrowhead). Numerous smaller inflammatory cells are present within the lesion. Fibroblast ingrowth from surrounding viable tissue had not yet begun at this

tubes between 1 and 3 d and acquired the appearance of maturing myofibers with well formed sarcomeres by 2 wk. The grafts expressed both embryonic MHC and MHC-fast at all times between 3 d and 3 mo. There was no expression of cardiac MHC- α at any time.

Electron microscopy. Electron microscopy was performed on one heart, 2 wk after myoblast grafting. Most of the grafted cells had well formed, slightly contracted sarcomeres which were aligned in registry (Fig. 2 A). Mitochondria were abundant. Multinucleation was evident in many cells, as were well formed T-tubules. However, in other cells a spectrum of morphological stages was present, consistent with developing skeletal muscle (Fig. 2 B). Cells at the earliest stage were small, had scanty myofibril content, and contained focal aggregations of electron-dense material suggestive of developing Z-discs. In these cells there were abundant ribosomes and glycogen, a prominent Golgi apparatus, and dilated segments of sarcoplasmic reticulum. Intermediate cells were larger and had increasing amounts of myofibrils with a corresponding decrease in ribosomes and glycogen. Some cells had well formed sarcomeres, but these were out of registry compared with the most mature cells. No intercalated discs were identified between cells in the graft region. Adjacent myofibers often had intimately apposed, interdigitating cell membranes. Occasional cells were identified with electron-dense membrane structures suggestive of intermediate adherens junctions and gap junctions (Fig. 2, C and D). Some mature myofibers were closely associated with mesenchymal cells, located within the basal lamina compartment of the myofiber. Their location within the basal lamina of the myofiber suggests that they might be new satellite stem cells (Fig. 2, E and F). Some of these mesenchymal cells had abundant rough endoplasmic reticulum, similar to fibroblasts. Cells with this morphology have also been described in regenerating skeletal muscle by Trupin et al. (15). Their location within the basal lamina of the myofiber and the

absence of collagen in this space make it unlikely that these cells are actually fibroblasts.

Myoblast grafts convert from fast to slow twitch fibers. The poor staining for MHC-fast in the 7-wk group with delayed transplantation seemed at variance with the morphology of the grafts, which showed relatively hypertrophic cells with well formed sarcomeres. We hypothesized that the grafts had undergone fiber type conversion to slow twitch muscles, which no longer expressed high levels of MHC-fast. Slow twitch fibers have physiological similarities to cardiac muscle, including a high capacity for oxidative phosphorylation and fatigue resistance. Furthermore, slow fibers use β -MHC as a major contractile protein, which is also the predominant myosin in developing rat hearts. In contrast, fast twitch fibers use glycolysis for ATP production, have a low aerobic capacity and fatigue rapidly, and do not express β -MHC (16). Therefore, we compared β -MHC expression with skeletal MHC-fast, to determine fiber types in the maturing grafts.

At 1 wk the grafts stained intensely for MHC-fast (Fig. 3 A) but did not stain with an antibody to β -MHC (Fig. 3 B). At 2 wk the grafts continued to express MHC-fast. In the group transplanted immediately after injury no expression of β -MHC was noted at 2 wk, yet in grafts transplanted 1 wk after injury some cells expressed β -MHC (not shown). At 7 wk after transplantation the two groups differed in expression of MHC-fast, with strong staining in the immediate transplant group (see Fig. 1 G) and weak staining in the group where transplantation was delayed for 1 wk after injury (Fig. 3 C). However, both the immediate and delayed transplantation groups exhibited extensive staining for β -MHC at 7 wk after transplantation (Fig. 3 D). At 3 mo there was continued expression of β -MHC and MHC-fast in the immediate transplantation group; we did not study the delayed transplantation protocol at 3 mo. Thus, myoblast grafts appeared to be undergoing conversion from fast twitch to slow twitch fibers. Conversion appeared to take place

time. Hematoxylin and eosin stain. $\times 800$. (B) Low magnification of 1-d graft stained for embryonic MHC. The freeze-thaw lesion (Wound) occupies approximately the upper 75% of the field, while residual subendocardial myocardium (Myo) is present in the lower 25%. None of the grafted cells express embryonic MHC, indicating no differentiation had taken place yet. Methyl green counterstain. $\times 80$. (C) 3-d graft. Multiple multinucleated myotubes (MT) are present. Note that myotubes are already aligned in parallel. The surrounding tissue contains numerous fibroblasts (some of which may be of graft origin), macrophages, and capillaries, characteristic of granulation tissue. Two mitotic figures are present at the lower right (arrowheads). Hematoxylin and eosin stain. $\times 800$. (D) 3-d graft stained for embryonic MHC. The multinucleated myotubes (MT) express embryonic MHC, indicated by brown staining. Note faint cross-striations present at the periphery of some myotubes (arrowheads). Comparable staining was seen using antibodies to MHC-fast (not shown). Methyl green counterstain. $\times 800$. (E) 2-wk graft. Multinucleated myofibers are present and many have peripherally placed nuclei (arrows); most of these nuclei appear to be within the sarcolemma, although some may be immediately external. Cross-striations were readily seen under the microscope but appear faint in the photograph. Hematoxylin and eosin staining. $\times 800$. (F) 2-wk graft stained for embryonic MHC. The myofibers of the graft stain vigorously for embryonic MHC, while the underlying granulation tissue (Wound) and subendocardial myocardium (Myo) do not stain. Methyl green counterstain. $\times 80$. (G) 2-wk graft stained for fast fiber isoforms of MHC. There is intense staining of the engrafted myofibers (Graft), indicating that they exhibit a fast twitch phenotype. Note that the residual myocardium (Myo) beneath the graft does not stain, nor does the granulation tissue of the injured region (Wound). $\times 80$. (H) 7-wk graft stained for embryonic MHC. The graft continues to stain vigorously for embryonic MHC. There is no staining in the underlying young scar tissue (Wound) or the residual subendocardial myocardium (Myo). Methyl green counterstain. $\times 80$. (I) 7-wk graft stained for cardiac MHC- α . The skeletal myofibers of the graft do not express MHC- α , nor does the underlying scar tissue (Wound). This indicates that the grafted skeletal muscle does not show cardiac differentiation. The subendocardial myocardium (Myo) stains vigorously for MHC- α . Methyl green counterstain. $\times 80$. (J) 7-wk graft. Mature myofibers are present. Most myofibers have peripheral nuclei. Cross-striations were readily apparent under the microscope, but again are faint in the photograph. Multiple capillaries are present within the muscle tissue (arrows). Hematoxylin and eosin stain. $\times 800$. (K) 3-mo graft. The myofibers (obliquely and cross-sectioned) have peripheral nuclei and are closely apposed with little intervening extracellular matrix. The myofibers are hypertrophic compared with the 7-wk grafts (compare fiber diameter with J). Most 3-mo grafts had this appearance. Hematoxylin and eosin stain. $\times 800$. (L) 3-mo graft. The myofibers (cross-sectioned) in this region are encased by dense scar tissue and are atrophic. Note the markedly diminished cell diameters compared with K. Such entrapment of myofibers by scar was seen in one region of one heart. Hematoxylin and eosin stain. $\times 800$.

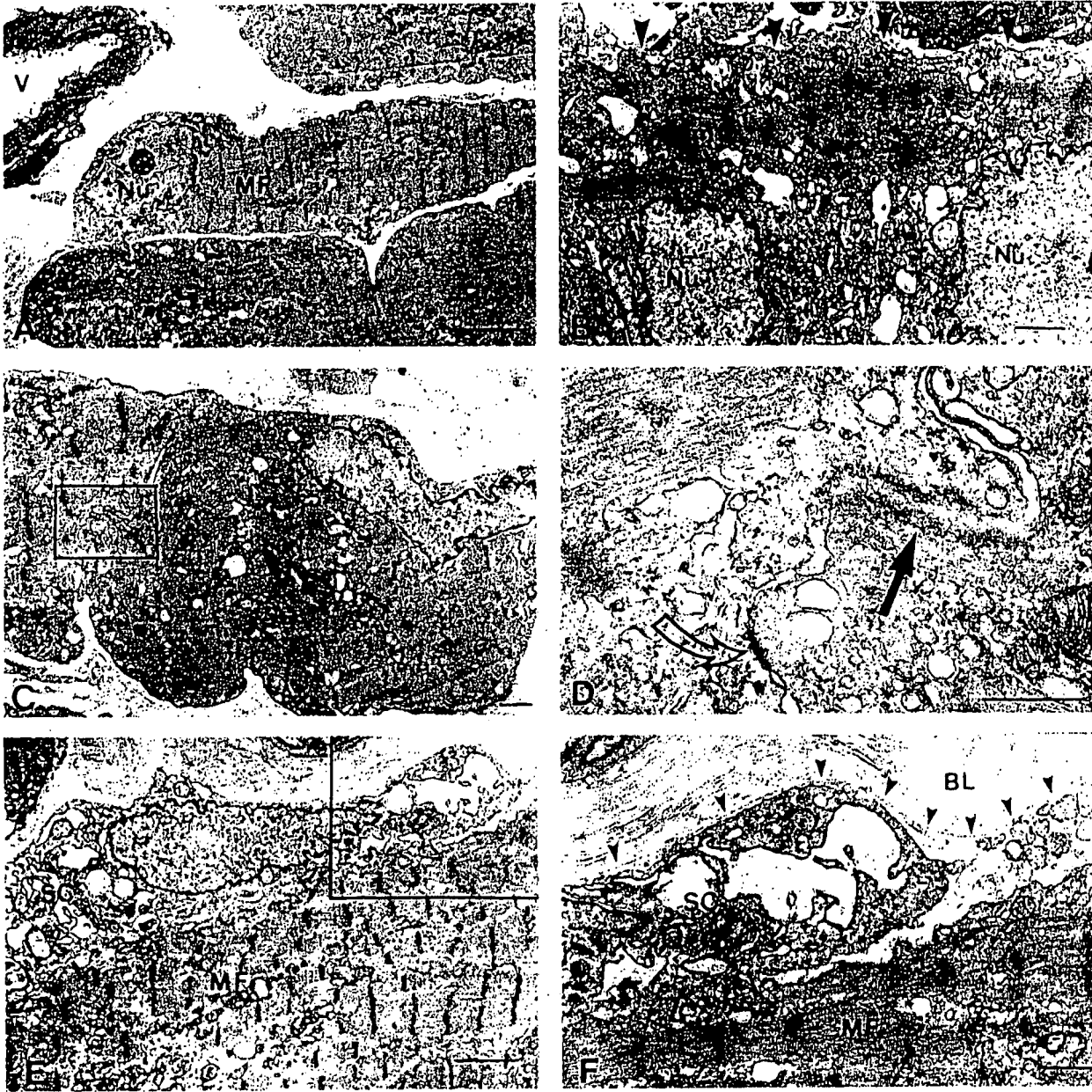


Figure 2. Transmission electron micrographs of 2-wk-old myoblast graft. The graft was placed immediately after cardiac freeze-thaw injury. (A) Low magnification overview showing well differentiated, striated skeletal myofibers (MF) within a collagen-rich matrix. A small venule (V) is shown at the left aspect. Nu, nucleus. Bar, 5 μ m. (B) Moderately differentiated skeletal myofiber containing two nuclei (Nu), a modest complement of myofibrils (mf), and abundant ribosomes and sarcoplasmic reticulum between the nuclei. The sarcolemma is delimited by arrowheads. Bar, 1 μ m. (C) Intercellular junction formation between adjacent myofibers. The two cells have closely apposed and interdigitated membranes. Two electron-dense plaques between the cells are present within the boxed region, suggestive of an adherens type intermediate junction and a gap junction, shown at higher magnification in D. Bar, 1 μ m. (D) Higher magnification of the junctional region boxed in C, showing putative intermediate junction between adjacent myofibers (solid arrow) and gap junction (open arrow). Bar, 0.5 μ m. (E) Skeletal myofiber (MF) with closely apposed mesenchymal cell atop it, suggestive of a satellite cell (SC). The boxed region is shown at higher magnification in F. Bar, 2 μ m. (F) Higher magnification of region boxed in E. The putative satellite cell (SC) and the myofiber (MF) are contained within the same basal lamina compartment (BL, outlined by arrowheads). Although the cell has abundant rough endoplasmic reticulum, its location within the basal lamina of the myofiber and the absence of fibrillar collagen from this space make it unlikely that this is a fibroblast. Bar, 1 μ m.

more rapidly when cells were transplanted into an injury with more advanced healing.

Proliferation of myoblast grafts. To identify cells undergoing DNA synthesis, the thymidine analogue BrdU was admin-

istered for 24 h before time of killing in most groups; animals in the day 1 group received a single pulse of BrdU 1 h before time of killing. Double immunostaining was performed with antibodies to the fast isoform of MHC and to BrdU, to detect

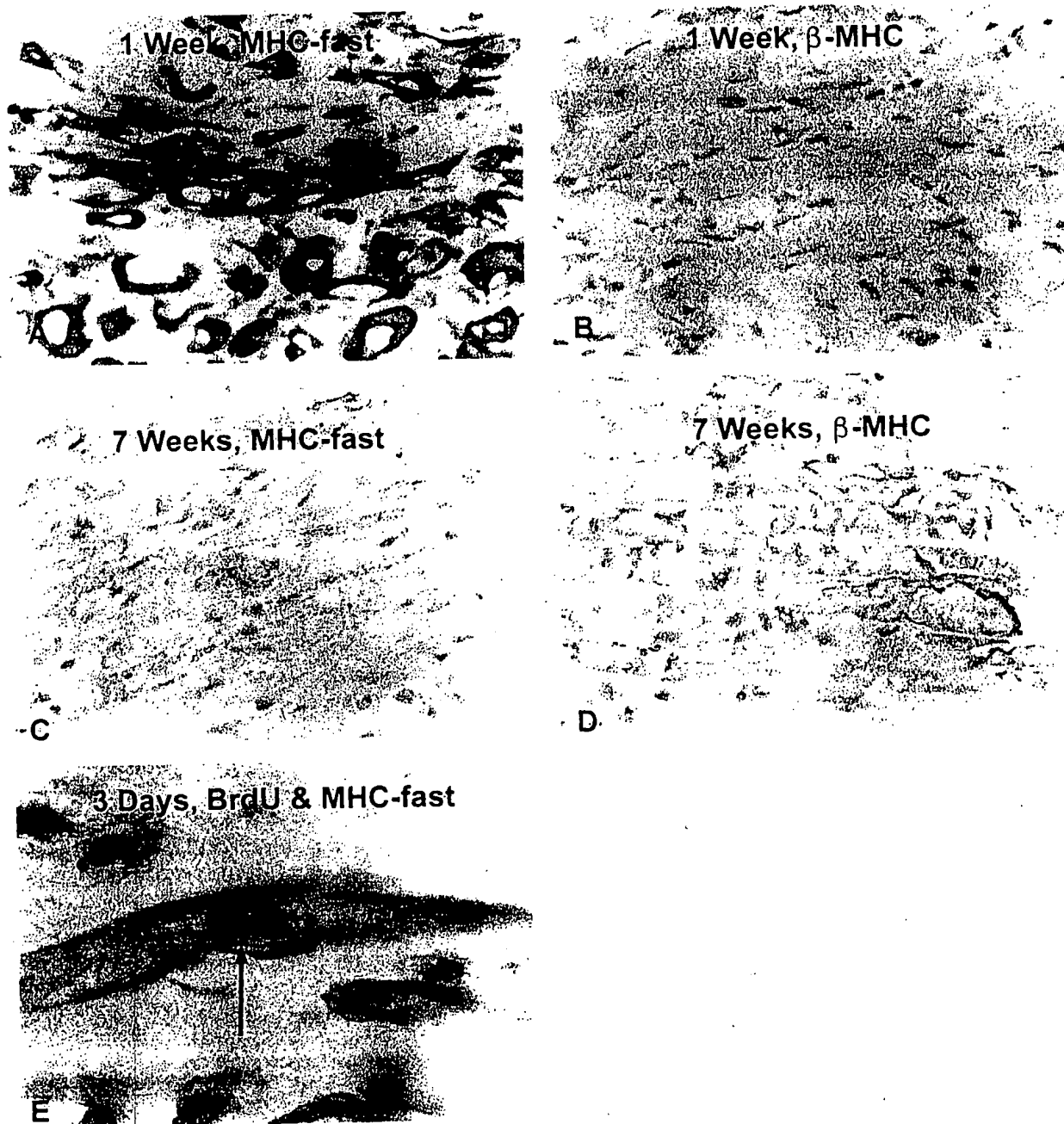


Figure 3. Fiber type conversion and proliferation of engrafted skeletal myoblasts. For the fiber typing experiments, rat hearts were injured by freeze-thaw and the lesions were allowed to heal for 1 wk. Syngeneic skeletal myoblasts were engrafted into the 1-wk-old wounds. For studies of cell proliferation, myoblasts were engrafted immediately after cardiac injury. Rats were killed at the indicated times after transplantation. Antibodies specific to fast twitch (MHC-fast) and slow twitch (β -MHC) fibers were used to define fiber types. Processing for frozen sections in A–D resulted in formation of contraction bands, artifactual clumping of the sarcomeres due to hypercontracture. BrdU was administered 24 h before time of killing to detect DNA synthesis. Double immunostaining for BrdU and MHC was then performed on paraffin sections. Appearance of a BrdU-positive nucleus within a myosin-positive cell indicated the myoblast had replicated and fused into the myotube within the last 24 h. (A) 1-wk graft stained for fast fiber isoforms of MHC. There is intense staining of the engrafted cells, indicating a fast fiber phenotype. Hematoxylin counterstain. $\times 960$. (B) 1-wk graft stained with an antibody to the slow fiber-specific β -MHC. None of the grafted cells express β -MHC at this time, indicating that the cells show no characteristics of slow fibers. Methyl green counterstain. $\times 960$. (C) 7-wk graft stained with an antibody to MHC-fast. There is weak staining compared with the 1-wk graft (A). Methyl green counterstain. $\times 960$. (D) 7-wk graft stained with an antibody to β -MHC. The grafted cells now express β -MHC, indicating that they are acquiring a slow fiber phenotype (compare with B). Methyl green counterstain. $\times 960$. (E) 3-d graft doubly stained for BrdU (purple) and MHC-fast (brown). One nucleus within the myotube stains purple (arrow), indicating it has undergone DNA replication before fusion into the myotube. The remaining nuclei in the myotube do not contain BrdU and pick up the red counterstain. Numerous myosin-negative cells in the surrounding wound tissue also stain positively for BrdU. Nuclear fast red counterstain. $\times 2,400$.

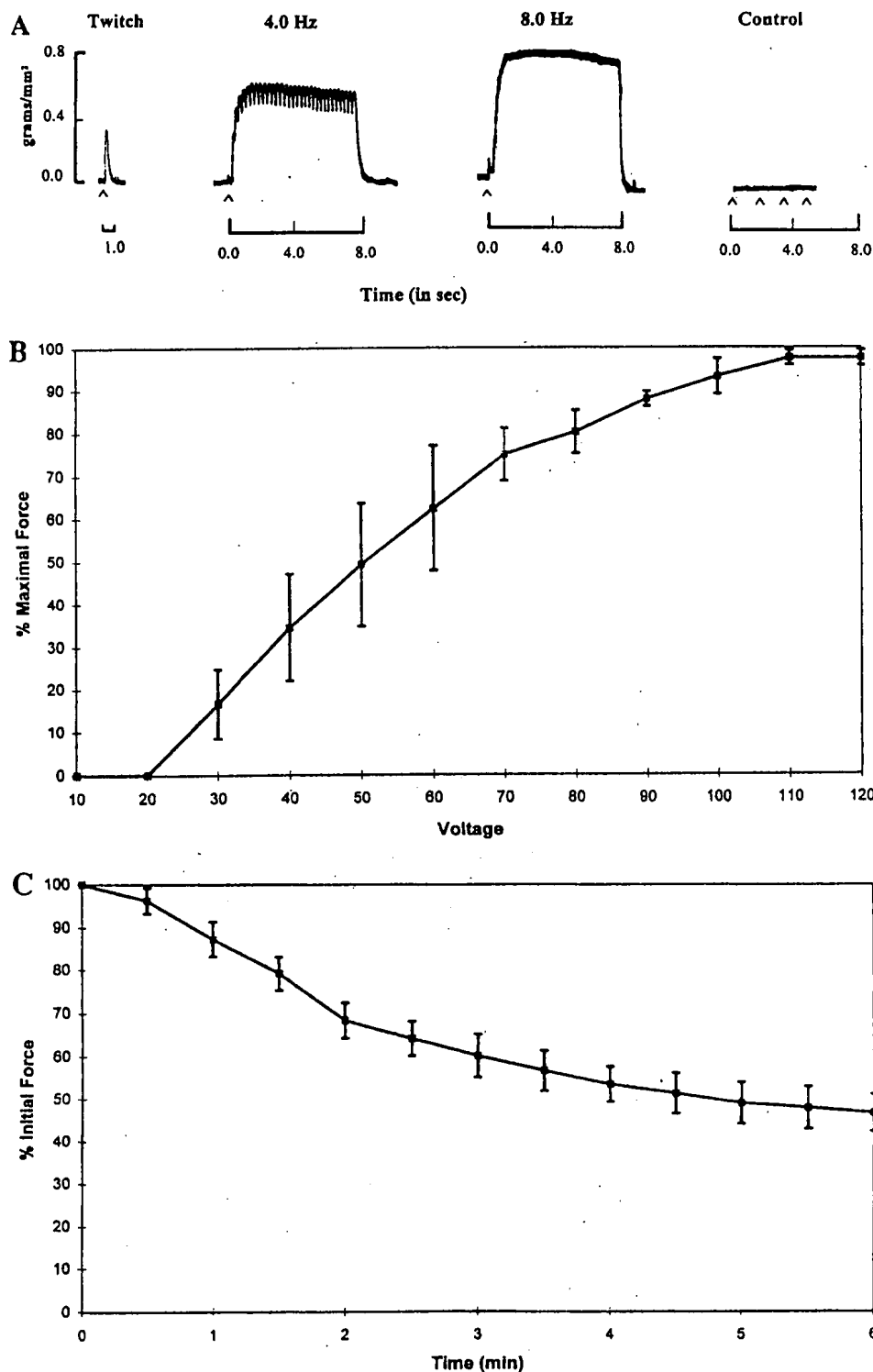


Figure 4. Contractile function of myoblast grafts *ex vivo*. Wound strips from injured hearts receiving either skeletal myoblasts or a sham saline injection were excised at 2 wk. Wounds were mounted on an isometric tension myograph in oxygenated buffer and electrically stimulated. The carats indicate the onset of electrical stimulation. Force has been normalized to cross-sectional area. (A) The first panel shows individual muscle twitch in a myoblast-injected wound. Note the rapid contraction and relaxation rates. The second panel shows that individual twitches began to superimpose with a stimulation frequency of 4 Hz, with a resulting potentiation in developed tension. The third panel shows that tetanus was induced with stimuli ≥ 7 Hz. Note the further increase in tension compared with the 4-Hz stimulation. Time to peak force in this preparation was ~ 1 s, faster than was typically observed for the overall group. The fourth panel shows that no tension was developed at any voltage in a sham-injected wound. This tracing is representative of six wound strips from three sham-injected hearts. (B) Force-voltage relationship. Developed tension for individual twitches increased as stimuli increased from 30 to 100 V, indicating recruitment of additional myofibers. Data have been normalized to maximal developed tension and are presented as mean \pm SEM of eight wound strips from six hearts. (C) Fatigue test. Wounds containing myoblast grafts were subjected to a cardiac-like duty cycle, consisting of repeated episodes of 0.33 s of tetanus/0.67 s of rest, to mimic a heart rate of 60 beats/min. There was a 53% decrease in developed tension at the end of the 6-min test. Note that most of the diminution in force occurred during the first 3 min. Data represent mean \pm SEM of seven wound strips from five hearts.

myoblasts which had proliferated and subsequently differentiated. In the day 1 grafts, proliferating cells were present within the necrotic lesion, which could have represented either graft cells or macrophages. As mentioned above, none of the cells

expressed MHC at this time, so it was not possible to determine which among these were myoblasts (versus transplanted fibroblasts or host macrophages). In the day 3 grafts, occasional BrdU-positive nuclei were identified within myosin-pos-

itive cells (Fig. 3 E). We observed a total of 12 such nuclei in three hearts. No attempt was made to quantify this low rate, but it was certainly $< 1\%$ of total nuclei in myosin-positive cells. Virtually no BrdU-positive nuclei were seen in myosin-positive cells at 1, 2, or 7 wk after transplantation (not shown). We conclude that myoblast proliferation occurs for at least 3 d after grafting, but by 1 wk virtually all cells have ceased replicating.

Contractile function of myoblast grafts. The contractile properties of 2-wk-old myoblast grafts were determined by attaching isolated wound strips to a tension myograph *ex vivo*. Virtually no spontaneous mechanical activity was detected, consistent with the paucity of cardiomyocytes histologically. Electrical stimulation caused muscle twitches in six of eight myoblast-engrafted hearts (Fig. 4 A, *first panel*); strips from the remaining two hearts may have been damaged during sample preparation, since skeletal muscle was present histologically. The grafts showed a stepwise increase in tension development as voltage was increased from 30 to ~ 100 V with a plateau thereafter (Fig. 4 B). This indicates that increasing voltage recruited additional myofibers to contract, implying that the graft myofibers are electrically insulated from one another. It should be noted that cardiac muscle does not increase contractile force with increasing voltage, since cardiocytes are coupled electrically via gap junctions.

Next, force-frequency relationships were determined. Using 120% of the voltage required for maximal tension development, the frequency of stimulation was increased incrementally from 0.5 to 10 Hz. Twitches began to superimpose at frequencies of 3–4 Hz, with a resulting increase in total developed tension (Fig. 4 A, *second panel*). Fully fused tetani were produced with 6–7 Hz stimulation (Fig. 4 A, *third panel*). Peak force during tetanus was 1.98 ± 0.45 grams (mean SEM); after normalization to cross-sectional area the peak force was 0.72 ± 0.14 grams/mm². The time to peak tetanic force averaged 2.3 ± 0.3 s, although 90% of peak force was typically generated within 1.5 s. The time to half-relaxation after tetanus was 240 ± 17 ms. It should be stressed that tetanus cannot be induced in cardiac muscle, due to the long refractory period of cardiocytes.

Finally, a fatigue test was performed to test the response of this muscle to a cardiac-like work load. The grafts were subjected to a duty cycle consisting of repeated 0.33 s of tetanic stimuli followed by 0.67 s of rest, mimicking a heart rate of 60 beats/min. The grafts showed a 32% decline in developed tension by 2 min and a 53% decline by the end of the 6-min test period (Fig. 4 C). No contractile activity could be elicited from six of seven wound strips from three injured hearts which received a sham injection of saline instead of myoblasts (Fig. 4 A, *fourth panel*). In one sham heart an adhesion had developed between the heart and chest wall, resulting in a small amount of intercostal muscle adhering to one of the two wound strips. In this preparation we detected a peak force of 0.04 grams/mm², $< 2\%$ of what was present in the myoblast-engrafted hearts.

Thus, the skeletal muscle grafts could be stimulated to contract *ex vivo* and could sustain a cardiac-like duty cycle over a 6-min test period. Furthermore, the grafts showed two physiological properties unique to skeletal muscle: recruitment of fibers with increasing voltage and the ability to sustain tetanic contraction. We do not know yet whether the grafts contract *in vivo*.

Discussion

The principal findings of this study are: (a) neonatal skeletal myoblasts can be grafted into an injured heart; (b) the engrafted myoblasts initially proliferate and then begin to form multinucleated myotubes by day 3; (c) the myotubes differentiate into mature myofibers, which initially have a phenotype similar to fast twitch muscle; (d) the myofibers develop characteristics of slow twitch muscle as the wound heals; (e) the new muscle may form satellite stem cells; and (f) the new muscle can be stimulated to contract *ex vivo*.

Strategies for muscle regeneration after myocardial injury. In principal, there are at least three strategies to induce muscle regeneration after myocardial infarction. First, the surrounding cardiac myocytes could be stimulated to migrate into the wound and proliferate to repair the defect. There is evidence that a limited amount of cell replication by adult cardiocytes occurs naturally after myocardial infarction in humans (17) and in rats (18, 19), but the response is clearly not adequate to repair the defect. The factors responsible for cell cycle arrest in cardiocytes are not well enough defined at present to begin exploring this as a therapy. (The interested reader is referred to references 20–23 for further information on this topic.)

A second strategy is to induce the cells of cardiac granulation tissue (the fibroblast-rich tissue of wound repair) to differentiate into muscle rather than forming a scar. There is not enough known about cardiac differentiation at present to attempt formation of new myocardium. However, much more is known about skeletal muscle determination. The discovery of myogenic determination genes (24, 25) has made it possible to induce a wide range of cultured cell types to differentiate into skeletal muscle. Recent studies from our group (26) and others (27) have shown that cells in cardiac granulation tissue can be induced to differentiate into skeletal muscle by transfection with the prototype myogenic determination gene, MyoD. In these early experiments, however, the frequency of muscle differentiation has been low after MyoD gene transfer. Until the frequency of myogenic conversion can be increased, it will be difficult to investigate the functional properties of the MyoD-induced skeletal muscle.

The third strategy for muscular repair of infarcts is to transplant either skeletal or cardiac myoblasts into the injured region. Studies by Koh et al. (3) and Soonpaa et al. (2) have demonstrated that fetal cardiocytes will form intercalated discs with host cardiocytes, including gap junctions and adherens junctions, when transplanted into normal hearts. No proliferation was detected in the grafted cardiocytes. Less information is available on grafting of cardiocytes into injured hearts. Our group (28) and others (29, 30) have preliminary data showing that neonatal rat or fetal human cardiocytes can be transplanted successfully into injured rat hearts. To our knowledge there is no information regarding proliferation of these grafts, nor are any functional data available. As discussed above, the principal limitation to this approach is the inability to induce cardiocytes to proliferate in culture. Until cardiocytes can replicate *in vitro*, or proliferation-competent cells can be induced reliably to differentiate into cardiocytes, cardiocyte grafting will not be feasible in humans.

In contrast to cardiocytes, proliferating skeletal muscle precursors are readily available, either as primary myoblasts in developing muscle or as satellite cells from quiescent muscle. In this study six rat pups yielded the myoblasts implanted into

27 injured hearts. In addition to their growth in culture, the myoblasts proliferated *in vivo* for several days after transplantation (Fig. 3 E). These properties have led us and several other groups to explore skeletal muscle grafting for cardiac repair. Koh et al. (7) demonstrated that the myogenic cell line C2C12 could be transplanted into the hearts of normal syngeneic mice, where the cells fused to form multinucleated myofibers. The same group also demonstrated that C2C12 cells stably transfected with a plasmid encoding active TGF- β could induce angiogenesis around the graft site (31). No coupling between the host cardiocytes and the grafted skeletal muscle was observed in either experiment.

Chiu et al. (8, 9) transplanted autologous satellite cells into cardiac freeze-thaw lesions in dogs. Comparable with our study, they also found that the grafts formed muscle cells within the healing lesion. In distinction to the current study, however, they hypothesized that their grafted skeletal muscle cells differentiated into cardiac muscle, via "milieu-dependent effects." The evidence for a cardiac phenotype was that some cells within the grafts had central rather than peripheral nuclei, and some cells contained refractile transverse structures under light microscopy interpreted to be intercalated discs. Although we observed some myofibers with persistent central nuclei in this study, as well as rare cells showing intermediate and gap junctions (Fig. 2, C and D), no intercalated discs were present by electron microscopy. More importantly, the grafted cells expressed skeletal muscle-specific proteins and failed to express the cardiac-specific isoform MHC- α up to 3 mo after transplantation. Thus, there clearly was no cardiac differentiation in this study.

Conversion of grafts from fast to slow twitch muscle. Although the skeletal muscle grafts expressed the fast fiber isoform of MHC at 1 and 2 wk, they expressed β -MHC, a marker for slow twitch fibers, at 7 wk and 3 mo. This indicates that the grafts were converting to slow twitch fibers. Conversion was apparently more rapid when the myoblasts were injected into wounds where healing had been allowed to progress for 1 wk, as opposed to immediately after injury. In the delayed transplantation model the grafts expressed β -MHC at 2 wk, while in the immediate transplantation model this protein was not detected until 7 wk. It is possible that the growth factors and cytokines present in the early wound delay myoblast differentiation and subsequent fiber type conversion.

Slow fibers exhibit several important differences from fast fibers, including a slower shortening velocity, use of oxidative phosphorylation for ATP production, a higher mitochondrial content, a higher myoglobin content, and a much greater resistance to fatigue (16, 32). An interesting parallel is that the latissimus dorsi muscle also undergoes fiber type switching when it is conditioned for dynamic cardiomyoplasty. Cardiomyoplasty is an experimental therapy for heart failure, where skeletal muscle is wrapped around the heart to serve as a ventricular assist device (33). Untrained latissimus dorsi is a mixed fiber type muscle which fatigues rapidly with repeated stimulation. When conditioned by repeated electrical stimulation for 6 wk before surgery, however, the latissimus dorsi converts entirely to slow twitch fibers and becomes fatigue resistant (6). Only the conditioned, slow twitch muscle is able to assist cardiac function. This parallel suggests the intriguing possibility that repeated electromechanical stimulation leads to activation of the slow fiber phenotype. Since we did not test whether the environment of the heart contributed to fiber type conversion,

additional experiments will be required to determine the mechanism. The fact that the grafts differentiated into slow twitch fibers suggests that they may be suited to perform a cardiac type work load.

Will skeletal muscle transplantation augment cardiac function? This study definitively showed that myoblast grafting can generate new contractile tissue. The skeletal muscle grafts exhibited characteristic twitches when stimulated *ex vivo* (Fig. 4 A) and showed recruitment of contractile units with increasing voltage (Fig. 4 B). Furthermore, tetanus could be induced with rapid stimulation (Fig. 4 A, *second* and *third* panels), and the grafts could perform a cardiac-like duty cycle for 6 min (Fig. 4 C). Peak force during tetanus averaged 0.7 ± 0.1 grams/mm². Since the wound strips contained < 50% of the myofiber content of normal muscle, due to inclusion of scar tissue, the force can be normalized to at least 1.4 grams/mm² muscle. Adult mammalian muscle can generate 15–35 grams/mm² force at tetanus, depending on fiber type (14, 34). Thus, the 2-wk grafts generated ~ 4–10% of the predicted force for mature skeletal muscle. Several factors may cause a lower than predicted force, including the relative immaturity of the 2-wk myofibers, stretching of the immature extracellular matrix, poor cell matrix attachments, or misalignment of some fibers relative to the axis of the wound strip.

Although preliminary, these results are encouraging and suggest that more detailed studies of contractile function are warranted in skeletal myoblast-engrafted hearts. A critical question is whether the skeletal muscle grafts contract *in vivo*. To provide coordinated mechanical assistance, the grafted cells ideally should form electrical and mechanical junctions with the host myocardium. In our grafts the skeletal muscle cells were insulated from the remaining myocardium by scar tissue, so there was no opportunity for myofiber–cardiocyte coupling to occur. Koh et al. (7) transplanted C2C12 myoblasts into normal mouse hearts and observed no cell junctions between grafted myofibers and host cardiocytes by electron microscopy. Although proliferating myoblasts have been reported to synthesize both gap junction proteins (35) and N-cadherin (36, 37), these proteins are typically absent from adult skeletal myofibers. By electron microscopy we observed evidence both for intermediate and gap junction formation between skeletal myofibers 2 wk after grafting (Fig. 2, C and D). This finding was infrequent, however, and it is unknown whether such junctions would persist in longer term grafts. If skeletal muscle will not couple spontaneously with cardiac muscle, it is possible that such junctions could be induced by stably transfecting skeletal muscle cells with genes for cardiac junctional proteins. Another possibility is that skeletal muscle grafts could be electrically paced in synchrony with the cardiac cycle. Pacing would require sufficient voltage to activate all of the fibers, and currently it is unknown whether this would have a deleterious effect on the surrounding myocardium.

In the uninjured heart there is a complex fiber geometry, where the outer fibers run in the long axis, the midwall fibers run in the short axis, and the inner fibers again run in the long axis. This geometry is established during embryogenesis and is thought to be important for mechanical efficiency. In this study, the grafted myofibers were predominantly aligned with the short (transverse) axis of the heart. Alignment was noted as early as day 3, when myotube formation was prominent (Fig. 1 C). This is the same orientation that wound fibroblasts and collagen fibers acquire during wound healing, and it seems

likely that all are aligned by local mechanical forces. It is not known whether alignment with the heart's short axis will influence the ability of these myofibers to restore mechanical function after injury.

There are two aspects of skeletal muscle which theoretically could make it superior to cardiac muscle for infarct repair. First, skeletal muscle is much more resistant to ischemia than cardiac muscle. Skeletal muscle can withstand many hours of severe ischemia without becoming irreversibly injured, whereas in myocardium irreversible injury begins within 20 min (38). A second difference is that skeletal myoblast grafts might establish satellite cells. Satellite cells are the resident stem cells in skeletal muscle and proliferate in response to injury. Once activated, satellite cells can fuse with damaged myofibers or establish new myofibers to replace those lost to necrosis. We observed cells within 2-wk grafts which were morphologically consistent with satellite cells by electron microscopy (Fig. 3, E and F). Thus, it is possible that infarcts repaired with skeletal myoblasts might become more resistant to a subsequent episode of ischemia or might be able to replace myofibers damaged by ischemia.

Acknowledgments

The authors are grateful for the adroit technical skills of Trudy Bartosek for performing all the immunocytochemistry, Cathy Gipay for assistance with animal surgery, DeeAnn Gregory for assistance with myoblast culture, and Rene Collman for assistance with electron microscopy. The β -MHC antibody was a gift from Dr. Jeff Miller, Massachusetts General Hospital, Boston, MA. We thank Kelly Hudkins for her helpful advice for double immunostaining.

This study was supported in part by a Grant-in-Aid from the American Heart Association (C.E. Murry), and National Institutes of Health grants HL-03174 (S.M. Schwartz and S.D. Hauschka), HL-26405 (S.M. Schwartz), AR-18860 (S.D. Hauschka), and AR-41973 (R.W. Wiseman). Dr. Murry is the recipient of a Burroughs Wellcome Career Award in the Biomedical Sciences.

References

- Reimer, K.A., and R.B. Jennings. 1979. The "wavefront phenomenon" of myocardial ischemic cell death. II. Transmural progression of necrosis within the framework of ischemic bed size (myocardium at risk) and collateral blood flow. *Lab. Invest.* 40:633-644.
- Soonpaa, M.H., G.Y. Koh, M.G. Klug, and L.J. Field. 1994. Formation of nascent intercalated disks between grafted fetal cardiomyocytes and host myocardium. *Science (Wash. DC)* 264:98-101.
- Koh, G.Y., M.H. Soonpaa, M.G. Klug, H.P. Pride, B.J. Cooper, D.P. Zipes, and L.J. Field. 1995. Stable fetal cardiomyocyte grafts in the hearts of dystrophic mice and dogs. *J. Clin. Invest.* 96:2034-2042.
- Hauschka, S.D. 1974. Clonal analysis of vertebrate myogenesis. II. Environmental influences upon human muscle differentiation. *Dev. Biol.* 37:329-344.
- Hauschka, S.D. 1982. Muscle cell culture: future goals for facilitating the investigation of human muscle disease. In *Disorders of the Motor Unit*. D.L. Schotland, editor. J. Wiley & Sons, New York. 925-936.
- Mannion, J.D., T. Bitto, R.L. Hammond, N.A. Rubenstein, and L.W. Stephenson. 1986. Histochemical and fatigue characteristics of conditioned canine latissimus dorsi muscle. *Circ. Res.* 58:298-304.
- Koh, G.Y., M.G. Klug, M.H. Soonpaa, and L.J. Field. 1993. Differentiation and long-term survival of C2C12 myoblast grafts in heart. *J. Clin. Invest.* 92:1548-1554.
- Zibaitis, A., D. Greentree, F. Ma, D. Marelli, M. Duong, and R.C. Chiu. 1994. Myocardial regeneration with satellite cell implantation. *Transplant. Proc.* 26:3294.
- Chiu, R.C., A. Zibaitis, and R.L. Kao. 1995. Cellular cardiomyoplasty: myocardial regeneration with satellite cell implantation. *Ann. Thorac. Surg.* 60:12-18.
- Clegg, C.H., and S.D. Hauschka. 1987. Heterokaryon analysis of muscle differentiation: regulation of the postmitotic state. *J. Cell Biol.* 105:937-947.
- Vracko, R., and D. Thorning. 1991. Contractile cells in rat myocardial scar tissue. *Lab. Invest.* 65:214-227.
- Vracko, R., D. Thorning, and R.G. Frederickson. 1990. Fate of nerve fibers in necrotic, healing and healed rat myocardium. *Lab. Invest.* 63:490-501.
- Vracko, R., D. Thorning, R.G. Frederickson, and D. Cunningham. 1988. Myocyte reactions at the borders of injured and healing rat myocardium. *Lab. Invest.* 59:104-114.
- Wiseman, R.W., T.W. Beck, and P.B. Chase. 1996. The effect of intracellular pH on force development depends on temperature in intact mouse skeletal muscle. *Am. J. Physiol.* 40:C878-C886.
- Trupin, G.L., L. Hsu, and Y.-H. Hsieh. 1979. Satellite cell mimics in regenerating skeletal muscle. In *Muscle Regeneration*. A. Mauro, editor. Raven Press, New York. 101-114.
- Kushmerick, M.J., T.S. Moerland, and R.W. Wiseman. 1992. Mammalian skeletal muscle fibers distinguished by contents of phosphocreatine, ATP, and Pi. *Proc. Natl. Acad. Sci. USA* 89:7521-7525.
- Quaini, F., E. Cigola, C. Lagrasta, G. Saccani, E. Quaini, C. Rossi, G. Olivetti, and P. Anversa. 1994. End-stage cardiac failure in humans is coupled with the induction of proliferating cell nuclear antigen and nuclear mitotic division in ventricular myocytes. *Circ. Res.* 75:1050-1063.
- Rumyantsev, P.P. 1979. Some comparative aspects of myocardial regeneration. In *Muscle Regeneration*. A. Mauro, editor. Raven Press, New York. 335-355.
- Reiss, K., J. Kajstura, J.M. Capasso, T.A. Marino, and P. Anversa. 1993. Impairment of myocyte contractility following coronary artery narrowing is associated with activation of the myocyte IGF1 autocrine system, enhanced expression of late growth related genes, DNA synthesis and myocyte nuclear mitotic division in rats. *Exp. Cell Res.* 207:348-360.
- Tam, S.K.C., W. Gu, V. Mahdavi, and B. Nadal-Ginard. 1995. Cardiac myocyte terminal differentiation. *Ann. NY Acad. Sci.* 752:72-79.
- Gu, W., J.W. Schneider, G. Condorelli, S. Kaushal, V. Mahdavi, and B. Nadal-Ginard. 1993. Interaction of myogenic factors and the retinoblastoma protein mediates muscle cell commitment and differentiation. *Cell* 72:309-324.
- Yoshizumi, M., W.S. Lee, C.M. Hsieh, J.C. Tsai, J. Li, M.A. Perrella, C. Patterson, W.A. Endege, R. Schlegel, and M.E. Lee. 1995. Disappearance of cyclin A correlates with permanent withdrawal of cardiomyocytes from the cell cycle in human and rat hearts. *J. Clin. Invest.* 95:2275-2280.
- Kim, K.K., M.H. Soonpaa, A.I. Daud, G.Y. Koh, J.S. Kim, and L.J. Field. 1994. Tumor suppressor gene expression during normal and pathologic myocardial growth. *J. Biol. Chem.* 269:22607-22613.
- Weintraub, H., R. Davis, S. Tapscott, M. Thayer, M. Krause, R. Ben-ezra, T.K. Blackwell, D. Turner, R. Rupp, S. Hollenberg, et al. 1991. The MyoD gene family: nodal point during specification of the muscle cell lineage. *Science (Wash. DC)* 251:761-766.
- Olson, E.N., and W.H. Klein. 1994. bHLH factors in muscle development: dead lines and commitments, what to leave in and what to leave out. *Genes Dev.* 8:1-8.
- Murry, C.E., M.A. Kay, T. Bartosek, S.D. Hauschka, and S.M. Schwartz. 1996. Muscle differentiation during repair of myocardial necrosis in rats via gene transfer with MyoD. *J. Clin. Invest.* 98:2209-2217.
- Prentice, H., R.A. Kloner, V. Sartorelli, S.D. Bellows, K. Alker, and L. Kedes. 1993. Transformation of cardiac fibroblasts into the skeletal muscle phenotype by injection of a MyoD-expressing retrovirus into ischemic heart. *Circulation* 88(Suppl.):1475a. (Abstr.)
- Murry, C.E., C. Fabre-Suver, S.D. Hauschka, and S.M. Schwartz. 1995. Skeletal and cardiac myoblast transplantation after myocardial necrosis: possible routes to muscle regeneration. *Circulation* 92:112a. (Abstr.)
- Leor, R., M. Patterson, M.J. Quinones, L. Kedes, and R.A. Kloner. 1995. Successful transplantation of fetal cardiomyoblasts into the infarcted myocardium of rat: a potential method for repair of infarcted myocardium. *Circulation* 92(Suppl.):150a. (Abstr.)
- Scorcin, M., F. Marotte, A. Sabri, O. Le Dref, M. Demirag, J.-L. Samuel, L. Rappaport, and P. Menasche. 1995. Can grafted cardiomyocytes colonize peri-infarction myocardial areas? *Circulation* 92(Suppl.):150a. (Abstr.)
- Koh, G.Y., S.J. Kim, M.G. Klug, K. Park, M.H. Soonpaa, and L.J. Field. 1995. Targeted expression of transforming growth factor-beta 1 in intracardiac grafts promotes vascular endothelial cell DNA synthesis. *J. Clin. Invest.* 95:114-121.
- Kelly, A.M., and N.A. Rubinstein. 1994. The diversity of muscle fiber types and its origin during development. In *Myology*. A.G. Engel and C. Franzini-Armstrong, editors. McGraw-Hill, New York. 119-133.
- Ninami, H., A. Pochettino, and L.W. Stephenson. 1991. Use of skeletal muscle grafts for cardiac assist. *Trends Cardiovasc. Med.* 1:122-126.
- Phillips, S.K., R.W. Wiseman, R.C. Woleged, and M.J. Kushmerick. 1993. The effects of metabolic fuel on force production and resting inorganic phosphate levels in mouse skeletal muscle. *J. Physiol. (Lond.)* 462:135-146.
- Balogh, S., C.C.G. Naus, and P.A. Merrifield. 1993. Expression of gap junctions in cultured rat L6 cells during myogenesis. *Dev. Biol.* 155:351-360.
- Cifuentes, D.C., M. Nicolet, D. Goudou, F. Reiger, and R.M. Mege. 1993. N-cadherin and N-CAM-mediated adhesion in development and regeneration of skeletal muscle. *Neuromuscul. Disord.* 3:361-365.
- Mege, R.M., D. Goudou, D. Diaz, M. Nicolet, L. Garcia, G. Geraud,

and F. Rieger. 1992. N-cadherin and N-CAM in myoblast fusion: compared localisation and effect of blockade by peptides and antibodies. *J. Cell Sci.* 103: 897-906.

38. Jennings, R.B., and K.A. Reimer. 1981. Lethal myocardial ischemic injury. *Am. J. Pathol.* 102:241-255.

39. Bader, D., T. Masaki, and D.A. Fischman. 1982. Immunochemical analysis of myosin heavy chain during avian myogenesis in vivo and in vitro. *J. Cell Biol.* 95:763-770.

40. Havenith, M.G., R. Visser, J.M.C. Schrijvers-van Schendel, and F.T. Bosman. 1990. Muscle fiber typing in routinely processed skeletal muscle with monoclonal antibodies. *Histochemistry.* 93:497-499.

41. Rudnicki, M.A., G. Jackowski, L. Saggin, and M.W. McBurney. 1990. Actin and myosin expression during development of cardiac muscle from cul-

tured embryonal carcinoma cells. *Dev. Biol.* 138:348-358.

42. Silberstein, L., S.G. Webster, M. Travis, and H.M. Blau. 1986. Developmental progression of myosin gene expression in cultured muscle cells. *Cell.* 46: 1075-1081.

43. Hughes, S.M., M. Cho, I. Karsch-Mizrachi, M. Travis, L. Silberstein, L.A. Leinwand, and H.M. Blau. 1993. Three slow myosin heavy chains sequentially expressed in developing mammalian skeletal muscle. *Dev. Biol.* 158:183-199.

44. Van Kleef, E.M., J.F.M. Smits, J.G.R. DeMey, J.P.M. Cleutjens, D.M. Lombardi, S.M. Schwartz, and M.J.A.P. Daemen. 1992. Alpha-1-adrenoreceptor blockade reduces the angiotensin II-induced vascular smooth muscle cell DNA synthesis in the rat thoracic aorta and carotid artery. *Circ. Res.* 70:1122-1127.

EVIDENCE APPENDIX

Item 5

UNITED STATES PATENT AND TRADEMARK OFFICE
CERTIFICATE OF CORRECTION

PATENT NO. : 5,328,470

Page 1 of 2

DATED : July 12, 1994

INVENTOR(S) : Elizabeth G. Nabel et al.

It is certified that error appears in the above-identified patent and that said Letters Patent is hereby corrected as shown below:

Column 1: line 25, "would an asset"
should read --would be an asset--.

Column 7: line 17, "cells though"
should read --cells through--.

Column 21: line 39, "DNA/liposomes comples"
should read --DNA/liposomes complex--;

line 41, "occasioinally"
should read --occasionally--;

line 43, "ogran"
should read --organ--;

line 68, "injuection"
should read --injection--.

Column 22: line 27, "plasmnid"
should read --plasmid--.

UNITED STATES PATENT AND TRADEMARK OFFICE
CERTIFICATE OF CORRECTION

PATENT NO. : 5,328,470

Page 2 of 2

DATED : July 12, 1994

INVENTOR(S) : Elizabeth G. Nabel et al.

It is certified that error appears in the above-identified patent and that said Letters Patent is hereby corrected as shown below:

Column 23: line 65, "incrase"
should read --increase--.

Column 24: line 5, "ande"
should read --and--.

Column 25: line 25, "that. within"
should read --that within--.

Signed and Sealed this
Twenty-sixth Day of March, 1996



Attest:

BRUCE LEHMAN

Commissioner of Patents and Trademarks

Attesting Officer

EVIDENCE APPENDIX

Item 6

FOCUS ISSUE: CARDIAC REGENERATION

Regeneration of Human Infarcted Heart Muscle by Intracoronary Autologous Bone Marrow Cell Transplantation in Chronic Coronary Artery Disease

The IACT Study

Bodo E. Strauer, MD,* Michael Brehm, MD,* Tobias Zeus, MD,* Thomas Bartsch, MD,* Christina Schannwell, MD,* Christine Antke, MD,† Rüdiger V. Sorg, PhD,‡ Gesine Kögler, PhD,‡ Peter Wernet, MD,‡ Hans-Wilhelm Müller, MD,† Matthias Köstering, MD*

Düsseldorf, Germany

OBJECTIVES	Stem cell therapy may be useful in chronic myocardial infarction (MI); this is conceivable, but not yet demonstrated in humans.
BACKGROUND	After acute MI, bone marrow-derived cells improve cardiac function.
METHODS	We treated 18 consecutive patients with chronic MI (5 months to 8.5 years old) by the intracoronary transplantation of autologous bone marrow mononuclear cells and compared them with a representative control group without cell therapy.
RESULTS	After three months, in the transplantation group, infarct size was reduced by 30% and global left ventricular ejection fraction (+15%) and infarction wall movement velocity (+57%) increased significantly, whereas in the control group no significant changes were observed in infarct size, left ventricular ejection fraction, or wall movement velocity of infarcted area. Percutaneous transluminal coronary angioplasty alone had no effect on left ventricular function. After bone marrow cell transplantation, there was an improvement of maximum oxygen uptake (VO_{2max} +11%) and of regional ^{18}F -fluor-desoxy-glucose uptake into infarct tissue (+15%).
CONCLUSIONS	These results demonstrate that functional and metabolic regeneration of infarcted and chronically avital tissue can be realized in humans by bone marrow mononuclear cell transplantation. (J Am Coll Cardiol 2005;46:1651-8) © 2005 by the American College of Cardiology Foundation

Cardiac performance after myocardial infarction (MI) is compromised by ventricular remodeling, which represents a major cause of late infarct-related chronic heart failure and death (1,2). Although conventional drug therapy (e.g., with beta-receptor blockers and/or angiotensin-converting enzyme inhibitors) may delay remodeling, there is no basic

See page 1659

therapeutic regimen available for preventing or even reversing this process. By the use of interventional therapeutics (percutaneous transluminal coronary angioplasty [PTCA], stent), recanalization of the occluded infarct-related artery is possible, thereby improving or normalizing coronary blood flow. However, despite sufficient reperfusion of infarcted tissue, the viability of the infarcted myocardium cannot, or can only insufficiently, be improved in most of these patients (3). Therefore, catheter-based therapy of acute MI is useful for vascular recanalization, but the second and crucial step,

the regeneration of necrotic heart muscle, is not realized by this vascular procedure alone.

Experimental (4) and clinical (5,6) studies have shown recently for the first time that bone marrow mononuclear cells (BMCs) may regenerate damaged myocardium in acute MI in humans. Because the regenerative potential of bone marrow-derived cells ought also to be expected to exist in chronically ischemic heart disease as well (7-12), we have assembled in an ongoing clinical investigation 18 patients with chronic MI to prove this new therapeutic possibility.

METHODS

Study population. All 18 patients (49 ± 11 years) were men and were recruited consecutively from January 2003 until March 2004. They had had transmural MI 27 ± 31 months before, at which point all infarcts had been treated acutely by PTCA and/or stent implantation (Table 1, Fig. 1).

The inclusion criteria were age <70 years, one-vessel disease with an open infarct-related artery at the time of stem cell therapy, sinus rhythm, a clear-cut demarcation of the ventriculographic infarct area, and no coronary bypass surgery. General exclusion criteria were severe comorbidity and alcohol or drug dependency. Although chronically infarcted myocardium usually does not regenerate sponta-

From the *Department of Internal Medicine, Division of Cardiology, Pneumology and Angiology; †Department of Nuclear Medicine; and ‡Institute for Transplantation Diagnostics and Cell Therapeutics, Heinrich-Heine-University, Düsseldorf, Germany.

Manuscript received October 31, 2004; revised manuscript received December 13, 2004; accepted January 25, 2005.

Abbreviations and Acronyms

BMC	= bone marrow mononuclear cell
CPK	= creatine phosphokinase
ECG	= electrocardiogram
LV	= left ventricular
MI	= myocardial infarction
PET	= positron emission tomography
PTCA	= percutaneous transluminal coronary angioplasty
Tx group	= transplantation group

neously, for comparison a control group, parallel to the recruitment of the stem cell transplantation group (Tx group), was recruited and analyzed, meeting the same inclusion criteria as the stem-cell group. The recruitment of patients was performed according to a randomization procedure in which all patients of the entire chronic infarction group were distributed to the treatment group, where they agreed with all the therapeutic regimen. Alternatively, all patients of the chronic infarction group who refused the therapeutic regimen (bone marrow puncture and aspiration, intracoronary cell transplantation, and another cardiac catheterization) were allocated to the control group. All medications with angiotensin-converting enzyme inhibitors and with beta receptor blockers were maintained constant during the study period.

The cell-treated patients had stable ventricular dynamics for infarct size, ejection fraction, and wall movement velocity of infarcted area at least 9 ± 6 months before cell transplantation. Infarct size at the time of cell therapy showed an amount of $27 \pm 8\%$ of the circumference of the left ventricle (LV), determined by ventriculography.

Preparation of BMCs. One day before cell therapy, bone marrow was taken (80 ml from the iliac crest) and mono-

nuclear cells were isolated and identified including CD34-positive cells, AC133-positive cells and CD45/CD14 negative cells (6). The cells were isolated under good manufacturing practice conditions by Ficoll density separation on Lymphocyte Separation Medium (Bio Whittaker, Walkersville, Maryland), before the residual erythrocytes were lysed with H_2O . For overnight cultivation, 1×10^6 BMCs/ml were placed in Teflon bags (Vuelife, Cell Genix, Gaithersburg, Maryland) and cultivated in X-Vivo 15 Medium (Bio Whittaker) supplemented with 2% heat-inactivated autologous plasma. The next day, BMCs were harvested and washed three times with heparinized saline before final resuspension in heparinized saline. Viability was $93 \pm 3\%$. Heparinization and filtration (cell strainer, FALCON) was carried out to prevent cell clotting and microembolization during intracoronary transplantation. These cells were used for therapy. All microbiologic tests of the clinically used cell preparations proved negative. All patients received extensive information about the procedure, which was approved by the ethical committee of our university, and all gave written informed consent.

Administration of BMCs. Following assessment of baseline examinations (coronary angiography, left ventriculography, spiroergometry, ^{99m}Tc -tetrafosmin single-photon emission computed tomography (SPECT) and ^{18}F -fluor-deoxy-glucose (^{18}F -FDG) positron emission tomography (PET), cell transplantation was performed via the intracoronary administration route (6,13) using four to six fractional infusions parallel to balloon inflation over 2 to 4 min of 3 to 5 ml of cell suspension, each containing 15 to 22×10^6 mononuclear cells. All cells were infused directly into the infarcted zone through the infarct-related artery via an angioplasty balloon catheter, which was inflated at a low pressure (2 to 4 atm) and was located within

Table 1. Demographic Data of Intracoronary Bone Marrow Stem Cell Transplantation Group and Control Group

Characteristics	Tx Group	Control Group	p
No. of patients	18	18	
Age, yrs	49 ± 11	52 ± 10	NS
Transmural myocardial infarction, months before Tx	27 ± 31	30 ± 34	NS
Coronary angiography			
LAD/LCX/RCA as affected vessel	16/0/2	10/3/5	
No. of patients with stent implantation	16	17	NS
Risk factors			
Diabetes mellitus, %	16	11	NS
Positive family history, %	44	33	NS
Smoker and ex-smoker, %	67	56	NS
Hyperlipoproteinemia, %	89	94	NS
Medication			
Beta-blocker, %	94	89	NS
Angiotensin-converting enzyme inhibitor, %	94	89	NS
Statin, %	94	100	NS
Laboratory parameters			
CPK, U/l	$1,504 \pm 979$	$1,489 \pm 952$	NS
Bone marrow mononuclear cells, n ($10^6 \times$)	90		

Values are mean \pm SD or number of patients.

CPK = creatine phosphokinase; LAD = left anterior descending coronary artery; LCX = left circumflex coronary artery; RCA = right coronary artery; Tx = intracoronary bone marrow stem cell transplantation.

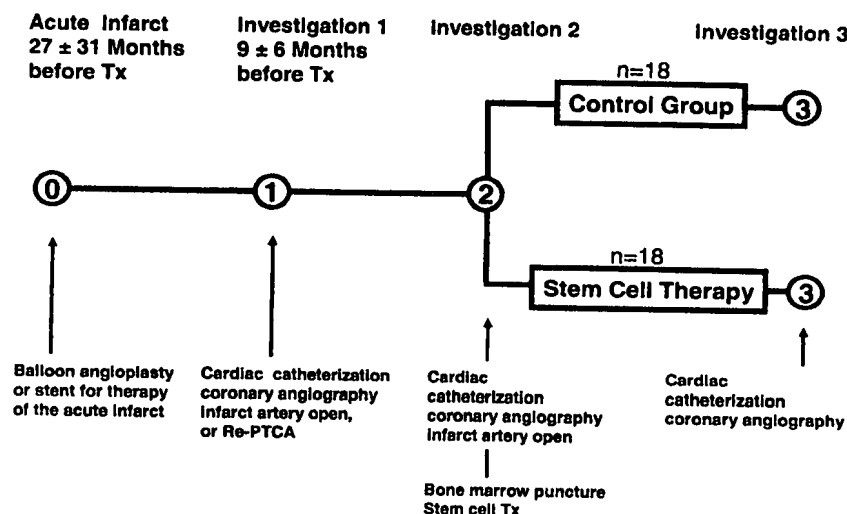


Figure 1. Diagrammatic representation of the algorithm of intracoronary stem cell therapy (Tx) in chronic ischemic heart disease after myocardial infarction. The infarcts occurred 27 ± 31 months before Tx. All infarct patients were treated with percutaneous transluminal coronary angioplasty (PTCA) or with stent implantation. 9 ± 6 months before (investigation 1) coronary angiography (including quantitative left ventriculography) was performed. If re-stenosis was present, re-PTCA was made. Investigation 2 embraces all patients for the evaluation of coronary morphology after PTCA/stent. Only patients with an open infarct-related artery were included in both groups. Patients who agreed to Tx received within 10 days after investigation 2 bone marrow punctures and Tx by the intracoronary administration route and had altogether five invasive investigations, including two for therapeutic reasons (nos. 0 and 1). Patients who were not eligible for Tx (disagreement with bone marrow puncture and with subsequent Tx) served as a control group. Investigation 3 represents all follow-up measurements 3 months after Tx (Tx patients) or after investigation 2 for control group patients.

the previously stented coronary segments. This prevented backflow of cells and produced stop flow beyond the site of balloon inflation to facilitate high-pressure infiltration of cells into the infarcted zone. Prolonged contact time for cellular migration was also enabled. Three months after catheter-guided cell transplantation, all functional tests were repeated, including coronary angiography and left ventriculography. There were no procedural or cell-induced complications, and there were no side effects in any patient.

Spiroergometry. Aerobic exercise capacity was examined before (<10 days) intracoronary cell transplantation and three months later during follow-up. All patients ($n = 18$) were subjected to initial bicycle spiroergometry to assess their functional fitness and to determine the limit of safe intensity of exercise. We chose a protocol with an intensified workload up to the symptom-limited maximum (basic load of 50 W, intensification at 25 W, 2-min duration of each workload step). We determined the anaerobic threshold for prescribing a suitable load intensity. During the whole spiroergometry, monitoring by a 12-lead electrocardiogram (ECG) was carried out. The exercise capacity was assessed on the basis of maximum load levels expressed in watts (W_{max}) and maximum peak oxygen uptake (VO_{2max}).

Coronary angiography and left ventriculography. Coronary angiography and biplane left ventriculography were performed 9 ± 6 months before cell transplantation and also a second time, within 10 days, immediately before cell therapy. The therapeutic follow-up was three months after the treatment. Thus, stable baseline conditions were documented (coronary vessel involvement, ventricular function, and geometry). Cardiac function was evaluated by left

ventricular (LV) ejection fraction and by auxotonic myocardial contractility index, evaluated by the wall movement velocity of the infarcted area. The infarct size was calculated according to the method of Sheehan (14) by plotting five axes perpendicular to the long axis of the heart in the main akinetic or dyskinetic segment of the ventricular wall. Systolic and diastolic lengths were then measured by two independent observers, and the mean difference was divided by the systolic duration in seconds.

Quantification of coronary stenosis (restenosis). Cinecoronangiograms were obtained during stem cell transplantation and at three months thereafter according to standard acquisition guidelines. The angiograms were evaluated by two independent observers and quantitative analysis was performed (15). Standard morphologic criteria were used to characterize the complexity of baseline lesions. The user-defined reference diameter proximal to the stenosis and the minimal luminal diameter within the culprit of the stenosis were used to calculate the percentage of stenosis. A value of 0 mm was assigned for the minimal luminal diameter in case of total occlusion at baseline or follow-up. Restenosis was defined as $\geq 50\%$ stenosis of the initial target lesion at follow-up. Calculations of restenosis were performed in both groups, with and without stem cell therapy, in the same way, thus enabling evaluation the differential effects of PTCA-guided cell therapy and of PTCA effects alone.

Ventricular function after PTCA in the control group. For the evaluation of a potential effect on the PTCA intervention itself on LV function, all patients in the control group were analyzed with regard to infarct size, ejection fraction, and infarction wall movement velocity.

Table 2. Single Values of Intracoronary Bone Marrow Stem Cell Transplantation Group

Patient Number	Area of Infarction, %*			LV Ejection Fraction, %*			Infarction Wall Movement Velocity, cm/s*		
	Investigation 1 9 ± 6 Mo Before Tx	Investigation 2 <10 Days Before Tx	Investigation 3 3 Mo After Tx	Investigation 1 9 ± 6 Mo Before Tx	Investigation 2 <10 Days Before Tx	Investigation 3 3 Mo After Tx	Investigation 1 9 ± 6 Mo Before Tx	Investigation 2 <10 Days Before Tx	Investigation 3 3 Mo After Tx
1	26	26	22	56	55	60	0.88	0.77	0.82
2	28	29	26	45	43	49	2.06	1.88	2.13
3	16	16	5	64	65	71	1.45	1.50	2.10
4	27	25	14	48	50	65	1.20	1.25	2.88
5	16	14	11	66	69	71	2.25	2.77	3.75
6	16	13	6	64	66	73	1.50	1.77	2.55
7	15	18	11	57	55	63	2.78	2.65	3.13
8	28	28	20	43	44	49	3.15	3.25	4.25
9	27	27	11	46	46	64	1.61	1.65	3.30
10	20	17	14	56	58	62	2.21	2.45	3.13
11	28	25	17	42	38	52	1.91	1.88	3.00
12	33	28	21	44	47	54	2.28	2.62	3.50
13	39	37	27	50	51	59	1.25	2.50	4.90
14	29	33	27	62	62	61	1.20	1.33	2.70
15	37	37	31	48	43	53	1.83	1.56	2.50
16	29	29	24	53	54	58	1.25	1.06	3.06
17		41	35		48	55		1.66	3.00
18		35	25		45	53		0.94	1.94
Mean	26	27	19	53	52	60	1.80	1.86	2.92
SD	7	8	9	8	9	7	0.63	0.70	0.91

*Calculated from left ventriculography.

LV = left ventricular; Mo = Months; other abbreviations as in Table 1.

Nuclear cardiologic investigations (PET and SPECT). ^{18}F -FDG-positron emission tomography (^{18}F -FDG PET) was performed with a Scanditronix SCX 4096 WB-Scanner (FWHM = 6 mm transaxial, axial field of view = 4.6 cm). Patients received an oral glucose load of 1 g/kg body weight 80 ± 30 min before the intravenous application of ^{18}F -FDG (380 ± 60 MBq). The ^{18}F -FDG was administered at the time of decrease of blood glucose level <130 mg/dl. An initial transmission scan was obtained using a ^{68}Ga -filled pin source to correct the subsequent emission scans for attenuation. The data acquisition was started 45 min after administration of FDG. Image data were recorded with a 256×256 matrix in 3 consecutive bed positions over 15 min per position. The data were reconstructed backprojected with a Hanning filter (5 mm).

$^{99\text{m}}\text{Tc}$ -tetrofosmin SPECT. Sixty minutes after intravenous injection of 600 ± 140 MBq of the perfusion-marker $^{99\text{m}}\text{Tc}$ -tetrofosmin under a "rest" condition, the images were obtained using a SPECT scanner with double-head detector (PRISM 2000, Marconi/Phillips), a low-energy, high-resolution collimator, and a 128×128 matrix. Image data were collected over 360° at 3° every 30 s. The images were reconstructed backprojected with a low-pass filter (order 12, cutoff 0.2).

PET and SPECT evaluation. Normalized values for FDG uptake and perfusion were calculated by comparing regional with maximum tracer uptake on the reconstructed images. We performed a regional analysis of glucose metabolism and perfusion using a set of standardized, individually adjusted circular regions of interest (diameter 18.06 mm, surface 256 mm^2). The reconstructed metabolic and perfusion images were realigned for each patient (MPI-Tool, version 3.0; Advanced Tomo Vision, Erftstadt, Germany) and were resliced according to cardiac axis (short-axis and horizontal and vertical long-axis views). The regions were positioned immediately neighboring, with no overlap, according to an overlay of the co-registered metabolic and perfusion images. The regions covered the infarct lesion as well as normal myocardium. In this way, we generated templates of regions for each patient, which could be used for the evaluation of metabolism and perfusion, before and after BMC transplantation without further modification. According to Segall et al. (16), regions with a normalized FDG uptake $<50\%$ were rated as transmural scar and regions with an uptake of 50% to 60% as non-transmural scar.

Further analysis was restricted to regions with FDG uptake $<60\%$ in the PET scans, pursuant to our intention to focus on the effects of BMC transplantation on scar tissue.

Safety parameters. To assess any inflammatory response and myocardial reaction after cell therapy, white blood cell count, the serum levels of C-reactive protein (CRP) and of creatine phosphokinase (CPK) were determined immediately before as well as after treatment. Additional analysis was done directly after transplantation and three months later: ECG at rest, 24-h Holter ECG, and echocardiography.

Statistical analysis. All data are presented as mean \pm SD. Statistical significance was accepted when $p < 0.05$. Intra-individual comparison of variables of investigation 1 (9 ± 6 months before cell transplantation for Tx group, 9 ± 5 months before investigation 2 for control patients) and investigation 2 (<10 days before cell transplantation for Tx group, no transplantation for control patients) and of variables of investigation 2 and follow-up investigation 3 (3 months after cell therapy for Tx group, 8 ± 5 months after investigation 2 for control patients) was performed using Wilcoxon rank-sum test. The missing values (Table 2) were omitted and not calculated for statistical analysis. The p values (by analysis of variance) have been given for LV ejection fraction, area of infarction, and infarction wall movement velocity. Statistical analysis was performed with SPSS-Windows 10.1 software.

RESULTS

Three months after intracoronary cell therapy, the infarct size was reduced by 30%, whereas the global LV ejection fraction increased by 15% and regional infarct wall movement velocity by 57% (Tables 2 and 3). In parallel, the clinical performance improved (Table 4), as evidenced by a higher work load demonstrated by a 11% increase in maximum oxygen uptake ($\text{VO}_{2\text{max}}$). SPECT investigation presented enhanced tetrofosmin uptake in the infarcted zone by 5%, and PET examination showed enhanced glucose uptake in the infarcted zone by 15%, demonstrating regeneration of formerly avital, chronically infarcted heart muscle (Fig. 2). An unchanged or even impaired LV function was not observed in any patient.

In the control group (18 patients with chronic MI, but without stem cell therapy) no significant changes were observed in infarct size, LV ejection fraction, or wall

Table 3. Cardiac Parameters in the Transplantation Group and in Control Group at the Three Investigation Time Points

	Area of Infarction, %			LV Ejection Fraction, %			Infarction Wall Movement Velocity, cm/s		
	Control Group	Tx Group	p Value*	Control Group	Tx Group	p Value*	Control Group	Tx Group	p Value*
Investigation 1	25 ± 9	26 ± 7	0.99	53 ± 10	53 ± 8	0.87	1.95 ± 0.66	1.80 ± 0.63	0.57
Investigation 2	27 ± 9	27 ± 8	0.83	51 ± 10	52 ± 9	1.00	1.88 ± 0.76	1.86 ± 0.70	0.94
Investigation 3	26 ± 9	19 ± 9	0.02	52 ± 10	60 ± 7	0.02	1.91 ± 0.79	2.92 ± 0.91	0.001

*Analysis of variance.

Abbreviations as in Table 1.

Table 4. Positron Emission Tomography and Spiroergometry Before and After Stem Cell Therapy in Chronically Infarcted Myocardium

	¹⁸ F-FDG-Positron Emission Tomography			VO _{2max} Spiroergometry	
	FDG Uptake, %	Difference in %		ml/min	Difference in %
Investigation 1	none			none	
Investigation 2	43.8 ± 8.0	>	+ 15	1,602 ± 533	>
Investigation 3	50.5 ± 11.6			1,776 ± 523	
p (Wilcoxon test)	0.012			0.0001	

¹⁸F-FDG = ¹⁸F-fluor-deoxy-glucose; VO_{2max} = maximum oxygen uptake.

movement velocity of the infarcted area (Figs. 3A to 3C). Electrocardiogram at rest and on exercise and 24-h Holter ECG revealed no rhythm disturbances at any time point. Only 1 patient (from 18 cell-treated patients, 6%) developed relevant restenosis due to quantitative angiographic criteria. The restenosis could be treated adequately by stent implantation. The other 17 patients showed good patency rates without restenosis after PCI and cell transplantation. They also revealed no alterations in LV function 8 ± 5 months after PTCA.

There was no inflammatory response or myocardial reaction (white blood cell count, CRP, CPK) after cell therapy, despite a moderate increase in CRP (before cell transplantation 0.58 ± 0.48 mg/dl, after cell transplantation 1.07 ± 0.73 U/l, p = 0.002), which is usual after bone marrow puncture and/or cardiac catheterization.

DISCUSSION

The results of these investigations demonstrate, for the first time, that the intracoronary transplantation of autologous bone marrow mononuclear cells may reduce infarct size and improve LV function as well as myocardial glucose uptake in chronic ischemic heart disease attributable to chronic MI (5 months to 8.5 years old). Infarct size decreased in all patients and cardiac performance (ejection fraction, wall movement velocity of infarcted area, maximum oxygen uptake, and exercise tolerance) and myocardial metabolism (FDG-PET) improved, all being between 11% and 57%. Furthermore, it is noteworthy that there were no complications immediately or three months after cell transplantation, especially that there was no cardiac arrhythmia and no signs of cardiac or systemic inflammation were present.

The effects of stem cell transplantation on infarct size, cardiac function, and contractility demonstrate significant improvement of these three parameters in the therapy group (before and after stem cell therapy) as well as in the comparison between the stem cell therapy group and the control group, thus giving evidence for a beneficial therapeutic effect of stem cell therapy on cardiac performance in chronic MI.

Patients in both the stem-cell group and the control group were recruited in parallel to each other and consecutively between January 2003 and March 2004. They all (n = 36) fulfilled the same inclusion criteria. Thus, representative patient characteristics were present for the stem cell group (n = 18) and the control group (n = 18) as well as in comparing both of them. Moreover, two subsequent investigations before stem cell transplantation have been performed for each patient: investigation 1 and 2 demonstrated the stability of LV dynamics before cell therapy (9 months respectively 10 days before transplantation) and investigation 3 compared the effects of stem cell therapy with the control group. The stable hemodynamics during the preceding 9 ± 6 months before stem-cell therapy and the stable hemodynamics within the control group at all three points of investigation underline the significant alterations of the left ventriculography-derived parameters investigated after stem cell transplantation.

The regenerative potential of bone-marrow-derived stem cells may be explained by any of four mechanisms: 1) direct cell differentiation from mononuclear cells to cardiac myocytes (17), 2) cytokine-induced growing and increase of residual viable myocytes, especially within the border zone of the infarcted area (18), 3) stimulation of intrinsic myocardial stem cells (endogenous stem cells) (19,20), and 4)

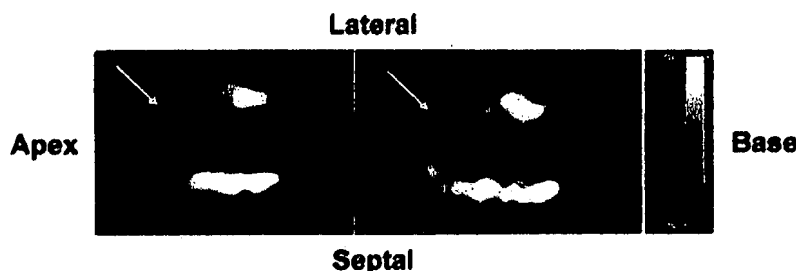


Figure 2. Representative illustration of ¹⁸F-FDG-positron emission tomography (PET) before (above) and 3 months after (below) cell therapy in the transversal (left) and longitudinal (right projection) in a 30-year-old male patient with an 8-month-old anteroapical infarction. Note the restoration of glucose uptake (below) within the infarcted area of the formerly completely avital anteroapical myocardium.

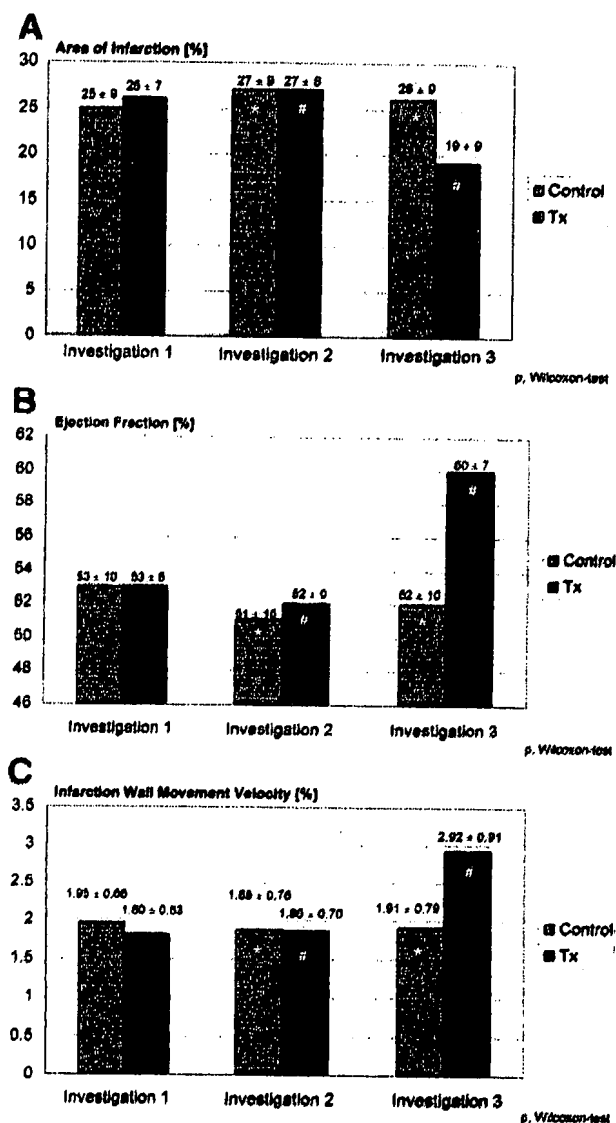


Figure 3. Illustration of the mean values of (A) area of infarction, (B) ejection fraction, and (C) infarction wall movement velocity, determined by quantitative left ventriculography in both groups (control group vs. transplantation [Tx] group) at the point of time: investigations 1, 2, and 3. Comparison of both groups with chronically infarcted myocardium (control group vs. Tx group), $n = 18$ patients. Investigation 1 was 9 ± 6 months before cell transplantation (controls: 9 ± 5 months before percutaneous transluminal coronary angioplasty [PTCA]); investigation 2 within 10 days before cell transplantation (controls: at the time point of PTCA) and investigation 3 was three months after cell transplantation (controls: 8 ± 5 months after PTCA). Note the significant decrease of infarct size and the increase in ejection fraction and in contractility (infarction wall movement velocity) 3 months after cell therapy in comparison with the control group. * $p =$ not significant (investigation 2 vs. investigation 3); # $p = 0.001$ (investigation 2 vs. investigation 3).

induction of cell fusion between transplanted bone marrow cells and resident myocytes (21–24).

Transdifferentiation has been described by previous investigators (4); however, it has been questioned by recent experimental studies (25). The influence of cytokines has

shown to restore coronary blood vessels and muscle cells after experimental myocardial infarction. This regeneration of blood vessels and muscle cells is most pronounced in the border zone of ischemic and/or infarcted tissue (26), demonstrating an enhancement of mitotic cells and cell cycles up four-fold, when compared to areas remote from the necrotic myocardium. Moreover, mononuclear bone marrow stem cells contain a lot of cytokines (VEGF, insulin-like growth factor, platelet-derived growth factor, and so on), thereby stimulating residual normal myocytes for regeneration and proliferation and intrinsic myocardial stem cells (endogenous stem cells) for cell regeneration and for cell fusion (27–31).

Mitotic indexes are three to four times more frequent within the border zone of myocardial necrosis when compared with non-injured heart muscle (26). Moreover, 20% to 40% of intracoronarily transplanted bone-marrow-derived stem cells may be accumulated within the border zone of MI. There were no signs of apparent microcirculation disturbances because all patients had Thrombolysis In Myocardial Infarction flow grade 3. Thus, it is conceivable that in MI the border zone represents the optimum “niche” for exogenously transplanted stem cells, stimulating mitosis rates and heart muscle regeneration, preferably originating in and expanding from these areas. Cell fusion may also contribute to heart muscle regeneration, which takes its origin from the border zone, expanding gradually to the necrotic core of the infarcted area.

Our study cannot determine which cell-biologic and molecular mechanisms are responsible for heart muscle repair or which of the studied factors may play the predominant role. However, the final functional outcome of this cell therapy demonstrates three main target effects: improvement in muscle function (pumping ability and contractility), myocardial perfusion (SPECT), and myocardial glucose metabolism (PET), thus giving evidence that heart muscle repair must have taken place by this intracoronary bone marrow cell transplantation procedure.

The clinical significance of this novel therapeutic approach may embrace a large number of patients with chronic coronary artery disease, preferably after previous or longstanding MI. It is conceivable that remodeling after infarction may be ameliorated or even stopped by this procedure. Thus, cell therapy may represent a new option of basic and causal therapy in chronic infarcted myocardium. It is an open question whether variations of the amount and kind of bone marrow cells, the administration technique, and the transplantation procedure itself, by enhanced environment and improvement of the angiogenic microenvironment, can further improve the milieu-dependent differentiation or regeneration of bone marrow cells in chronic infarcted heart disease. Therefore, our clinical results represent a stable basis to proceed to the next necessary step: to a larger prospective randomized study.

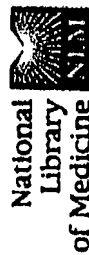
Reprint requests and correspondence: Dr. Bodo E. Strauer, Head of Department of Medicine, Division of Cardiology, Pneumology, and Angiology, Heinrich-Heine-University, Moorenstr. 5, 40225 Düsseldorf, Germany. E-mail: strauer@med.uni-duesseldorf.de.

REFERENCES

- Pfeffer MA, Braunwald E. Ventricular remodeling after myocardial infarction. Experimental observations and clinical implications. *Circulation* 1990;81:1161-72.
- Pfeffer MA. Left ventricular remodeling after acute myocardial infarction. *Annu Rev Med* 1995;46:455-66.
- Zijlstra F, de Boer M, Beukema W, et al. Mortality, reinfarction, left ventricular ejection fraction and costs following reperfusion therapies for acute myocardial infarction. *Eur Heart J* 1996;17:382-7.
- Orlic D, Kajstura J, Chimenti S, et al. Bone marrow cells regenerate infarcted myocardium. *Nature* 2001;410:701-5.
- Strauer BE, Brehm M, Zeus T, et al. Myocardial regeneration after intracoronary transplantation of human autologous stem cells following acute myocardial infarction. *Dtsch Med Wochenschr* 2001;126:932-8.
- Strauer BE, Brehm M, Zeus T, et al. Repair of infarcted myocardium by autologous intracoronary mononuclear bone marrow cell transplantation in humans. *Circulation* 2002;106:1913-8.
- Kudo M, Wang Y, Wani MA, et al. Implantation of bone marrow stem cells reduces the infarction and fibrosis in ischemic mouse heart. *J Mol Cell Cardiol* 2003;35:1113-9.
- Fuchs S, Baffour R, Zhou YF, et al. Transendocardial delivery of autologous bone marrow enhances collateral perfusion and regional function in pigs with chronic experimental myocardial ischemia. *J Am Coll Cardiol* 2001;37:1726-32.
- Smits PC, van Geuns RJ, Poldermans D, et al. Catheter-based intramyocardial injection of autologous skeletal myoblasts as a primary treatment of ischemic heart failure: clinical experience with six-month follow-up. *J Am Coll Cardiol* 2003;42:2063-9.
- Perin EC, Dohmann HF, Borojevic R, et al. Transendocardial, autologous bone marrow cell transplantation for severe, chronic ischemic heart failure. *Circulation* 2003;107:2294-302.
- Stamm C, Westphal B, Kleine HD, et al. Autologous bone-marrow stem-cell transplantation for myocardial regeneration. *Lancet* 2003;361:45-6.
- Galinanes M, Loubani M, Davies J, et al. Autotransplantation of unmanipulated bone marrow into scarred myocardium is safe and enhances cardiac function in humans. *Cell Transplant* 2004;13:7-13.
- Strauer BE, Kornowski R. Stem cell therapy in perspective. *Circulation* 2003;107:929-34.
- Sheehan FH, Bolson EL, Dodge HT, et al. Advantages and applications of the Centerline method for characterizing regional ventricular function. *Circulation* 1986;74:293-305.
- Gronenschild E, Janssen J, Tijdens F, CAAS II, a second generation system for off-line and on-line quantitative coronary angiography. *Cathet Cardiovasc Diagn* 1994;33:61-75.
- Segall G. Assessment of myocardial viability by positron emission tomography. *Nucl Med Commun* 2002;23:323-30.
- Beltrami AP, Barlucchi L, Torella D, et al. Adult cardiac stem cells are multipotent and support myocardial regeneration. *Cell* 2003;114:763-76.
- Orlic D, Kajstura J, Chimenti S, et al. Mobilized bone marrow cells repair the infarcted heart, improving function and survival. *PNAS* 2001;98:10344-9.
- Leri A, Kajstura J, Anversa P. Myocyte proliferation and ventricular remodeling. *J Card Fail* 2002;8 Suppl:S518-25.
- Urbanek K, Quaini F, Tasca G, et al. Intense myocyte formation from cardiac stem cells in human cardiac hypertrophy. *PNAS* 2003;100:10440-5.
- Alvarez-Dolado M, Pardal R, Garcia-Verdugo JM, et al. Fusion of bone-marrow-derived cells with Purkinje neurons, cardiomyocytes and hepatocytes. *Nature* 2003;425:968-73.
- Oh H, Bradfute SB, Gallardo TD, et al. Cardiac progenitor cells from adult myocardium: homing, differentiation, and fusion after infarction. *PNAS* 2003;100:12313-8.
- Terada N, Hamazaki T, Oka M, et al. Bone marrow cells adopt the phenotype of other cells by spontaneous cell fusion. *Nature* 2002;416:542-5.
- Balsam LB, Wagers AJ, Christensen JL, et al. Haematopoietic stem cells adopt mature haematopoietic fates in ischaemic myocardium. *Nature* 2004;428:668-73.
- Murry CE, Soonpaa MH, Reinecke H, et al. Haematopoietic stem cells do not transdifferentiate into cardiac myocytes in myocardial infarcts. *Nature* 2004;428:664-8.
- Anversa P, Torella D, Kajstura J, et al. Myocardial regeneration. *Eur Heart J* 2002;23 Suppl G:G67-71.
- Kajstura J, Rota M, Whang B, et al. Bone marrow cells differentiate in cardiac cell lineages after infarction independently of cell fusion. *Circ Res* 2005;96:127-37.
- Torella D, Rota M, Nuszynska D, et al. Cardiac stem cell and myocyte aging, heart failure, and insulin-like growth factor-1 overexpression. *Circ Res* 2004;94:514-24.
- Nadal-Ginard B, Kajstura J, Leri A, Anversa P. Myocyte death, growth, and regeneration in cardiac hypertrophy and failure. *Circ Res* 2003;92:139-50.
- Pennica D, King KL, Shaw KJ, et al. Expression cloning of cardiotrophin 1, a cytokine that induces cardiac myocyte hypertrophy. *PNAS* 1995;92:1142-6.
- Oh H, Bradfute SB, Gallardo TD, et al. Cardiac progenitor cells from adult myocardium: homing, differentiation, and fusion after infarction. *PNAS* 2003;100:12313-8.

EVIDENCE APPENDIX

Item 7



PubMed

Nucleotide

Protein

Genome

Structure

PMC

Taxonomy

Books

Search PubMed



for

Limits

Preview/Index

History

Clipboard

Details

About Entrez

Text Version

Entrez PubMed

Overview

Help | FAQ

Tutorial

New/Noteworthy

E-Utilities

PubMed Services

Journals Database

MeSH Database

Single Citation Matcher

Batch Citation Matcher

Clinical Queries

LinkOut

Cubby

Related Resources

Order Documents

NLM Gateway

TOXNET

Consumer Health

Clinical Alerts

ClinicalTrials.gov

PubMed Central

Privacy Policy

1: Circulation 2003 Apr 21; [epub ahead of print]

Full text article at
circ.ahajournals.org

Related Articles, Links

Transendocardial, Autologous Bone Marrow Cell Transplantation for Severe, Chronic Ischemic Heart Failure.

Perin EC, Dohmann HF, Borojevic R, Silva SA, Sousa AL, Mesquita CT, Rossi ML, Carvalho AC, Dutra HS, Dohmann HJ, Silva GV, Belem L, Vivacqua R, Rangel FO, Esporcatte R, Geng YJ, Vaughn WK, Assad JA, Mesquita ET, Willerson JT.

Texas Heart Institute at St Luke's Episcopal Hospital, Houston, Tex.

BACKGROUND: This study evaluated the hypothesis that transendocardial injections of autologous mononuclear bone marrow cells in patients with end-stage ischemic heart disease could safely promote neovascularization and improve perfusion and myocardial contractility. **METHODS AND RESULTS:** Twenty-one patients were enrolled in this prospective, nonrandomized, open-label study (first 14 patients, treatment; last 7 patients, control). Baseline evaluations included complete clinical and laboratory evaluations, exercise stress (ramp treadmill), 2D Doppler echocardiogram, single-photon emission computed tomography perfusion scan, and 24-hour Holter monitoring. Bone marrow mononuclear cells were harvested, isolated, washed, and resuspended in saline for injection by NOGA catheter (15 injections of 0.2 cc). Electromechanical mapping was used to identify viable myocardium (unipolar voltage ≥ 6.9 mV) for treatment. Treated and control patients underwent 2-month noninvasive follow-up, and treated patients alone underwent a 4-month invasive follow-up according to standard protocols and with the same procedures used as at baseline. Patient population demographics and exercise test variables did not differ significantly between the treatment and control groups; only serum creatinine and brain natriuretic peptide levels varied in laboratory evaluations at follow-up, being relatively higher in control patients. At 2 months, there was a significant reduction in total reversible defect and improvement in global left ventricular

function within the treatment group and between the treatment and control groups ($P=0.02$) on quantitative single-photon emission computed tomography analysis. At 4 months, there was improvement in ejection fraction from a baseline of 20% to 29% ($P=0.003$) and a reduction in end-systolic volume ($P=0.03$) in the treated patients. Electromechanical mapping revealed significant mechanical improvement of the injected segments ($P<0.0005$) at 4 months after treatment. CONCLUSIONS: Thus, the present study demonstrates the relative safety of intramyocardial injections of bone marrow-derived stem cells in humans with severe heart failure and the potential for improving myocardial blood flow with associated enhancement of regional and global left ventricular function.

PMID: 12707230 [PubMed - as supplied by publisher]

Abstract	Show: 20	Sort	Text
----------	----------	------	------

Write to the Help Desk
 NCBI | NLM | NIH
 Department of Health & Human Services
[Freedom of Information Act](#) | [Disclaimer](#)

May 2 2003 16:34:23

EVIDENCE APPENDIX

Item 8

Milestones in the development of pediatric hematopoietic stem cell transplantation—50 years of progress

Trigg ME. Milestones in the development of pediatric hematopoietic stem cell transplantation—50 years of progress
Pediatr Transplantation 2002; 6: 465–474. © 2002 Blackwell Munksgaard

Abstract: In the 1950s, the first infusions of hematopoietic stem cells were given as a form of treatment for childhood leukemia. This heralded the beginning of a field that has expanded to include the treatment of immune deficiencies, a variety of leukemias and solid tumors, and then genetic diseases. A number of milestones are highlighted, particularly in regard to the use of alternative sources of hematopoietic stem cells such as unrelated donors, peripheral blood stem cells and umbilical cord stem cells. In addition, newer techniques of using non-myeloablative preparative regimens helped to reduce the toxicity and long-term consequences of hematopoietic stem cell transplant. Many diseases now benefit from the replacement of the marrow stem cells and the provision of a new immune system and improved immune surveillance.

Michael E. Trigg

Department of Pediatrics, Division of Blood & Bone Marrow Transplantation, A. I. duPont Hospital for Children, Wilmington, Delaware; Jefferson Medical College, Thomas Jefferson University, Philadelphia, Pennsylvania, USA

Key words: stem cell transplantation – pediatrics – blood formation

Michael E. Trigg, MD, Professor of Pediatrics, Jefferson Medical College, Chief, Division of Blood & Bone Marrow Transplantation, Department of Pediatrics, Alfred I. duPont Hospital for Children, 1600 Rockland Road, Wilmington, DE 19803, USA
Tel.: +1-302-651-5565
Fax: +1-302-651-5575
E-mail: mtrigg@nemours.org

Accepted for publication 26 March 2002

With the development of the age of atomic energy and interest in radiation, there developed an appreciation of the potential harm of radiation and the side-effects of exposing those in the industry to its effects, as well as the consequences of using atomic weapons. With this came an interest in understanding sensitivity to radiation and an appreciation for the hematopoietic toxicity it caused (1). In 1955–56, it was shown that mice could be protected from the lethal effects of total body irradiation with an infusion of allogeneic marrow, and in fact mice given an allogeneic marrow infusion could subsequently permanently accept a skin graft from the same marrow donor (2–4). These and other experiments showed clearly the radiation protective effect of transferring-in new cells and the development of a form of long-term chimerism and tolerance (5–10).

Abbreviations: ALL, acute lymphocytic leukemia; DLI, delayed lymphocyte infusions.

In this brief report, a number of milestones will be highlighted from the development of our current knowledge base in the field of pediatric hematopoietic stem cell transplantation (Table 1). Even the words 'hematopoietic stem cell transplantation' have evolved over time. Most work in this field of blood formation began using bone marrow. Blood as a source of peripheral stem cells was used with syngeneic transplants during the 1950s and 1960s (11, 12). However, our appreciation of the value of peripheral blood stem cells did not come about until the late 1980s and early 1990s (13). In the 1970s, the use of fetal liver tissue as a source of hematopoietic stem cells was investigated and applied in children with immune deficiencies (14). Early in their development, hematopoietic stem cells migrate from the yolk sack to the liver and temporarily make a home there before migrating to the spleen and bone marrow. Since that time in the 1980s, umbilical cord blood has been utilized as a source of hematopoietic stem cells. Motivated by the high concentration of these

Table 1. Milestones in pediatric hematopoietic stem cell transplantation

1	Experimental work
2	Initial work with children
3	Immune deficiencies
4	Selection of patients
5	Acute myelogenous leukemias
6	Neuroblastoma
7	Adding donors
8	Peripheral stem cell transplants
9	Cord blood transplants
10	Genetic diseases
11	Non-myeloablative transplants
12	Conclusions

cells during fetal development and soon after birth, the use of stem cells have been expanded *in vitro* and also been used for transplant purposes (15). More recently, the words 'stem cells' have taken on a new meaning because they describe cells which are usually obtained from early blastocysts or embryos. These cells have the capability of forming tissues or, in some cases, a whole living being. Thus, pediatric marrow transplantation has taken on new meaning and the sources of these hematopoietic stem cells have been expanded. This accounts for the change in name from pediatric marrow transplantation to pediatric hematopoietic stem cell transplantation.

Experimental work

Out of the atomic age and the atomic bomb experience came the work of the late 1940s and 1950s on protection from the hematologic effects of irradiation (16). A number of animal models were developed, and from this came the understanding that infusions of allogeneic marrow from inbred strains of mice could protect allogeneic irradiated mice (4, 8, 9). Infusions of marrow from related mice could also induce a tolerance to skin grafts (5). This led the way to an understanding of the cellular protective effects of infusions of marrow in otherwise irradiated individuals.

Initial work with children

In the 1950s, Thomas and others published their groundbreaking experience with infusions of syngeneic marrow into twins with leukemia who had been treated with supralethal doses of total body irradiation (12, 16). The prompt hematologic recovery and well being of these children against more than twice the lethal dose of irradiation attested to the protective effects of the marrow. However, these experiments took place during the initial phases of transplantation

and also during the time of our initial leukemia trials, as we tried to understand how best to treat that disease. The role of continuation and maintenance chemotherapy in the treatment of childhood leukemias was not understood at all, nor were there very many chemotherapeutic agents from which to choose. As a result, by current standards these children were inadequately treated for their underlying leukemia and unfortunately had a subsequent recurrence of their underlying disease. This experience does not speak to a failure of transplantation but rather to a failure of our understanding of how to best treat leukemia back in the 1950s. It speaks to the fact that infusions of hematopoietic stem cells can provide a protective effect from the hematopoietic-suppressive effects of total body irradiation (17).

Soon after this, work began in closely related dog litter mates and this provided us with a sound scientific basis for most, if not all, of the future work to come out of the Seattle bone marrow transplant group and other groups around the world (7, 10).

Immune deficiencies

In the 1960s, a group from Minnesota used tissue typing to select a sibling marrow donor for a child with severe combined immune deficiency (14). The infusion of marrow provided immunologic recovery for this child, who is currently alive and in his 30s (14). Again, this milestone in the development of pediatric hematopoietic stem cell transplantation provided a basis for large numbers of transplants for children with a variety of immune deficiencies and set the stage for utilizing hematopoietic stem cell transplants to correct the hematopoietic and immunologic deficiencies induced by large doses of chemotherapy and radiation that are employed to treat underlying malignancies (18, 19). Although a number of other cellular and humoral factors have been utilized to treat immune deficiencies, none have been as successful as the use of hematopoietic stem cells (14, 20). Although the first child to receive such a transplant had severe combined immune deficiency and required no preparative therapy, our understanding of these many immune deficiencies has increased over time to include a realization that some of these children require immune suppression or ablation in order to make room for a new graft or to prevent stem cell graft rejection (21). These techniques, of making a space for new stem cells resulted, in some cases, in incomplete engraftment or in

History of pediatric organ transplantation

Table 2. Indications for bone marrow transplantation

Malignant disorders	Non-malignant disorders
Leukemia Acute myeloblastic leukemia Acute lymphoblastic leukemia Chronic myelogenous leukemia (adult type) Chronic myelogenous leukemia (juvenile type) Myelodysplastic syndromes Acute myelofibrosis Some less severe combined immunodeficiency disorders Lymphoproliferative disorders Hodgkin lymphoma Non-Hodgkin lymphoma Multiple myeloma Chronic lymphocytic leukemia Solid tumors Neuroblastoma Bronchial carcinoma Breast carcinoma Melanoma Brain tumors Osteosarcoma Ewing's Sarcoma Teratomas Rhabdomyosarcoma Others	Bone marrow failure syndromes Acquired severe aplastic anemia Fanconi's aplastic anemia Reticular dysgenesis Immunodeficiency states Severe combined immunodeficiency disease Wiskott-Aldrich syndrome Some acquired immune deficiency syndrome (AIDS) Hematological disorders Some Thalassemia syndromes Some sickle cell anemias Some congenital neutropenia Some severe congenital platelet disorders Some Osteopetrosis Genetic disorders Mucopolysaccharidoses Leukodystrophies Other rare metabolic disorders Connective tissue disorders Some juvenile rheumatoid arthritis Some systemic lupus erythematosus

others, in complete hematopoietic engraftment (14). At the present time, almost 75% of all those with severe immune deficiencies can be cured with hematopoietic stem cell transplants and the rate is even higher when a matched sibling donor is available (14).

Selection of patients

Indications for hematopoietic stem cell transplant have, to some extent, changed over the years, particularly related to the curative treatments provided for children with leukemia (Table 2) (22). In the 1980s, the most common indication for a hematopoietic stem cell transplant was salvage therapy for patients with acute lymphocytic leukemia (ALL) who failed to maintain a remission from conventional therapy. However, the likelihood of attaining long-term remission from conventional therapy is now approaching 80%, therefore eliminating the need for a marrow transplant consideration for the vast majority of such children (23). In addition, overall survival rates have also improved for children with lymphoma. Thus, the majority of children with ALL can be cured with conventional approaches and no longer require a transplant. Considering how conventional therapy has increased in intensity over time, it is true that the vast majority of children with ALL and other diseases

who require a transplant have already been through a variety of therapeutic protocols and are thus more resistant to the curative effects of the preparative therapy which is provided (Table 3) (24).

Acute myelogenous leukemias

Throughout the 1980s, a number of studies suggested the utility of high dose therapy for patients with acute myelogenous leukemias (25). However, there are now available randomized results from a large-scale study of children with acute non-lymphocytic leukemia undergoing marrow transplantation with a matched sibling donor, compared to those who were treated with conventional chemotherapy because of a lack of a matched sibling donor (25). Very clearly, the long-term survival rates are higher in patients with acute non-lymphocytic leukemia who underwent transplantation with a matched sibling donor. The results from these trials would indicate that those with acute non-lymphocytic leukemia will do much better with a marrow transplant, provided there is a related matched sibling available, than with alternative forms of therapy. The recognition of this phenomena in a biologically randomized study is a key milestone in the development of hematopoietic stem cell transplantation, showing how this disease is quite

Table 3. Remaining stem cell transplant questions in 2002 for children with leukemia

1	What factors best define those to benefit from transplant therapy: (a) initial response; (b) minimal residual disease; (c) biologic markers such as chromosomes, cell morphology, cellular markers or <i>in vitro/in vivo</i> growth characteristics; (d) age and/or (e) genetic/pharmacologic profiles.
2	Are all stem cell donors alike in regards to outcome and relapse rates? Matched sibling vs. mismatched family member vs. unrelated adult donors vs. umbilical cord derived stem cells.
3	Role of peripheral blood vs. marrow as a source of stem cells.
4	Methods to accelerate immune maturation of stem cell transplants or methods to minimize toxicity to the immune system.

sensitive to the preparative therapy which has been provided early in the course of the disease, as well as the immunologic effects of providing a marrow graft (25).

Neuroblastoma

Of all the solid tumors that occur in the pediatric age group, neuroblastoma has been the most resistant to conventional therapy. However, slow but steady improvement has been made in overall survival (26).

The landmark study in the development of pediatric hematopoietic stem cell transplant was the cooperative group trial which compared transplantation vs. no transplant for those with Stage IV neuroblastoma (27). Previous studies had indicated that allogeneic transplants seemed to provide no evidence for an immunologic effect of the graft itself on the underlying tumor. Fewer problems were associated with high dose therapy and autologous rescue vs. allogeneic stem cell transplant. Thus, the autologous transplant was accepted as a standard therapy for those with neuroblastoma. In fact, the current Children's Oncology Group protocol offers a course of very high dose therapy with stem cell rescue to all patients (27).

One of the major questions that remains to be decided in the next few years is whether there has been an improvement in long-term survival in those with neuroblastoma by the utilization of peripheral stem cells vs. marrow, since marrow runs the risk of contamination with tumor cells. Although most trials of autologous stem cells for transplant purposes utilized a laboratory methodology to purge neuroblastoma cells from the stem cell inoculum, the sensitivity of the assay is such that there may still be quite a number of neuroblastoma cells in the stem cell preparation

that is eventually infused into patients (27). Occasionally, patterns of relapse in children following recovery of hematopoiesis suggests that there has been a miliary spread of neuroblastoma cells to a variety of tissues never previously involved with neuroblastoma, and in all likelihood, these tumor cells were contaminating the stem cell inoculum that was infused intravenously (Table 4).

Adding donors

In the late 1970s and early 1980s, a variety of clinical experiments showed the potential of applying high dose chemotherapy with hematopoietic stem cell rescue for an ever-expanding number of children and adults with malignancies, by utilizing allogeneic donors who were less than perfectly matched (28). The first group of donors to be added were those family members who were haploidentical, such as parents or siblings, or in situations where there were more extensive familial investigations to find potentially more closely matched donors (29, 30). Mismatched familial transplants are characterized by one or more major histocompatibility loci differences between donor and recipient, or complete haploidentical transplants, where there is a complete disparity at one of the two HLA A, B or DR loci, and these transplants took place in great numbers in the 1980s and early 1990s (13). Most, but not all, of these clinical protocols attempted to overcome the significant graft-vs.-host disease problems by removing more than two logs of T-lymphocytes from the donor stem cell inoculum (29, 31). From these experiments, the investigators derived a great deal of knowledge and experience in the role of T-lymphocytes when removing them from the transplant inoculum. High rates of rejection and non-engraftment

Table 4. Best approach to treatment of Stage IV neuroblastoma in 2002

1	Diagnostic biopsy with complete resection if possible.
2	Induction chemotherapy—combination—usually 5–6 cycles.
3	Subsequent surgery to resect residual disease.
4	Consolidation therapy—usually high dose chemotherapy and stem cell rescue with marrow or peripheral stem cells, purged of neuroblastoma cells if present.

Table 5. Peripheral stem cells—advantages in adults

- 1 Easier/less painful to obtain than marrow.
- 2 Quicker recovery of neutrophils and platelets when used in place of marrow as a source of stem cells
- 3 No exposure to any anesthetic as would be necessary for collecting marrow stem cells.

suggested either that the process of removing the T-lymphocytes had eliminated extensive numbers of stem cells meant to engraft and produce hematopoietic progeny, or alternatively (28), T-lymphocytes from the donor derived inoculum were important in providing further immune suppression within the host to enable hematopoietic stem cell engraftment. To overcome the latter problem, more preparative therapy was given to some patients but this did not necessarily seem to make a difference. In addition, the removal of T-lymphocytes increased the potential for opportunistic infections due to the subsequent delay in T-lymphocyte recovery, and the delay in T-lymphocyte recovery led to an increased relapse rate in those patients receiving such marrow grafts which were T-lymphocyte depleted. However, the goal of using mismatched or haploidentical donors had achieved an important milestone which was now making it possible for every patient who had a disease that could be treated with hematopoietic stem cell transplant to be eligible for such treatment, without limitations as to whether a donor was available or not (31). The only children who failed to find a donor when considering the use of hematopoietic stem cells from mismatched or haploidentical family donors were those who had been adopted and whose families were not traceable, or alternatively children who had no siblings or parents due to some sort of environmental accident or early death.

Alongside the development of mismatched or haploidentical transplantation was the typing and recruitment of unrelated adult donors. Considering the increased knowledge of the histocompatibility loci and the further delineation of the unique HLA disparities between individuals using molecular techniques, investigations continued on typing large numbers of individuals who volunteered to register with a number of private marrow donor banks and who agreed to be called upon to donate marrow if necessary for an individual who needed a transplant and who had a tissue type similar or identical to that of the potential donor. Reports of these transplants began to appear in the 1980s and some of these were quite successful (30). As expected, most were associated with significant graft-vs.-host disease, either because our preventive treatments for GvHD were inadequate or

alternatively, because there had been a lack of appreciation of minor histocompatibility differences between donor and recipient. As an increasing number of allogeneic transplants using unrelated donors developed, there also developed a national interest in bringing together all of these donor banks and setting up a single national registry supplying all allogeneic hematopoietic stem cells for transplant purposes. Thus came the government directive to set up the National Marrow Donor Program and the National Marrow Donor Registry. With increasing recruitment efforts through the 1980s and into the 1990s, well over 4,000,000 donors have been registered world-wide and listed in the NMDP registry, making it possible to find close if not perfect matches more than half the time for individuals without a matched familial donor. Several studies have documented that children will tolerate some histocompatibility difference between donor and recipient, making it possible to find unrelated donors for children with an ever increasing frequency (31). Again, another milestone had been achieved in the development of the pediatric hematopoietic stem cell field by continuing to increase the population of children eligible for transplant by defining new groups of donors, thereby circumventing the earlier problem whereby many patients could not avail themselves of marrow transplant therapy for lack of a matched related allogeneic donor.

Peripheral stem cell transplants

Although experience with syngeneic transplants had yielded the necessary information about circulating peripheral hematopoietic stem cells, to a large extent the application of this technique did not begin until the late 1980s and early 1990s (32–34). Several considerations came about to spearhead this development:

1 Some patients with malignancies undergoing high dose chemotherapy with hematopoietic stem cell reconstitution had previously received irra-

Table 6. Non-myeloablative transplant preparation

- 1 Reduced preparative therapy and thus potential reduced toxicity
- 2 Potential to eliminate all radiation.
- 3 Reduce growth retardation and neuro-psychological effects of full myeloablative therapy.

diation to large marrow spaces or alternatively, had a potential contamination of marrow spaces with tumor cells. Collection of peripheral blood might obviate this problem since there were few if any detectable malignant cells in the peripheral blood and thus collections of stem cells in the peripheral blood would theoretically be less likely to be contaminated with tumor cells.

2 Going to the operating room to obtain hematopoietic stem cells from the marrow space was traumatic and painful, and collections of peripheral blood stem cells could be done as an outpatient in a less painful setting with far less morbidity (33).

As a result, many studies were published on the utilization of peripheral stem cells on how best to collect them and on the timing of collection (32-34). With the development of hematopoietic stem cell cytokines stimulating the production of CD34+ cells, there was now an understanding of how best to improve the yield of collected stem cells. Subsequent studies showed that peripheral stem cells, when given to a host who had undergone ablative chemotherapy or chemotherapy plus radiation, resulted in faster hematopoietic engraftment, fewer hospital days, and potentially improved immune function more quickly than when using hematopoietic stem cells from the marrow space (34). Although similar studies have not been done in children due to the numbers of children available for such studies, in some diseases we have already accepted the utility of obtaining peripheral blood stem cells for transplant purposes (33, 35, 36). However, in many situations in pediatrics, the donors are small children and it therefore becomes more difficult to collect peripheral stem cells from them and more likely that we can obtain an equal or higher total number of hematopoietic stem cells by taking the donor to the operating room and obtaining bone marrow in the traditional manner (33). Thus, even though the use of peripheral stem cells was first documented in pediatric transplants more than 30 years ago, when blood from one twin was infused into another (11), the large scale use of peripheral stem cells, the appreciation of the fact that the stem cells collected from the peripheral blood were somewhat different from those collected from the marrow space, and the extension of this technique to those who have potential marrow involvement with tumor was a whole new era and marked a new milestone in the further development of pediatric hematopoietic stem cell transplantation (Table 5) (32, 33).

Cord blood transplants

In the 1970s, the migration of hematopoietic stem cells from the yolk sack to the liver was understood to the extent that fetal livers were obtained from aborted fetuses and successfully used for transplant purposes (37). Over a period of 10 years, there developed an appreciation of the vast numbers of hematopoietic stem cells in the developing fetus, which at the time of birth is more than 100-fold greater per volume of blood than that found in adults. The sense was that the developing fetus was rapidly growing and needed ever increasing quantities of hematopoietic and immunologic cells. This was translated into an increased number of hematopoietic stem cells, developing in a naïve individual and incapable of recognizing and rejecting maternal tissue and probably having little capacity to reject other tissue soon after birth. We knew from hundreds of years of clinical observation that infants were highly susceptible to a wide variety of infectious problems, only because of the lack of immunity at birth and that there was a normal maturation of the immune system in the first 1-2 years of life. In fact, it was well known that children with immune deficiencies did not usually present within the first few months of birth because of the protective levels of cellular and humoral immune factors that are passed from mother to fetus, and that when this protection eventually waned, then the infant's immunodeficiency was exposed (20). With this background in mind, there developed an appreciation of the idea of obtaining umbilical cord blood at the time of birth and using it as a rich source of hematopoietic stem cells which could then be used for transplant purposes (15, 38). Several significant advantages immediately became apparent:

1 With millions of births worldwide, it should be possible to collect umbilical cord blood from every child soon after birth and such blood/cells could be typed and cryopreserved and made available to any individuals with a similar tissue type.

2 Unlike the situation where an adult allogeneic donor needs to be taken to the operating room to obtain marrow or needs to visit a blood bank in order to provide peripheral blood stem cells, the use of these umbilical cord cells entailed no harm to the donor since these blood cells were usually discarded with the placenta at the time of birth.

3 Because the umbilical cord-derived hematopoietic stem cells were naïve, it should be possible to use them more readily in situations where there was not a perfect HLA match between donor and

recipient and still see a good hematopoietic engraftment without the lethal complications of graft-vs.-host disease.

In 1988, the first such transplant took place in a child with Fanconi's anemia, receiving hematopoietic stem cells from a previously collected umbilical cord blood specimen from a sibling who was perfectly matched (39, 40). Since that time, a number of umbilical cord blood banks have been set up around the world, some linked by computer and some not, but all dedicated to the business of providing umbilical cord blood for transplant purposes, particularly in situations where there were no available matched sibling donors (15). These transplants have continued in select centers around the world and to date, there still are some problems and difficulties:

1 Umbilical cord-derived hematopoietic stem cells tend to grow a little more slowly, perhaps related to their concentration and numbers following cryopreservation, than freshly obtained peripheral blood stem cells or marrow stem cells from adult donors. The slowness to recovery of blood counts may predispose the transplanted patient to an increased number of infectious problems and complications (38).

2 Although one of the advantages of using hematopoietic stem cells derived from the umbilical cord is their relative naiveté and the potential for a lack of graft-vs.-host disease, there developed a sense in the 1980s and 1990s of the importance of graft-vs.-host disease in providing an immunologic method for controlling the subsequent recurrence of a malignancy following high dose preparative therapy. The absence of such graft-vs.-host disease following the transplant of hematopoietic stem cells derived from umbilical cord predisposes some patients to an increased risk for recurrence of their underlying leukemia (15); and

3 The amount of cord blood obtained in the delivery room is directly related to the experience of the operator. The amount of cord blood collected correlates with the number of stem cells and the number of stem cells in a sample used for transplant purposes correlates with the rate of engraftment. The rate of engraftment and the number of stem cells appears to correlate with ultimate survival (41). Thus, there are many specimens of umbilical cord blood obtained in the delivery room which are low in volume and which are therefore probably not well suited for use in larger children or adults (38).

The availability of umbilical cord blood has once again been a milestone because it has enabled more children who otherwise could not

find a suitable donor to have one available. In addition, many children undergoing marrow transplantation need not suffer graft-vs.-host disease and its attendant morbidity/mortality, such as those with genetic diseases or immune deficiencies (40-42). As a result, cord blood has then become an ideal source of hematopoietic stem cells. The first patient transplanted with umbilical cord blood as previously mentioned was a child with Fanconi's anemia, and there had not yet been any evidence of myelodysplasia or leukemic changes (39, 40). As a result, the child would not theoretically benefit from the occurrence of any graft-vs.-host disease and thus the use of umbilical cord blood as a source of hematopoietic stem cells was an ideal source in that particular situation. The overall place of umbilical cord blood in the field of hematopoietic stem cell transplantation remains to be determined, but clearly the advent of this technology and the establishment of numerous umbilical cord blood banks has been a major milestone in the field of pediatric hematopoietic transplantation.

Genetic diseases

For some patients, hematopoietic stem cell transplants are a very crude way of providing new genetic material. It is not selective but rather provides a host of hematopoietic and immunologic stem cells, all of which may bring missing enzymes or missing substrates not present in a child undergoing such a transplant for what is presently defective. Two such examples are noteworthy.

Almost 20 years ago, the first marrow transplant took place in a child with sickle cell anemia (42). Sickle cell anemia is an inherited genetic disorder characterized by abnormal hemopoiesis, deformities and increased sickling of red cells secondary to abnormal hemoglobin synthesis and an associated array of clinical difficulties. The replacement of these abnormal hematopoietic stem cells with new ones making cells with normal hemoglobin eliminates future problems associated with the disease for the child undergoing such a transplant (43). In fact, hematopoietic transplants have been curative (40). The problem is related primarily to the selection of patients: when to offer the transplants to patients and when too much damage may have been done by the underlying disease to warrant proceeding with the transplants. Since the vast majority of children with sickle cell disease make it into adult life, families have a very difficult time

making a decision to proceed with a marrow transplant when there might be as high as a 15-30% significant morbidity and mortality associated with the procedure (44, 45).

A number of mucopolysaccharide disorders, as an example of another genetic disorder, have been cured with a hematopoietic stem cell transplant (46). In these cases, there is a missing enzyme which can be found within neutrophils and other cells originating from the bone marrow space, and these new cells will replace the missing enzyme and deal with substrates which accumulate in abnormal places where such accumulation may result in significant damage. In most of the mucopolysaccharide disorders, proteins accumulate in the nervous system as well as in the heart and liver and eventually lead to the dysfunction of these organs and death. A marrow transplant provides enzymes which are missing to help degrade and deal with the substrate which accumulates. Damage previously associated with the abnormal condition may not be repaired over time but in general, one is looking to halt the progression of the underlying disease.

Significant progress has been made in learning for which diseases it is best to offer such transplants, the appropriate timing, and the limitation of the procedures in other patients in terms of correcting or halting the further progression of the underlying illness.

Non-myeloablative transplants

It has been well known for some time that one of the beneficial aspects of an allogeneic stem cell transplant has been the immunologic surveillance and immunologic treatment which is brought about by the growth of a new immune system. Patients experience graft-vs.-host disease with variable frequency and graft-vs.-host disease has been shown in several studies to be helpful if not essential in preventing the recurrence of an underlying malignancy. There appears to be a graft-vs.-tumor or graft-vs.-leukemia effect from the new source of allogeneic hematopoietic stem cells (47, 48).

This fact was further emphasized by studies using delayed lymphocyte infusions (DLI) to treat recurrent disease, as well as to treat significant viral infections which occur post-transplantation (47-49). Considering the delay in the recovery of normal T-lymphocytes following an allograft, lymphocytes can be obtained from living hematopoietic stem cell donors and these cells can be utilized for treatment, either to prevent recurrent diseases by prophylactic infu-

sions of such lymphocytes or to treat the recurrent chronic myelogenous leukemia which may occur following a transplant.

Considering the immunologic effects of the graft, and animal work which showed that with minimal conditioning therapy and minimal establishment of an allograft within the bone marrow space, further immune suppression from donor lymphocytes would effectively create complete chimerism and full engraftment, thereby providing individuals with the benefit of the new immune system provided with the new allograft (50). This approach holds great promise in establishing the presence of a new allograft and as such is really the goal of the transplant process and at the same time, limit the preparative therapy given, since a complete establishment of hematopoiesis is not initially necessary but will be effected by subsequent infusions of lymphocytes. This has been one of the milestones in the development of pediatric hematopoietic stem cell transplantation. Those with an enzyme disorder, and potentially those with disorders only characterized by missing cellular function, need a small proportion of hematopoietic stem cells to fully engraft and function to correct the underlying defect (50). A recent patient reported with Chédiak Higashi syndrome is a good example whereby only a small proportion of the neutrophils are actually of donor origin and therefore normal, but the actual number is high enough to have prevented any significant infections occurring in the years following the establishment of the allograft (51).

The use of these non-myeloablative transplants, meaning that the preparative therapy has been limited in extent and toxicity, has permitted a very small proportion of donor cells to engraft, but these cells are then engineered to become fully chimeric with the recipient by the use of DLI to further suppress the host. These types of transplants, whereby the preparative therapy is minimized, are undergoing trials in the elderly and in more fragile individuals who would otherwise not qualify for transplant, in the hope that they will receive the beneficial effects of a new immune system and the immunologic surveillance thus provided, without the toxicity which often is associated with the preparative therapy given to bring about full engraftment. In addition, there are children with disorders that would potentially be made worse with full doses of preparative therapy, including total body irradiation. Thus a non-myeloablative transplant limits the amount and type of preparative therapy given, thereby limiting the potential toxicity to

the patient. Limiting the preparative therapy reduces the long-term effects but still permits full eventual engraftment and provision of the missing components of the immune system or immune surveillance (Table 6).

Conclusions

When hematopoietic stem cell transplants began almost 50 years ago, they were a shot in the dark to try and correct underlying significant diseases and were eventually utilized to correct immune deficiencies. Over the ensuing 40 years, a variety of newer techniques were developed to perfect the source of stem cells and increase our understanding of clinical situations in which one source of stem cells might be better than another (52, 53). Improvements in supportive care made it possible for an ever increasing number of children to survive the effects of the preparative therapy and the subsequent graft-vs.-host disease and immaturity of the immune system. High dose therapy with hematopoietic stem cell rescue has become a mainstay of modern therapy for children with Stage IV neuroblastoma and a significant salvage therapy for patients with a variety of other diseases when these children have failed to respond to more conventional approaches. The approaching frontier involves the *in utero* identification of genetic defects and immunodeficiency diseases, thereby utilizing *in utero*-administered hematopoietic stem cells to provide definitive curative therapy (54, 55). Although children with leukemia make up an ever decreasing number of those eligible for transplantation because of the success of the initial therapeutic non-transplant treatment they receive (22), there still are a number of children benefiting from transplantation who have been treated rather heavily with alternative therapy and we know that there are a variety of diseases that now benefit from replacement of the marrow graft and provision of a new immune system and improved immune surveillance.

References

1. CAVINS JA, SCHIEER SC, THOMAS ED, FERREBEE JW. The recovery of lethally irradiated dogs given infusions of autologous leukocytes preserved at -80°C . *Blood* 1964; 23: 8-43.
2. FORD CE, HAMERTON JL, BARNES DWH, LOMITT FJ. Cytological identification of radiation-chimaeras. *Nature* 1956; 177: 452-454.
3. JACOBSON LO, MARKS EK, ROBINSON MJ, GASTON EO, ZIRKLE RE. Effect of spleen protection on mortality following x-irradiation. *J Clin Med* 1949; 34: 1538-1543.
4. LORENZ E, UHROPF D, REID TR, SHELTON E. Modification of irradiation injury on mice and guinea pigs by bone marrow injections. *J Natl Cancer Inst* 1951; 12: 197-201.
5. MAIN JM, PREHN RT. Successful skin homografts after the administration of high dosage x radiation and homologous bone marrow. *J Natl Cancer Inst* 1955; 15: 1023-1029.
6. MANNICK JA, LOCITE HL Jr, ANHLEY CA, THOMAS ED, FERREBEE JW. Autografts of bone marrow in dogs after lethal total-body radiation. *Blood* 1960; 15: 255-266.
7. THOMAS ED, COLLINS JA, KERNAN EC Jr, FERREBEE JW. Marrow transplants in lethally irradiated dogs given methotrexate. *Blood* 1962; 19: 217-228.
8. MATTHE G, ANIEL JL, SCHWARZENBERG L, CATTON A, SCHNEIDER M. Adoptive immunotherapy of acute leukemia: Experimental and clinical results. *Cancer Res* 1965; 25: 1525-1531.
9. SANTOS GW, OWENS AH Jr. Allogeneic marrow transplants in cyclophosphamide treated mice. *Transplant Proc* 1969; 1: 44-46.
10. STORN R, EPSTEIN RB, GRAHAM TC, THOMAS ED. Methotrexate regimens for control of graft-versus-host disease in dogs with allogeneic marrow grafts. *Transplantation* 1970; 9: 240-246.
11. THOMAS ED, LOCITE HL Jr, CANNON JH, SAHLER OD, FERREBEE JW. Supralethal whole body irradiation and isologous marrow transplantation in man. *J Clin Invest* 1959; 38: 1709-1716.
12. THOMAS ED, LOCITE HL Jr, LU WC, FERREBEE JW. Intravenous infusion of bone marrow in patients receiving radiation and chemotherapy. *N Engl J Med* 1957; 257: 491-496.
13. TRIGG ME. The sources and uses of hematopoietic stem cells keep increasing. *Del Med J* 1998; 70: 387-392.
14. GOOD RA, VERJEE T. Historical and current perspectives on bone marrow transplantation for prevention and treatment of immunodeficiencies and autoimmunities. *Biol Bone Marrow Transpl* 2001; 7: 123-135.
15. WAGNER JE, KERNAN NA, STEINMUTH M, BROXMEYER HE, GLUCKMAN E. Allogeneic sibling umbilical-cord-blood transplantation in children with malignant and non-malignant disease. *Lancet* 1995; 346: 214-219.
16. THOMAS ED, BLUME KG. Historical markers in the development of allogeneic hematopoietic cell transplantation. *Biol Bone Marrow Transpl* 1999; 5: 341-346.
17. THOMAS ED, FLORENOV N, BUCKNER CD, et al. Cure of leukemia by marrow transplantation. *Leuk Res* 1977; 1: 67-70.
18. THOMAS ED, STORN R, CLIFT RA, et al. Bone marrow transplantation. *N Engl J Med* 1975; 292: 832-843, 895-902.
19. KRIVAN G, TIMAR L, GODA V, et al. Bone marrow transplantation in non-malignant disorders. *Bone Marrow Transpl* 1998; 22 (Suppl. 4): S80-S83.
20. BUCKLEY RH. Primary immunodeficiency diseases due to defects in lymphocytes. *N Engl J Med* 2000; 343: 1313-1324.
21. MOEN RC, HOROWITZ SD, SONDEL PM, et al. Immunologic reconstitution after haploidentical bone marrow transplantation for immune deficiency disorders. Treatment of bone marrow cells with monoclonal antibody CT-2 and complement. *Blood* 1987; 70: 664-669.
22. WHEELER KA, RICHARDS SM, BAILEY CC, et al. Bone marrow transplantation versus chemotherapy in the treatment of very high-risk childhood acute lymphoblastic leukemia in first remission. Results from Medical Research Council UKALL X and XI. *Blood* 2000; 96: 2412-2418.
23. GAYNON PS, TRIGG ME, HEEREMA NA, et al. Children's Cancer Group trials in childhood acute lymphoblastic leukemia. *Leukemia*, 1983-95; 14: 2223-2233, 2000.
24. HARRISON G, RICHARDS S, LAWSON S, et al. Comparison of allogeneic transplant versus chemotherapy for relapsed childhood acute lymphoblastic leukemia in the MRC UKALL R1 trial. *Biol Oncol* 2000; 11: 999-1006.
25. WOODS WG, NEIDORF S, GOLD S, et al. A comparison of allogeneic bone marrow transplantation, autologous bone marrow trans-

1815-1822.

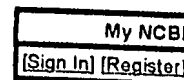
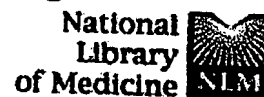
39. GUICHMAN E, BAUMMEYER H, JURECH A, et al. Hematopoietic reconstitution in a patient with Fanconi's anemia by means of umbilical-cord blood from an HLA-identical sibling. *N Engl J Med* 1994; 331: 1774-1778.

transplantation. *Bone Marrow Transpl* 1991; 7 (Suppl. 3): 92-97.

55. JONES DRE, BUI T-H, ANDERSON EM, et al. In utero haematopoietic stem cell transplantation: Current perspectives and future potential. *Bone Marrow Transpl* 1996; 18: 831-837.

EVIDENCE APPENDIX

Item 9



Entrez

PubMed

Nucleotide

Protein

Genome

Structure

OMIM

PMC

Journals

Books

Search PubMed for

Go

Direct



Limits

Preview/Index

History

Clipboard

Details

Limits: Publication Date from 1990 to 1990



Abstract

Show:

20

Sort

Send to

Text

All: 1

☐ 1: Prog Clin Biol Res. 1990;333:379-85.

Related Articles, Links

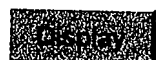
Automated isolation of mononuclear cells using the Fenwal CS3000 blood cell separator.

Areman EM, Cullis H, Sacher RA, Cottler-Fox M, Deeg HJ.

Division of Transfusion Medicine and Bone Marrow Transplantation, Georgetown University Hospital, Washington, D.C.

We describe a method for in vitro isolation of mononuclear cells from peripheral blood or bone marrow using a Fenwal CS3000 Apheresis device without employing density gradients or sedimenting agents. The automatic processing program requires minimal operator intervention and no subjective operator decisions. A mean of 67% of starting mononuclear cells were recovered in a 100 ml product having 95% mononuclear cells and less than 1% of the original red blood cells. The average processing time was 35 minutes.

PMID: 2308992 [PubMed - indexed for MEDLINE]



Abstract

Show:

20

Sort

Send to

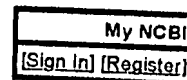
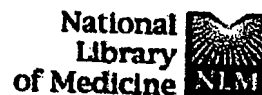
Text

[Write to the Help Desk](#)
[NCBI](#) | [NLM](#) | [NIH](#)
[Department of Health & Human Services](#)
[Privacy Statement](#) | [Freedom of Information Act](#) | [Disclaimer](#)

Feb 10 2005 12:03:04

EVIDENCE APPENDIX

Item 10



Entrez PubMed

Nucleotide

Protein

Genome

Structure

OMIM

PMC

Journals

Books

Search PubMed



for



Limits

Preview/Index

History

Clipboard

Details

Limits: Publication Date from 1990 to 1990

Abstract

Show: 20

Sort

Text

All: 1

About Entrez

Text Version

Entrez PubMed

Overview

Help | FAQ

Tutorial

New/Noteworthy

E-Utilities

PubMed Services

Journals Database

MeSH Database

Single Citation Matcher

Batch Citation Matcher

Clinical Queries

LinkOut

My NCBI (Cubby)

Related Resources

Order Documents

NLM Catalog

NLM Gateway

TOXNET

Consumer Health

Clinical Alerts

ClinicalTrials.gov

PubMed Central

☐ 1: Haematologica. 1990 Jan-Feb;75 Suppl 1:43-7.

haematologica

Related Articles, Links

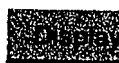
Human bone marrow processing using Cobe 2991 and CS 3000 blood cell separators for further ex vivo manipulation.

Angelini A, Dragani A, Iacone A, D'Antonio D, Accorsi P, Quaglietta AM, Berardi A, Fioritoni G, Di Bartolomeo P, D'Emilio G, et al.

Divisione di Ematologia e Centro Trasfusionale, Ospeda-le Civile, Pescara, Italy.

Several automated procedures are now available to enrich stem cells from large bone marrow (BM) volumes prior to ex vivo treatment or cryopreservation. This report details our experience using a ficoll-hypaque (F/H) gradient on Cobe 2991 cell washer and a CS 3000 continuous flow separator, on 90 BM processed for allogeneic and autologous transplantation. In the Cobe series, from 70 BM aspirates, 89 +/- 5% of the original mononuclear cells (MNC) was found in the light density fraction with a erythrocytes (RBC) and granulocytes (PMN) removal of 98 +/- 1 and 97 +/- 4.5%, respectively. Over 80% of the initial myeloid precursors (CFU-GM) were recovered in a small final volume. Twenty BM processing were performed with a CS 3000 separator using program "3" and granulocyte chamber. This technique yielded 86 +/- 9% of the initial MNC while 85 +/- 10% of RBC and 90 +/- 1.1% of PMN was removed. Over 75% of the original CFU-GM was recovered in the final product. Both techniques are effective to large-scale purification of progenitor cells and readily available as routine procedures for marrow processing.

PMID: 2351343 [PubMed - indexed for MEDLINE]



Abstract

Show: 20

Sort

Text

Write to the Help Desk

NCBI | NLM | NIH

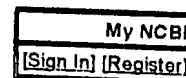
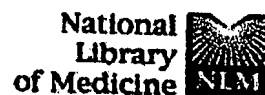
Department of Health & Human Services

Privacy Statement | Freedom of Information Act | Disclaimer

Feb 10 2005 12:03:04

EVIDENCE APPENDIX

Item 11



Entrez

PubMed

Nucleotide

Protein

Genome

Structure

OMIM

PMC

Journals

Books

Search PubMed



for

Go

Order



Limits

Preview/Index

History

Clipboard

Details

Limits: Publication Date from 1992 to 1992



Abstract

Show:

20

Sort

Send to

Text

All: 1

☐ 1: J Hematother. 1992 Winter;1(4):349-59.

Related Articles, Links

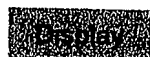
Use of the Terumo SteriCell for the processing of bone marrow and peripheral blood stem cells.

Janssen WE, Lee C, Smilee R, Carter R.

University of South Florida, Department of Internal Medicine, Tampa.

The SteriCell cell processing instrument is a good choice for a stem cell processing laboratory that is of sufficient size that they cannot share an apheresis machine with the blood bank. It is a laboratory instrument, with no facility for patient connection. Because of its minimal size and weight, it is easily stored in a cramped laboratory. Its automated programs are appropriate for processing of bone marrow and peripheral blood stem cells, and it is quite easy to learn how to use (in our laboratory, most individuals have been completely facile with the SteriCell after fewer than six processings). Based on reported results from other instruments, the SteriCell provides cell yields that are comparable to competing instruments. Service (provided by Haemonetics) has been satisfactory, and support from Terumo has been excellent. We can recommend this instrument to any other laboratory.

PMID: 1345677 [PubMed - indexed for MEDLINE]



Abstract

Show:

20

Sort

Send to

Text

[Write to the Help Desk](#)
[NCBI | NLM | NIH](#)
[Department of Health & Human Services](#)
[Privacy Statement](#) | [Freedom of Information Act](#) | [Disclaimer](#)

Feb 10 2005 12:03:04

EVIDENCE APPENDIX

Item 12

Mesenchymal Stem Cells*

Arnold I. Caplan

The Skeletal Research Center, Department of Biology, Case Western Reserve University, Cleveland, Ohio, U.S.A.

Summary: Bone and cartilage formation in the embryo and repair and turnover in the adult involve the progeny of a small number of cells called mesenchymal stem cells. These cells divide, and their progeny become committed to a specific and distinctive phenotypic pathway, a lineage with discrete steps and, finally, end-stage cells involved with fabrication of a unique tissue type, e.g., cartilage or bone. Local cuing (extrinsic factors) and the genomic potential (intrinsic factors) interact at each lineage step to control the rate and characteristic phenotype of the cells in the emerging tissue. The study of these mesenchymal stem cells, whether isolated from embryos or adults, provides the basis for the emergence of a new therapeutic technology of self-cell repair. The isolation, mitotic expansion, and site-directed delivery of autologous stem cells can govern the rapid and specific repair of skeletal tissues. **Key Words:** Mesenchymal stem cells—Bone—Cartilage—Differentiation—Self-cell therapy—Skeletal tissue—Embryo—Adult.

THE CONCEPT

It is generally agreed that in an embryo a mesenchymal stem cell is a pluripotent progenitor cell which divides many times and whose progeny eventually gives rise to skeletal tissues: cartilage, bone, tendon, ligament, marrow stroma, connective tissue (Fig. 1). By definition, these stem cells are not governed by or limited to a fixed number of mitotic divisions. Their progeny are affected by a number of factors, however, as they become tracked into very specific developmental pathways in which both intrinsic and extrinsic factors combine to control the molecular and cellular pattern of expression that results in specific tissues that perform specific functions based on their molecular repertoire (9,11).

Indeed, the progression from stem cell to final end phenotype is marked by discrete stages with transit from one stage to the next dependent on local cuing from surrounding cells (paracrine regulation) as well as signals emitted by the cell itself and the reception of its own signaling (autocrine regulation) (10,57). The sum of these various intrinsic and extrinsic signals defines the developmental position of the cells. Although difficult to reconstruct on a cell culture dish, such "positional information" has been experimentally approached by studying embryonic cells in culture, cells that have the potential to differentiate into various phenotypes (7,9,11,15).

The concept of stem cells is now well established (21,60). Two systems serve as models for such a concept: First, *Caenorhabditis elegans* is a small worm whose entire developmental lineage map has been described (21); every cell found in the adult has been carefully tracked and its progenitor tree precisely established with every branch and sub-branch delineated. Second, and to be emphasized, the hematopoietic cell lineage has been described with its several diverging pathways (21,52). It is now clear that each separate pathway and, indeed,

Received September 17, 1990; accepted January 17, 1991.

Address correspondence and reprint requests to Dr. Arnold I. Caplan at the Skeletal Research Center, Department of Biology, Case Western Reserve University, Cleveland, OH 44106, U.S.A.

* This publication was, in part, the basis for Dr. Caplan's receiving the Elizabeth Winston Lanier Award given by the American Academy of Orthopaedic Surgeons as part of their 1990 Kappa Delta Awards.

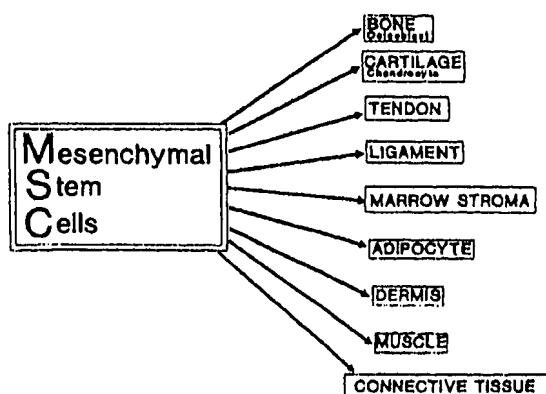


FIG. 1. Mesenchymal stem cell phenotypes. Mesenchymal stem cells are theoretically capable of differentiating through a series of separate and unique lineage transitions into a variety of end-stage phenotypes as shown.

progression through each separate stage within a discrete pathway is controlled by a balance of extrinsic and intrinsic macromolecules. Molecular biologists continue to isolate, clone, and express large amounts of these proteins, which allows use of cell culture systems to identify accurately the factor that controls progression to which stage and when (51,70). The challenge for skeletal biologists is to use the new information and new molecular tools to translate these advances into a better understanding of skeletal development, physiology, and repair.

EMBRYONIC MESENCHYMAL CELLS

The middle embryonic layer, the mesoderm, gives rise to all of the body's skeletal elements.* The term, mesenchyme, is derived from the Greek meaning "middle" (meso) "infusion" and refers to the ability of mesenchymatous cells to spread and migrate in early embryonic development between the ectodermal and endodermal layers. This characteristic migratory, space-filling ability is the key element of all wound repair in adult organisms involving mesenchymal cells in skin (dermis), bone (periosteum), or muscle (perimysium). Proteins that serve as chemoattractants, chemicals that specifically encourage this migratory activity to wound or developmental sites have been identified (24,32,59). The migratory activity of mesenchymal cells is complemented by their capacity to aggregate spe-

* For the sake of clarity, I address only issues related to cartilage or bone, although the same general experimental approach and logic can be used for other mesenchymal tissues.

cifically to form unique developmental structures or, in adults, to form repair blastemas, which are then capable of responding to local cues and differentiating accordingly to achieve regenerative repair (10,11).

Chick Limb Cells

More than 20 years ago, my collaborators and I attempted to define experimentally the conditions and cues necessary to control the differentiation of embryonic mesenchymal cells into cartilage and bone (5,7,17). Both in vivo and in vitro studies were used, but development of cell cultures and the general approach of using cell cultures has provided the experiential basis for approaching the study of mesenchymal stem cells from adults. The system we developed was the culturing of stage 24 (day 4.5) embryonic chick limb mesenchymal cells under conditions that promoted differentiation of cartilage (chondrocytes) (5,7,13,20) and bone (osteoblasts) (42,65).

Chondrocytes

Our first experimental effort with embryonic chick limb mesenchymal cells was to focus on chondrocyte development, which we learned was controlled by the initial plating density (5,17), oxygen levels (14), or, as recently shown by other investigators, a variety of physical and chemical factors (53,58,61). The key factor in the conversion of a mesenchymal cell to a chondrocyte is maintaining the progenitor cell in a round, unspread conformation. This can be accomplished simply by plating the cells initially under very compact, high-density conditions: 5×10^6 embryonic stage-24 limb mesenchymal cells per 35-mm dish (5,17). Even in a simple, defined medium consisting of insulin, transferrin, bovine serum albumin (BSA), and hydrocortisone in Eagle's minimum essential medium (MEM), the differentiation of chondrocytes and their further development can be documented as long as the cells are initially seeded at high density (18,30).

The high-density, limb cell-derived chondrocyte in culture makes two cartilage-specific molecules in abundance: type II collagen (68) and a large chondroitin sulfate, keratan sulfate proteoglycan (CSPG) (13,18,20). By detailed chemical and physical characterization of the CSPG synthesized on each day of culture, we showed that the glycosaminoglycan chains are biosynthesized slightly differently with

time (Fig. 2). Peptide maps show that the newly synthesized core protein (26) is identical on each day of culture, whereas the chondroitin sulfate chains are synthesized progressively shorter (30,000 D on day 2 to 15,000 D on day 20) and the keratan sulfate chains are synthesized progressively larger (0 to 10,000 D) (13,20). This biosynthetic progression is exactly what has subsequently been shown to occur in the cartilages of embryonic, adult, and aging human (50) and bovine specimens (62).

That embryonic chondrocytes have an aging-dependent program of changing biosynthesis is further documented when cultured embryonic chick chondrocytes are transplanted in a fibrin-based delivery vehicle into defects at the articular surface of adult chickens (29). Such chondrocytes produce what appears to be appropriate cartilaginous matrix and have been followed >18 months. The resulting repair cartilage appears to integrate perfectly into the defect and to provide the animal with a healthy, normal articular surface. These experiments and others clearly establish the concept of repairing cartilage with embryonic or appropriate reparative cells.

Osteoblasts

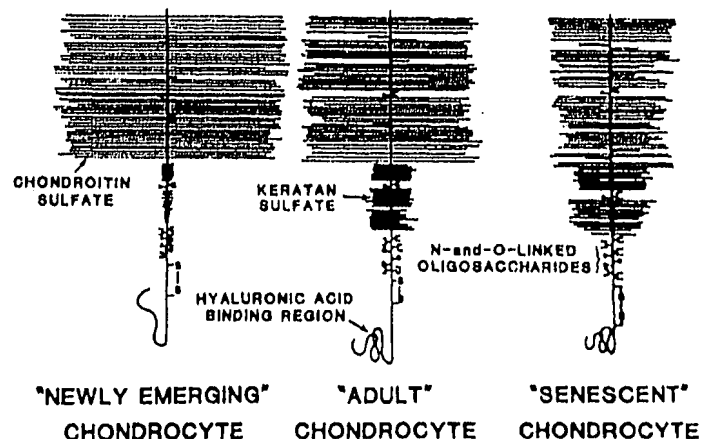
Our initial success in studying emergence of chondrocytes and formation of cartilaginous tissue from cultures of limb mesenchymal cells encouraged us to study differentiation of osteoblasts and formation of bone as well. Our initial logic was that high-density conditions caused cartilage formation and that cartilage was the progenitor tissue of bone. (Some investigators have reported that cartilage

provides the scaffold for bone formation.) After 2 years of frustrating experimentation, we realized that when infrequent bone and osteoblasts could be identified, the bone had formed at a distance from cartilage and never on or in the cartilage (42). By carefully decreasing the initial cell density of limb mesenchymal cells to just below the density at which some mineralized cartilage could form (2×10^6 cells/35-mm dish), we observed numerous deposits of bone and abundant osteoblasts which, again, were clearly at some distance from cartilage (6,42,44). In addition, these osteoblasts exhibited the classic response to parathyroid hormone (PTH) of elevated cyclic AMP levels (71,72) and possessed a bone-specific alkaline phosphatase (43). These studies clearly indicated that embryonic chick limb mesenchymal cells were capable of differentiating into osteoblasts and that the culture conditions supporting optimum osteoblast emergence were different from the conditions optimum for chondrogenesis.

Mouse and Human Limb Cells

With regard to cartilage and bone, the properties of mouse and human limb mesenchymal cells in culture appear to be quite similar, if not identical (25,46). Likewise, cartilage and bone development *in vivo* are also quite comparable, with the major exception that embryonic cartilage of chick does not calcify whereas that of mammals always calcifies (16). The comparable developmental properties of aves, rodents, and humans encourages us to continue experimentation with animal cells as an approximation of better understanding of the properties of human cells and tissues.

FIG. 2. Proteoglycans synthesized by newly differentiated, mature, and senescent chondrocytes. With increasing age, chondrocytes synthesize proteoglycans that have smaller chondroitin sulfate chains and larger keratan sulfate chains (7,8,12,13,20).



LINEAGE OF MESENCHYMAL CELLS

Cartilage

The important inference from the above discussion is that chondrocytes have a programmed (i.e., genetically dictated) sequence of changes in their end-stage expression (8,12). The differences in glycosaminoglycan chain lengths or chemistry are stable to cell culturing or metabolic perturbation. The control of these events is not known, but all experiments designed to slow this sequence of biosynthetic alterations or reverse them have failed. The inference is that a genomic mechanism somehow "tells time" and that this clock is hard-wired and unidirectional (8,12).

Such biosynthetic changes in articular cartilage are different from the lineage changes observed in adult growth plate or embryonic limb cartilage. A discrete set of expressional stages or lineage states, comprising dividing, maturing, and hypertrophic chondrocytes, is apparent in embryonic limb tissue, cell culture (13,58,61), and in the growth plate (19,28). Eventually, the hypertrophic cartilage in vivo is eroded by vascular, marrow, and phagocytic cells and replaced by bone. Each chondrocytic lineage state is uniquely different from its predecessor, as shown in Fig. 3. For example, hypertrophic chondrocytes synthesize a unique small collagen, type X, and a unique proteoglycan (54,55); neither of these molecules is synthesized by mature chon-

drocytes. In this particular circumstance, several factors are proposed to contribute to conversion of mature chondrocytes to hypertrophic chondrocytes (35); reversal of this process has not been reported.

Bone

We recently reviewed the major aspects of embryonic bone development. Figure 4 shows several important elements or rules governing this complex process (10,11,16). First, a discrete positioning of progenitor cells, stacked cells, existed in proximity to the developing bone (47). The stacked cells give rise to osteoblasts in a discrete series of lineage steps (described below). The end stage or secretory osteoblast is positioned by its proximity to vasculature, with the "back" of the osteoblast to the capillary and osteoid deposited from the "front" of this highly oriented secretory cell (47,48). The vasculature is the orientor of osteogenesis and the osteoblast is the formative element. Cartilage is not replaced by bone, but is instead the target for vascular (marrow) replacement (48); in the early limb, the cartilage model exactly defines the eventual marrow cavity.

That a discrete series of individual lineage stages exists between the progenitor cells in the stacked cell layer and the secretory osteoblasts is now clear, as shown in Fig. 5. We recently isolated four monoclonal antibodies, SB1, 2, 3, and 5, which have helped provide evidence for an osteoblast lineage (3,4). Progenitor cells in the stacked cell layer and osteocytes do not interact with SB1, 2, or 3. Newly differentiated osteogenic cells react with SB1, but not with SB2 or 3, whereas fully secretory osteoblasts react with SB1, 2, and 3. A subpopulation of osteogenic cells reacts with SB2, but not SB3. Osteocytes react with OB7.3 of Nijweide and Mulder (38) or with our SB5, but not with SB1, 2, or 3. The lineage tree in Fig. 3 is based on these observations and not only establishes the existence of an osteoblastic lineage but suggests that osteocytes are derived directly from osteoblasts with SB1, 2, and 3 antigens that are suppressed as SB5 and OB7.3 are turned on. Experiments are now in progress to use these monoclonal antibodies to isolate representatives of each lineage stage so that studies can be conducted to identify the agents that promote the progression from one lineage stage to the next. Central to the thesis presented below is the existence of osteoprogenitor cells in the stacked cell layer, the future periosteum.

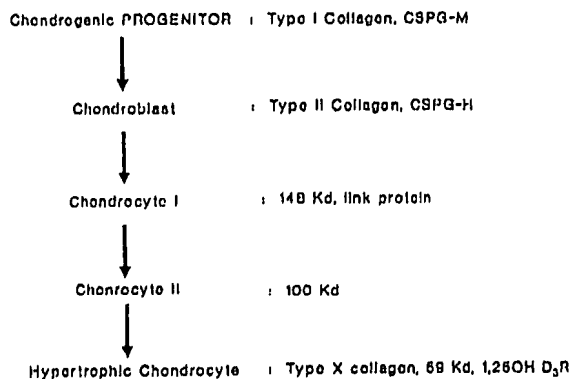
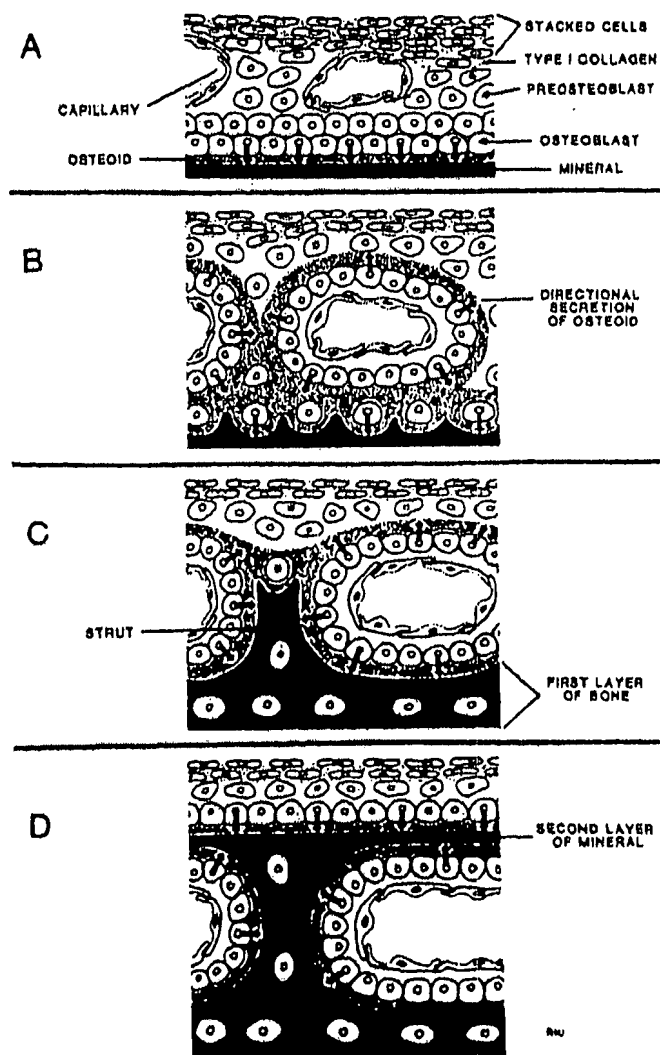


FIG. 3. Chondrogenic lineage. Based on the experiments of Solursh et al. (58,61) a hypothetical lineage map can be constructed to consist of at least five separate stages based on the changing biosynthesis of proteins (named or by molecular weight, K_d) or chondroitin sulfate proteoglycan (CSPG). The receptor for 1,25-dihydroxy Vitamin D_3 is represented as 1,25OHD $_3$ R.

FIG. 4. Sequence of progressive *in vivo* bone development. Progressive repositioning of the vasculature from outside the stacked cell layer to a position in close approximation to the first layer of secretory osteoblasts responsible for formation of the first bony collar of the chick tibia (11,47,48). The osteoblast is oriented with its back toward the invading capillary and secretion of osteoid toward the cartilage core from the osteoblast's face. In this model, osteoblasts secrete osteoid in a direction away from vasculature (B), causing formation of a strut (C) and eventually forming the second layer of bone (D). These observations show that an intimate relationship exists between vasculature and newly forming bone.



BIOACTIVE FACTORS IN BONE

From the earliest days of modern humans, bone has been recognized to have the powerful capacity to repair discontinuities (22). A variety of bioactive factors combine in a complex multicellular, multi-step response in which reparative cells are specifically attracted to the repair site. These cells then aggregate, multiply, bridge the bone gap, and differentiate into chondrocytes or osteoblasts as controlled by the proximity to vasculature. Recently, an intensive research activity to identify and characterize these various bioactive factors was largely

successful (56,66,67,69). Our laboratory has described the purification of a protein factor, chondrogenic stimulating activity (CSA), which converts embryonic limb mesenchymal cells to chondrocytes (63,64). We are also attempting to purify a bone-derived chemoattractant for mesenchymal cells by using the now standard modified Boyden chamber (31,33).

Relevant to the thesis developed below, the identity and manipulation of the cells responding to bone-derived bioactive factors is directly related to successful bone repair. Such responding cells are present in the adult periosteum (36), dermis (49),

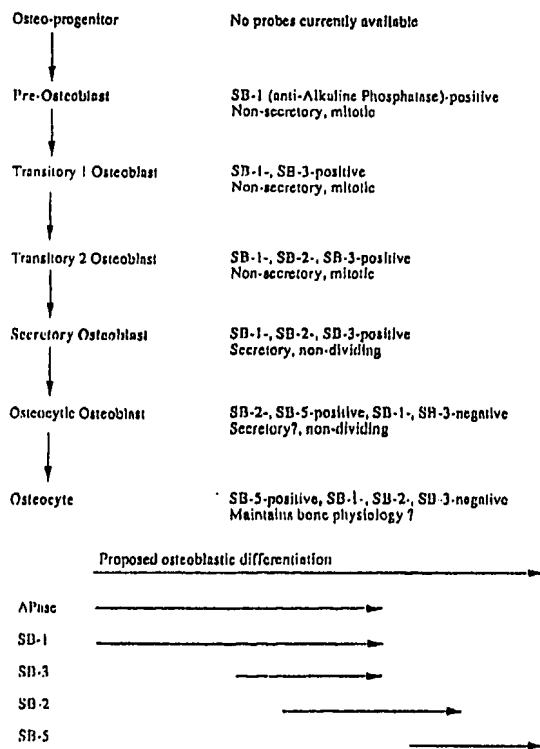


FIG. 5. Osteogenic cell lineage. Based on recent experimentation in which monoclonal antibodies were generated to cell surface antigens of osteogenic cells (3,4), a tentative lineage diagram reflects acquisition or loss of specific antigenic determinants. The characteristics of SB1, 2, and 3 were described previously (3); SB-5 (4) has been characterized and is similar to OB7.3 of Nijweide and Mulder (38). The individual lineage states are not weighted in terms of their prevalence or dwell-time; e.g., "transitory osteoblast 1" occurs rarely and cannot be recognized easily except at specific times and locations, whereas the "secretory osteoblast" is easily recognized and plentiful.

bone marrow (1,40,41,45), and connective tissue associated with muscle (34,37). One or all of these repositories are capable of forming bone when appropriately delivered bioactive factors are presented.

Alternately, when the responsive cells, stem cells, are placed in suitable delivery vehicles that can retain these cells while encouraging vascular invasion, bone can be observed to form. Recently, we used calcium phosphate porous ceramics in composite with marrow to encourage bone formation at both heterotopic and orthotopic sites (40,41). Whole disaggregated marrow cells in suspension are loaded into porous ceramic and transplanted to subcutaneous, intramuscular, or bone defect sites

in vivo. In 1-2 months, the few mesenchymal stem cells in the marrow have replicated massively and differentiated into osteoblasts. In the dead-end pores of the ceramic, which are devoid of vasculature, these stem cells differentiate into chondrocytes and form cartilage.

MESENCHYMAL STEM CELLS

From the above discussion several key facts are evident. First, embryonic mesenchymal stem cells in the limb which give rise to cartilage and bone in vivo can be manipulated in vitro. Second, these cells have a lineage progression of separate, individual steps, whether it be the chondrogenic or osteogenic pathway. Third, local cuing, sometimes involving highly potent protein factors, is responsible for providing positional information and causing lineage progression. Cell culture conditions have been refined to the extent that not only can these progressive events be studied in detail, but manipulation of the cells is also possible to provide control of tissue size and function.

Fourth, although chondrocytes and osteoblasts are derived from a common mesenchymal cell, the conditions for their initial differentiation and progression through the individual steps of their lineages are uniquely different. For example, osteogenesis is dependent on proximity to vasculature whereas chondrogenesis requires the complete absence of vasculature (7,10,11,16); osteogenesis is optimum at an initial cell culture seeding density in 35-mm dishes of 2×10^6 embryonic limb mesenchymal cells, whereas chondrogenesis is optimum at 5×10^6 cells (5,17,42).

Fifth, bone forms from mesenchymal stem cells in a cartilage-independent manner with vasculature providing a determinative discriminator between these two tissues; embryonic cartilage is not replaced by bone, but rather by vasculature and marrow (10,11,16). Sixth, we can demonstrate that three tissue sites are the repositories of mesenchymal stem cells: marrow (1,40,41,45), periosteum (36), and muscle connective tissue (34,37).

MARROW

Figure 6 outlines an assay to demonstrate that marrow contains mesenchymal stem cells capable of differentiation into cartilage and bone. Whole marrow is disrupted into single cells by passing it through needles of successively smaller sizes; the

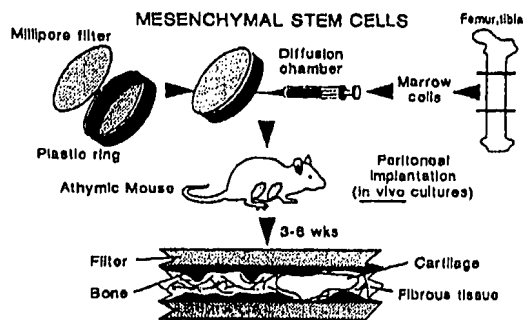


FIG. 6. Diffusion chamber assay in nude mice. Cell samples from marrow or other sources can be loaded into chambers composed of two Millipore filters glued to the edges of a plastic ring. These chambers are then implanted in the peritoneal cavity of athymic (nude) mice as a highly vascular in vivo incubation site. The filters prevent host cells from entering the chambers but permit rapid diffusion of nutrients and other factors into or out of the chamber. Histologic identification of two distinctive phenotypes, cartilage and bone, indicates that mesenchymal stem cells were present in the initial inoculum (1,2,45).

cells are counted, and $1-10 \times 10^6$ cells are placed in a small diffusion chamber (1,2,45). This chamber is of simple construction consisting of a small plastic ring onto which two Millipore filters have been glued. The filters allow body fluids (salts, nutrients, proteins, large protein complexes) to pass in and out of the chamber, but cells inside are not mixed with host cells, and tissues such as the vasculature are completely excluded. These chambers are implanted into the peritoneal cavity of an athymic (nude) mouse as an in vivo incubator, and they quickly become surrounded by host vasculature. Detailed studies have shown that the hematopoietic cells are eliminated, whereas mesenchymal cells vigorously divide and differentiate into cartilage in the middle of the chamber and bone at the filter interfaces closest to the enveloping vasculature (1,2,45). The presence of both cartilage and bone in the diffusion chamber has been compared to the presence of predominantly bone inside the highly vascularized pore regions of porous calcium phosphate ceramics loaded with marrow cells and implanted at heterotopic or orthotopic sites described above (40,41).

As a refinement of these experiments, we have been able to purify marrow mesenchymal cells by their differential adhesion to culture dishes and have successfully cultured cells through many passages (23). These cultured marrow mesenchymal cells from rat or chicken retain their capacity to differentiate into osteoblasts in ceramics through

such subculturing. Of importance is the demonstrated success of isolating marrow mesenchymal cells and mitotically expanding these cells with retention of their full developmental potency to differentiate into osteoblasts or chondrocytes.

Periosteum

Another repository for mesenchymal stem cells is the periosteum, a complex layer of cells that composes the outermost layer of long bone; we have termed the periosteum the stacked cell layer in developing embryos (1,16,47,48). This layer clearly responds to injury by rapidly expanding and forming woven bone; it also has cells capable of differentiating into chondrocytes when the periosteum is transplanted into an articular cartilage defect (39). In experimentation paralleling that described above for marrow mesenchymal cells, we have been successful in culturing and passaging periosteal cells (36). In porous ceramics implanted in nude mice, these cultured periosteal cells differentiate into osteoblasts (36). When the same cell preparation is injected into a subcutaneous site in a nude mouse, the cultured periosteal cells differentiate into both bone and cartilage (36). The important point is that culture-expanded periosteal cells retain their full developmental potency and can be manipulated to form two very complex and different tissues, bone or cartilage.

THE FUTURE: (SELF-CELL THERAPY)

Several important conceptual and technical advances have converged to allow us to consider the possibility of using a patient's own mesenchymal stem cells as starting material for tissue repair protocols. Mesenchymal stem cells must exist to maintain the living organisms, just as hematopoietic stem cells must exist to support both red and white blood cell turnover. Developmental biology has taught us that differentiated cells arise in a sequence of definitive cellular and molecular transitions, a lineage, from stem cell to end phenotype. Bone, for example, turns over; new osteoblasts arise, have a defined half-life, make new bone, and then die, to be replaced by other newly differentiating end-stage osteoblasts. Such osteoblasts must arise from stem cells; thus, a living organism must have repositories of stem cells.

Therefore, we might be able to isolate such human mesenchymal stem cells and place them in cell

culture, where we could mitotically expand their numbers. Eventually, if we had enough of these cells, we could reintroduce them into the original donor in a manner that guaranteed that they would massively differentiate into a specific tissue, such as cartilage or bone, at a transplantation or repair site. Immunorejection would not be a problem because the donor and host would be one and the same.

The first experimental step to test this idea is to determine if the animal-based technology described above can be modified to be used with human material. The first attempts at this have been highly encouraging. Recently, human marrow was introduced into diffusion chambers which were placed in nude mice; both cartilage and bone were eventually observed in the chamber (2). We recently cultured human marrow and isolated mesenchymal cells that were passaged, introduced into porous ceramics, and implanted subcutaneously in nude mice. In the pore regions of these highly vascularized composites, bone clearly formed in every sample of culture-expanded, marrow-derived mesenchymal cells tested (27). These preliminary experiments provide hope that the animal-based technology developed for mesenchymal cells from marrow or periosteum will be translatable to humans.

The concept of ex vivo manipulation of cells and their reimplantation into a donor is the basis for proposing self-cell therapy as a future possibility. Massive bone regeneration to fill gaps from tumor excision, regeneration of damaged articular cartilage, and maintenance of bone formation in the elderly at risk for osteoporosis are clinical protocols that require large numbers of the appropriate reparative skeletal cells. The patient's own mesenchymal stem cells may prove to be the basis of a new, cell-based treatment plan requiring the merging of molecular biology to produce specific bioactive factors, cell biology to develop ex vivo manipulation regimens, and surgeons able to implant cells capable of repairing skeletal defects by the regeneration process.

Acknowledgment: I thank the members of my laboratory, both past and present, for providing the fabric and labor of the cloth of our scientific pursuits. The resultant material of many colors provides both the backdrop and carpet for our scientific accomplishments and progress. My thanks are not enough to repay their kindnesses, contributions, and stimulation. This work was supported by grants from the NIH.

REFERENCES

1. Bab I, Howlett CR, Ashton BA, Owen ME: Ultrastructure of bone and cartilage formed in vitro in diffusion chambers. *Clin Orthop* 187:243-254, 1984
2. Bab I, Passi-Even L, Gazit D, Sekeles E, Ashton BA, Peylan-Ramu N, Ziv I, Ulmansky M: Osteogenesis in in vivo diffusion chamber cultures of human marrow cells. *Bone Mineral* 4:373-386, 1988
3. Bruder SP, Caplan AI: First bone formation and the dissection of an osteogenic lineage in the embryonic chick tibia is revealed by monoclonal antibodies against osteoblasts. *Bone* 10:359-375, 1989
4. Bruder SP, Caplan AI: Terminal differentiation of osteogenic cells in the embryonic chick tibia is revealed by a monoclonal antibody against osteocytes. *Bone* 11:189-198, 1990
5. Caplan AI: Effects of the nicotinamide-sensitive teratogen 3-acetylpyridine on chick limb cells in culture. *Exp Cell Res* 62:341-355, 1970
6. Caplan AI: Muscle, cartilage and bone development and differentiation from chick limb mesenchymal cells. In: *Vertebrate Limb and Somite Morphogenesis*, ed by DA Ede, JR Hinchliffe, and M Balls, Cambridge, England, Cambridge University Press, 1977, pp 199-213
7. Caplan AI: The molecular control of muscle and cartilage development. In: *39th Annual Symposium of the Society for Developmental Biology*, ed by S Subtelney, U Abbott, New York, Alan R. Liss, 1981, pp 37-68
8. Caplan AI: Cartilage. *Scientific American* 251:84-94, 1984
9. Caplan AI: The extracellular matrix is instructive. *BioEssays* 5:129-132, 1986
10. Caplan AI: Bone development and repair. *BioEssays* 6:171-175, 1987
11. Caplan AI: Bone development. In: Wiley J, ed. *Cell and Molecular Biology of Vertebrate Hard Tissues*. Chichester, CIBA Foundation Symposium 136. 1988:3-21.
12. Caplan AI, Fiszman MY, Eppenberger HM: Molecular and cell isoforms during development. *Science* 221:921-927, 1983
13. Caplan AI, Hascall VC: Structure and developmental changes in proteoglycans. In: *Dilatation of the Uterine Cervix*, ed by F Naftolin, and PG Stubblefield, New York, Raven Press, 1980, pp 79-98
14. Caplan AI, Koutropas S: The control of muscle and cartilage development in the chick limb: The role of differential vascularization. *J Embryol Exp Morphol* 29:571-583, 1973
15. Caplan AI, Ordahl CP: Irreversible gene repression model for control of development. *Science* 201:120-130, 1978
16. Caplan AI, Pechak DG: The cellular and molecular embryology of bone formation. In: *Bone and Mineral Research*, vol. 5, ed by WA Peck, Elsevier, New York, 1987, pp. 117-184
17. Caplan AI, Zwilling E, Kaplan NO: 3-Acetylpyridine: effects in vitro related to teratogenic activity in chicken embryos. *Science* 160:1009-1010, 1968
18. Carrino DA, Kujawa MJ, Lennon DP, Caplan AI: Altered proteoglycans synthesized by chick limb bud chondrocytes cultured in serum-free defined medium. *Exp Cell Res* 183:62-71, 1989
19. Cruz-Orive L-M, Hunziker EB: Stereology for anisotropic cells: Application to growth cartilage. *J Microsc* 143:47-80, 1986
20. DeLuca S, Heinegard D, Hascall VC, Kimura JH, Caplan AI: Chemical and physical changes in proteoglycans during development of chick limb bud chondrocytes grown in vitro. *J Biol Chem* 252:6600-6608, 1977

21. Dexter TM, Spooner E: Growth and differentiation in the hematopoietic system. *Annu Rev Cell Biol* 3:423-441, 1987
22. Goldberg K, and editors of U.S. News: The human body: The skeletal-fantastic framework. *U.S. News* 1982
23. Goshima J, Goldberg VM, Caplan AI: The osteogenic potential of culture-expanded rat marrow mesenchymal cells as assayed in vivo in calcium phosphate ceramic blocks. *Clin Orthop Related Res* 262:298-311, 1991
24. Grotendorst GR, Martin GR: Cell movements in wound healing and fibrosis. *Rheumatology* 10:385-403, 1986
25. Hauschka SD: Clonal analysis of vertebrate myogenesis, III. Developmental changes in the muscle colony-forming cells of the human fetal limb. *Dev Biol* 37:345-368, 1974
26. Haynesworth SE, Carrino DA, Caplan AI: Comparison of the cartilage proteoglycan core protein synthesized by chondrocytes of different ages. *Connect Tissue Res* 25:311-320, 1991
27. Haynesworth SE, Goshima J, Goldberg VM, Caplan AI: Isolation and expansion of cells with osteogenic potential from human marrow. *Bone* (submitted for publication)
28. Hunziker EB: Growth plate structure and function. *Pathol Immunopathol Res* 7:9-13, 1988
29. Itay S, Abramovici A, Nevo Z: Use of cultured embryonic chick epiphyseal chondrocytes as grafts for defects in chick articular cartilage. *Clin Orthop* 220:284-303, 1987
30. Kujawa MJ, Lennon DP, Caplan AI: Growth and differentiation of stage 24 limb mesenchymal cells in a serum-free, chemically defined medium. *Exp Cell Res* 183:45-61, 1989
31. Lucas PA, Caplan AI: Chemotactic response of embryonic limb bud mesenchymal cells and muscle-derived fibroblasts to transforming growth factor beta. *Connect Tissue Res* 18:1-7, 1988
32. Lucas PA, Price PA, Caplan AI: Chemotactic response of mesenchymal cells, fibroblasts and osteoblast-like cells to bone GLA protein. *Bone* 9:319-323, 1988
33. Lucas PA, Syftestad GT, Caplan AI: Partial isolation and characterization of a chemotactic factor from adult bone for mesenchymal cells. *Bone* 7:365-371, 1986
34. Lucas PA, Syftestad GT, Caplan AI: A water-soluble fraction from adult bone stimulates the differentiation of cartilage in explants of embryonic muscle. *Differentiation* 37:47-52, 1988
35. Manduca P, Castagnola P, Cancedda R: Dimethyl sulfoxide interferes with in vitro differentiation of chick embryo endochondral chondrocytes. *Dev Biol* 125:234-236, 1988
36. Nakahara H, Bruder SP, Goldberg VM, Caplan AI: In vivo osteochondrogenic potential of cultured cells derived from the periosteum. *Clin Orthop Rel Res* 259:223-232, 1990
37. Nathanson MA, Hay ED: Analysis of cartilage differentiation from skeletal muscle grown in bone matrix I. Ultrastructural aspects. *Dev Biol* 78:301-331, 1980
38. Nijweide P, Mulder P: Identification of osteocytes in osteoblast-like cell cultures using a monoclonal antibody specifically directed against osteocytes. *Histochemistry* 84:342-347, 1986
39. O'Driscoll SW, Salter RB: The induction of neochondrogenesis in free intra-articular periosteal autografts under the influence of continuous passive motion. *J Bone Joint Surg [Am]* 66:1248, 1984
40. Ohgushi H, Goldberg VM, Caplan AI: Heterotopic osteogenesis in porous ceramics induced by marrow cells. *J Orthop Res* 7:568-578, 1989
41. Ohgushi H, Goldberg VM, Caplan AI: Repair of segmental long bone defect by composite graft of marrow cells and porous calcium phosphate ceramic. *Acta Orthop Scand* 60:334-339, 1989
42. Osdoby P, Caplan AI: Osteogenesis in cultures of limb mesenchymal cells. *Dev Biol* 73:84-102, 1979
43. Osdoby P, Caplan AI: Characterization of bone-specific alkaline phosphatase in cultures of chick limb mesenchymal cells. *Dev Biol* 86:136-146, 1981
44. Osdoby P, Caplan AI: First bone formation in embryonic chick limbs. *Dev Biol* 86:147-156, 1981
45. Owen M: Lineage of osteogenic cells and their relationship to the stromal system. In: *Bone and Mineral Research*, Vol. 3, ed by WA Peck, New York, Elsevier, 1985, pp 1-25
46. Owens EM, Solursh M: In vitro histogenic capacities of limb mesenchyme from various stage mouse embryos. *Dev Biol* 88:297-311, 1981
47. Pechak DG, Kujawa MJ, Caplan AI: Morphological and histochemical events during first bone remodeling in embryonic chick limbs. *Bone* 7:441-458, 1986
48. Pechak DG, Kujawa MJ, Caplan AI: Morphology of bone development and bone remodeling in embryonic chick limbs. *Bone* 7:459-472, 1986
49. Reddi AH: Cell biology and biochemistry of endochondral bone development. *Cell Rel Res* 1:209-226, 1981
50. Roughley PJ, White RJ: Age-related changes in the structure of the proteoglycan subunits from human articular cartilage. *J Biol Chem* 255:217-224, 1980
51. Sachs L: Hematopoietic growth and differentiation factors after the reversal of malignancy. In: *Tumor Cell Differentiation*, ed by J Aarbakke, PK Chiang, and HP Koeffler, Clifton, NJ, Humana Press, 1987, pp 3-27
52. Sachs L: The molecular control of blood cell development. *Science* 238:1374-1379, 1987
53. San Antonio JD, Tunn RS: Chondrogenesis of limb bud mesenchyme in vitro: stimulation by cations. *Dev Biol* 115:313-324, 1986
54. Schmid TM, Conrad HE: Metabolism of low molecular weight collagen by chondrocytes obtained from histologically distinct zones of the chick embryo tibiotarsus. *J Biol Chem* 257:12451-12457, 1982
55. Schmid TM, Linsenmayer TF: Developmental acquisition of type X collagen in the embryonic chick tibiotarsus. *Dev Biol* 107:373-381, 1985
56. Seyedin SM, Thomas TC, Thompson AY, Rosen DM, Piez KA: Purification and characterization of two cartilage-inducing factors from bovine demineralized bone. *Proc Natl Acad Sci USA* 82:2267-2271, 1985
57. Slavkin HC: Molecular biology of dental development: a review. In: *The Biological Mechanism of Tooth Eruption and Root Resorption*, ed by Z Davidovitch, EbscoMed, Birmingham, AL, 1988, pp 107-116
58. Solursh M, Reiter RL, Ahrens FB, Vertel BM: Stage- and position-related changes in chondrogenic response of chick embryonic wing mesenchyme to treatment with dibutyryl cyclic AMP. *Dev Biol* 83:9-19, 1981
59. Somerman M, Hewitt AT, Varner HH, Schiffman E, Termine J, Reddi AH: Identification of a bone matrix-derived chemotactic factor. *Calcif Tissue Int* 35:481-485, 1983
60. Sulston J, Schierenberg E, White J, Thomson J: The embryonic cell lineage of the nematode *Caenorhabditis elegans*. *Dev Biol* 100:64-119, 1983
61. Swalla BJ, Solursh M: Inhibition of limb chondrogenesis by fibronectin. *Differentiation* 26:42-48, 1984
62. Sweet MBE, Thonar EJ-MA, Marsh J: Age-related changes in proteoglycan structure. *Arch Biochem Biophys* 198:439-448, 1979
63. Syftestad GT, Caplan AI: A fraction from extracts from demineralized bone stimulates the conversion of mesenchymal cells into chondrocytes. *Dev Biol* 104:348-356, 1984

64. Syfkestad GT, Lucas PA, Caplan AI: The in vitro chondrogenic response of limb bud mesenchyme to a water soluble fraction prepared from demineralized bone matrix. *Differentiation* 29:230-237, 1985
65. Syfkestad GT, Weitzhandler M, Caplan AI: Isolation and characterization of osteogenic cells derived from first bone of the embryonic tibia. *Dev Biol* 110:275-283, 1985
66. Takaoka K, Ono K, Ametani K, Kishimoto R, Nakata Y: Solubilization and concentration of bone-inducing substance from a murine osteosarcoma. *Clin Orthop* 148:274-280, 1980
67. Urist MR, Delange RJ, Finerman GAM: Bone cell differentiation and growth factors. *Science* 220:680-686, 1983
68. von der Mark K, Osdoby P, Caplan AI: Effect of 4-methylumbellifer-yl- β -D-xyloside on collagen synthesis in chick limb bud mesenchymal cell cultures. *Dev Biol* 90:24-30, 1982
69. Wozney JM, Rosen V, Celeste AJ, Mitsock LM, Whitters MJ, Kriz RW, Kewick RM, Wang EA: Novel regulators of bone formation: Molecular clones and activities. *Science* 242:1528-1534, 1988
70. Zipori D, Lee F: Introduction of interleukin-3 gene into stromal cells from the bone marrow alters hematopoietic differentiation but does not modify stem cell renewal. *Blood* 71:586-596, 1988
71. Zull JE, Krug S, Abel D, Caplan AI: Development of parathyroid hormone and calcitonin-activated adenylate cyclases in the embryonic limb and in cultured cells from embryonic chick limb. *Proc Natl Acad Sci USA* 75:3871-3875, 1978
72. Zull JE, Youngman K, Caplan AI: The development of hormonal responses of cultured embryonic chick limb mesenchymal cells. *Dev Biol* 86:61-68, 1981

EVIDENCE APPENDIX

Item 13

**The Journal**• **Current Issue**

Contents in full
Talking points
Analysis and
Interpretation
Primary Research
Review and Opinion

• Supplements
• Back Issues

[Search Journal](#)

[Subscribe](#)

[E-Toc](#)

[Log Out](#)

• **Jobs**

• Awards and
Announcements
• Conferences
• Press Services
• General Info
• Free Sample
• Contact Us

THE LANCET
Infectious Diseases

THE LANCET
Neurology

THE LANCET
Oncology

[Home](#) > [The Journal](#) > [Current Issue](#) > [Primary Research](#)

Volume 364, Number 9429
10 July 2004

**Articles**

Intracoronary autologous bone-marrow cell transfer after myocardial infarction: the BOOST randomised controlled clinical trial

Kai C Wollert, Gerd P Meyer, Joachim Lotz, Stefanie Ringes-Lichtenberg, Peter Lippolt, Christiane Breidenbach, Stephanie Fichtner, Thomas Korte, Burkhard Homig, Diethelm Messinger, Lubomir Arseniev, Bernd Hertenstein, Arnold Ganzer, Helmut Drexler

Lancet 2004; 364: 141-48

[See Comment](#)

Departments of Cardiology and Angiology (K C Wollert MD, G P Meyer MD, S Ringes-Lichtenberg MD, P Lippolt MD, C Breidenbach MD, S Fichtner BS, T Korte MD, B Homig MD, Prof H Drexler MD), **Diagnostic Radiology** (J Lotz MD), and **Haematology and Oncology** (L Arseniev MD, B Hertenstein MD, Prof A Ganzer MD), **Hanover Medical School, Hanover, Germany; and IST, Mannheim, Germany** (D Messinger MSc)

Correspondence to: Prof Helmut Drexler, Abteilung Kardiologie und Angiologie, Medizinische Hochschule Hannover, Carl-Neuberg Strasse 1, 30625 Hannover, Germany
drexler.helmut@mh-hannover.de

[Summary](#)
[Introduction](#)
[Methods](#)
[Results](#)
[Discussion](#)

Summary

Background Emerging evidence suggests that stem cells and progenitor cells derived from bone marrow can be used to improve cardiac function in patients after acute myocardial infarction. In this randomised trial, we aimed to assess whether intracoronary transfer of autologous bone-marrow cells could improve global left-ventricular ejection fraction (LVEF) at 6 months' follow-up.

Methods After successful percutaneous coronary intervention (PCI) for acute ST-segment elevation myocardial infarction, 60 patients were randomly assigned to either a control group (n=30) that received optimum postinfarction medical treatment, or a bone-marrow-cell group (n=30) that received optimum medical treatment and intracoronary transfer of autologous bone-marrow cells 4-8 days (SD 1.3) after PCI. Primary endpoint was global left-ventricular ejection fraction (LVEF) change from baseline to 6 months' follow-up, as determined by cardiac MRI. Image analyses were done by two investigators blinded for treatment assignment. Analysis was per protocol.

Findings Global LVEF at baseline (determined 3-5 days [SD 1.5] after PCI) was 51.3 (9.3%) in controls and 50.0 (10.0%) in the bone-marrow cell group (p=0.59). After 6 months, mean global LVEF had increased by 0.7 percentage points in the control group and 6.7 percentage points in the bone-marrow-cell group (p=0.0026). Transfer of bone-marrow cells enhanced left-ventricular systolic function primarily in myocardial segments adjacent to the infarcted area. Cell transfer did not increase the risk of adverse clinical events, in-stent restenosis, or proarrhythmic effects.

Interpretation Intracoronary transfer of autologous bone-marrow-cells promotes improvement of left-ventricular systolic function in patients after acute myocardial infarction.

▲ top

Introduction

Rapid reperfusion of the infarct-related coronary artery is of great importance in salvaging ischaemic myocardium and limiting the infarct size in patients with acute myocardial infarction. When done expeditiously and expertly, percutaneous transluminal coronary angioplasty with stent implantation is the method of choice to re-establish coronary flow.¹ Unfortunately, myocardial necrosis starts rapidly after coronary occlusion, usually before reperfusion can be achieved.² The loss of viable myocardium initiates a process of adverse left-ventricular remodelling, leading to chamber dilatation and contractile dysfunction in many patients.³ In this context, much interest has followed from experimental studies showing that cardiac transfer of unfractionated bone-marrow cells, or stem cells and progenitor cells derived from bone marrow can enhance functional recovery after acute myocardial infarction.^{4,5} Based on these data, stem cells and progenitor cells derived from bone marrow have been proposed for use in the repair of cardiac tissue after acute myocardial infarction in patients.⁶⁻⁸

Early clinical investigations indicate that infusion of autologous bone-marrow cells into the infarct-related coronary artery is feasible after acute myocardial infarction.^{9,10} However, because these studies were not randomised trials, the efficacy of intracoronary transfer of bone-marrow cells for functional recovery after acute myocardial infarction in patients has remained uncertain. We did a randomised controlled trial to assess the effect of intracoronary transfer of autologous bone-marrow cells on left-ventricular functional recovery in patients after acute myocardial infarction and successful percutaneous coronary intervention (PCI).

▲ top

Methods

Patients

Patients were eligible if they were admitted within 5 days of the onset of symptoms of a first ST-segment elevation myocardial infarction, had undergone successful PCI with stent implantation in the infarct-related artery, and had hypokinesia or akinesia involving more than two thirds of the left-ventricular anteroseptal, lateral, and/or inferior wall, as shown by angiography done immediately after PCI. We excluded patients who had multivessel coronary artery disease, pulmonary oedema, cardiogenic shock, advanced renal or hepatic dysfunction, or documented terminal illness or cancer.

This randomised-controlled study of BOne marrOw transfer to enhance ST-elevation infarct regeneration (the BOOST trial) was approved by our local Ethics Committee. Patients provided written informed consent.

Randomisation and baseline cardiac MRI

Patients were randomly allocated in a 1:1 ratio to either the control or bone-marrow-cell groups, with use of sequentially numbered, sealed envelopes provided by IST (DM). After randomisation, all patients underwent cardiac MRI.

Harvest and transfer of bone-marrow cells

After baseline cardiac MRI, bone marrow was harvested from patients in the bone-marrow-cell group. Bone marrow was processed by 4% gelatine-polysuccinate density gradient sedimentation according to current Good Manufacturing Practice (GMP) regulations (Cytonet, Hannover, Germany), to reduce the volume of the preparation and to deplete erythrocytes and platelets. The final suspension of bone-marrow cells was washed and resuspended in saline with 10 000 U/L heparin.

We used an automated haemocytometer to measure the number of nucleated cells, packed-cell volume, and platelet count in the initial bone marrow aspirate and in the final preparation of bone-marrow cells. Nucleated cell viability was assessed by trypan blue exclusion. We measured the number of CD34+ cells with flow cytometry analysis (FACSCalibur, BD Biosciences, Heidelberg, Germany) using an antibody from Beckman Coulter (Krefeld, Germany). Haemopoietic colony-forming cell growth was measured by a methylcellulose assay (StemCell Technologies, St Katharinen, Germany).

6-8 h after bone-marrow harvest, the final preparation of bone-marrow cells was infused into the infarct-related artery via the central lumen of an over-the-wire balloon catheter (Concerto, Occam International, Eindhoven, Netherlands). To allow bone-marrow cells maximum contact time with the microcirculation of the infarct-related artery, the balloon was inflated inside the stent to transiently interrupt antegrade blood flow during infusions. The entire bone-marrow-cell preparation was infused during four to five coronary occlusions, each lasting 2.5-4 min. Between occlusions, the coronary artery was reperfused for 3 min.

Follow-up

All patients were treated with aspirin (300 mg daily for 4 weeks after PCI, then 100 mg daily), clopidogrel (300 mg loading dose, then 75 mg daily for at least 4 weeks after PCI), an angiotensin-converting enzyme (ACE) inhibitor or angiotensin-receptor blocker, a β blocker, and a statin (if LDL

cholesterol concentrations were above 2.6 mmol/L), unless these agents were contraindicated. At both 6 weeks and 3 months after discharge, patients had follow-up examinations to assess their clinical status and to review their current medication. Where necessary, dosages of angiotensin-converting enzyme inhibitors (ACE-inhibitors), angiotensin-receptor blockers, β blockers, and statins were adjusted in accordance with current practice guidelines.^{11,12} 6 months after discharge, cardiac MRI was repeated in all patients. In addition, patients were scheduled to undergo coronary angiography to assess the degree of restenosis in the stented segment of the infarct-related artery. Restenosis was quantified with a computer-based system (CMS, Medical Imaging Systems, Leiden, Netherlands) by an investigator unaware of treatment assignment (AM).

To assess whether intracoronary bone-marrow-cell transfer was associated with proarrhythmic effects, we obtained 24 h Holter recordings from all patients before hospital discharge, and at 6 weeks', 3 months', and 6 months' follow-up. From these recordings, the mean number of premature ventricular complexes per h was calculated. We also recorded the number of non-sustained and sustained ventricular tachycardias per recording. In addition, patients were scheduled to undergo programmed ventricular stimulation at 6 months' follow-up. Ventricular stimulation was done at the right-ventricular apex and the right-ventricular outflow tract with single, double, and triple extra stimuli at twice the diastolic threshold and basic cycle lengths of 500 ms and 400 ms.

Cardiac MRI

Cardiac MRI was done with the patient in supine position in a 1.5-T scanner (CV/i, General Electric, Munich, Germany) using electrocardiogram (ECG) gating and a four-element phased array receiver coil. To measure left-ventricular volumes, we used repeated breath-hold fast gradient echo sequences in a steady state (FIESTA, General Electric). Sequence parameters were as follows: TR/TE 3.8/1.6 ms, 40° flip angle, 224x224 matrix, field of view 36-38 cm, in-plane resolution 1.6x1.6-1.7x1.7 mm, 38-40 phases per RR-interval, 10 mm slice thickness. An end-diastolic, horizontal long-axis plane of the left ventricle at end-expiration provided the reference image on which a stack of contiguous short-axis slices was positioned to cover the entire left ventricle.

Contrast-enhanced MRI was used to assess myocardial injury after acute myocardial infarction.¹³ A breath-hold k-space segmented T1 weighted inversion recovery gradient echo sequence was used to cover the entire left ventricle with 7-8 mm short-axis slices as described above (TR/TE 7.1/3.1 ms, 256x192 matrix, field of view 36-38 cm, in-plane resolution 1.4x1.9-1.5x2.0 mm). Inversion time (200-220 ms) was individually adapted to null the signal of the myocardium. End-diastolic images were obtained starting 15 min after an intravenous bolus injection of 0.15 mmol/kg gadobutrol, a gadolinium-based extracellular contrast agent (Schering, Berlin, Germany).

All image analyses were done by two investigators who were unaware of treatment assignment (CB and SF), using the MASS 4.0.1 software (Medical Imaging Systems). Endocardial and epicardial borders were traced in all end-diastolic and end-systolic short-axis slices to determine left-ventricular end-diastolic volumes (LVEDV) and end-systolic volumes (LVESV) for global and regional calculation of left-ventricular ejection fraction (LVEF), and left-ventricular mass. For assessment of infarct volumes, late contrast enhancement was quantified. LVEDV index, LVESV index, and left-ventricular-mass index were calculated by dividing LVEDV, LVESV, and left-ventricular mass by body surface area. Regional LVEF was derived by calculating LVEF only in slices showing late contrast enhancement at

baseline. Regional left-ventricular function was assessed by determining systolic wall motion in the infarct region and border zone. Systolic-wall motion was defined as the radial displacement of the endocardial contour at systole. Myocardial segments showing late contrast enhancement at baseline were defined as the infarct region. Segments adjacent to the infarct region were defined as the border zone.

Statistical analysis

Primary endpoint was the change from baseline in global LVEF at 6 months' follow-up. Secondary endpoints were changes in LVEDV index, LVESV index, left-ventricular-mass index, and late contrast enhancement. We calculated that we would need 30 patients in each group to achieve a power of at least 80% to detect a difference in global LVEF change of 5 percentage points between study groups, with a two-sided significance level of $p < 0.05$, and a common standard deviation of 6.5 percentage points for the global LVEF change from baseline to 6 months' follow-up. We used ANCOVA to compare global LVEF changes in the two study groups, with bone-marrow-cell treatment as the main factor and LVEF at baseline as a covariate. To estimate the treatment effect, differences in least-squares means and corresponding 95% CI were calculated based on the ANCOVA model. We analysed secondary endpoints using the same methods. The consistency of the treatment effect on global LVEF change was assessed across several subgroups. All statistical tests were two-sided with a significance level of $p < 0.05$.

Homogeneity of treatment groups at baseline was assessed using Student's *t* test for continuous variables showing no marked deviations from the normal distribution. For other continuous variables or ordinal baseline data, the Wilcoxon rank-sum test was used. Categorical baseline data were investigated using χ^2 tests. The relation between the number of nucleated cells, CD34+ cells, and haemopoietic colony-forming cells infused into the infarct-related coronary artery and subsequent global LVEF changes were assessed with Pearson's correlation coefficient. Subgroup analyses were not prespecified but were exploratory in nature. All subgroup analyses are reported.

▲ top

Results

Between January, 2002, and May, 2003, 78 patients were informed about the trial. 65 patients were randomly allocated to treatment. After randomisation, five patients were withdrawn because they could not undergo cardiac MRI, either because of claustrophobia or severe obesity. The final cohort included 30 controls and 30 patients in the bone-marrow-cell group (figure 1). Table 1 shows patients' baseline characteristics. All patients received optimum postinfarction medical treatment (table 1).

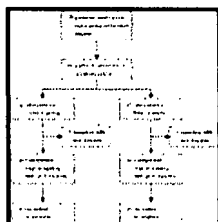


Figure 1: Trial profile

BMC=bone-marrow cell.

	Control group (n=30)	BMC group (n=30)	p
Age (years)	59.2 (13.5)	53.4 (14.8)	0.11
Men	22 (73%)	20 (67%)	0.57
Body-mass index (kg/m ²)	26.2 (4.2)	25.8 (3.0)	0.67
Diabetes mellitus	3 (10%)	3 (10%)	1.0
Hyperlipidaemia*	7	9	0.56
Hypertension	13 (43%)	9 (30%)	0.28
Current cigarette use (number of patients)	17 (57%)	18 (60%)	0.79
Median time from symptom onset to PCI (h) (range)	8.0 (3-120)	9.8 (2-22)	0.92
Killip class			
1	25 (83%)	23 (77%)	0.51
2	5 (17%)	7 (23%)	
3 or 4	0	0	
Infarct-related artery			
Right coronary artery	7 (23%)	7 (23%)	1.0
Left coronary artery	23 (77%)	23 (77%)	
TIMI flow grade before PCI:			0.73
Grade 0 or I	16 (53%)	13 (43%)	
Grade II	13 (43%)	16 (53%)	
Grade III	1 (3%)	1 (3%)	
after PCI:			0.75
Grade 0 or I	0	0	
Grade II	7 (23%)	6 (20%)	
Grade III	23 (77%)	24 (80%)	
Maximum serum creatine kinase concentration (U/L)	2844 (1161)	2968 (1867)	0.77
Maximum serum creatine kinase MB concentration (U/L)	156 (51)	175 (123)	0.46
Maximum serum troponin T concentration (µg/L)	7.4 (4.4)	7.4 (5.5)	0.99
Periprocedural therapy			
Thrombolytic therapy before PCI	10 (33%)	14 (47%)	0.29
Platelet glycoprotein IIb/IIIa inhibitors	1.0	14 (47%)	
Median number of stents (range)	1 (1-5)	1 (1-2)	0.40
Size of stent (mm)	3.3 (0.4)	3.3 (0.4)	1.0
Length of stent (mm)	17.5 (9.6)	17.6 (6.4)	0.97
Lesion characteristics			0.71
Type A	8 (27%)	6 (20%)	
Type B	16 (53%)	19 (64%)	
Type C	6 (20%)	5 (3%)	
Medication at primary discharge:			
Aspirin† and clopidogrel	29 (97%)	30 (100%)	
ACE-inhibitors or angiotensin-	30 (100%)	30 (100%)	

receptor blockers		
β blockers	30 (100%)	29 (97%)
Statins	29 (97%)	30 (100%)
at 6 months' follow-up:		
Aspirin†	27 (97%)	29 (97%)
ACE-inhibitors or angiotensin-receptor blockers	30 (100%)	30 (100%)
β blockers	30 (100%)	29 (97%)
Statins	28 (93%)	28 (93%)

BMC=bone-marrow cell. ACE=angiotensin-converting enzyme. Data are means (SD) or n (%) unless otherwise stated. *Serum cholesterol >5.2 mmol/L. †Patients not receiving aspirin were treated with phenprocoumon.

Table 1: Patients' characteristics

Mean time from PCI to baseline cardiac MRI was 3.5 days (SD 1.5). Mean time from PCI to bone-marrow harvest was 4.8 days (1.3). Time from symptom onset to harvest of bone-marrow cells was 5.7 days (1.2). On average, 128 mL (33) of bone marrow was aspirated from the posterior iliac crest during a brief general anaesthesia with midazolam and etomidate. No bleeding complications at the harvest site were noted.

During preparation of bone-marrow cells, the sedimentation process reduced the volume of bone-marrow cells to a mean of 26 mL (SD 4) and recovered 75% (12) of nucleated cells from the initial bone-marrow aspirate. The final preparation of bone-marrow cells contained 24.6×10^8 (SD 9.4×10^8) nucleated cells (viability 99% [2]), 9.5×10^6 (6.3×10^6) CD34+ cells, and 3.6×10^6 (3.4×10^6) haemopoietic colony-forming cells. The packed cell volume of the final bone-marrow-cell preparation was 31% (11), and the platelet count was 182×10^6 (93×10^6) per mL.

Changes of LVEDV index, LVESV index, left-ventricular-mass index, and late-contrast enhancement from baseline to 6 months' follow-up did not differ significantly between the control and bone-marrow-cell groups (table 2). The increase in LVEDV index at 6 months was slightly higher in the bone-marrow-cell group, whereas LVESV index tended to decrease more in the bone-marrow-cell group (table 2). 6 months after randomisation, global LVEF increased significantly in the bone-marrow-cell group compared with controls ($p=0.0026$) (table 2 and figure 2). The effects of bone-marrow-cell transfer on global LVEF change at 6 months' follow-up were consistent in all investigated subgroups (figure 3). The improvement in global LVEF after 6 months' follow-up was not correlated with the number of nucleated cells ($r=-0.11$, $p=0.57$), CD34+ cells ($r=0.13$, $p=0.48$), or haemopoietic colony-forming cells ($r=-0.14$, $p=0.46$) infused into the infarct-related coronary artery.

	Baseline		6 months		Change		BMC treatment effect*	p
	Controls	BMC group	Controls	BMC group	Controls	BMC group		
LVEDV index (mL/m ²)	81.4 (16.9)	84.2 (17.2)	84.9 (21.9)	91.7 (26.0)	3.4 (11.1)	7.6 (20.0)	4.0 (-4.4 to 12.5)	0.32
LVESV index	40.6 (16.9)	43.0 (14.7)	42.6 (23.5)	42.4 (23.9)	2.0 (11.1)	-0.6 (14.9)	-3.2 (-9.7 to 3.3)	0.33

(mL/m ²)								
Global LVEF (%)	51.3 (9.3)	50.0 (10.0)	52.0 (12.4)	56.7 (12.5)	0.7 (8.1)	6.7 (6.5)	6.0 (2.2 to 9.9)	0.0026
LVM index (g/m ²)	78.2 (18.3)	82.7 (18.7)	71.7 (14.2)	71.9 (14.6)	-6.5 (12.8)	-10.8 (10.6)	-2.5 (-7.3 to 2.3)	0.30
LE (mL)	30.3 (17.4)	33.0 (21.1)	19.8 (9.8)	18.9 (12.2)	-10.5 (10.6)	-14.1 (13.0)	-2.2 (-5.4 to 1.0)	0.18

BMC=bone-marrow cell. Data are mean (SD) unless otherwise stated.

*Treatment effects expressed as differences in least-squares means (ANCOVA model) with 95% CI. LVM=left ventricular mass. LE=late contrast enhancement. There were no differences between groups at baseline.

Table 2: Left ventricular volume and mass indices, global LVEF, and late enhancement as determined by contrast-enhanced MRI at baseline and 6 months' follow-up



Figure 2: Global LVEF at baseline and 6 months' follow-up

*p=0.0026 for difference between groups. Small dots show data for individual patients; large dots show mean values. Vertical bars show SD.

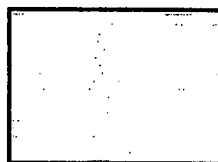


Figure 3: Subgroup analyses of global LVEF changes from baseline at 6 months' follow-up

LE=late contrast enhancement. BMC=bone-marrow cell. *Median values of the whole study population were used to create subgroups of equal size. †Cardiovascular risk factors were diabetes, total cholesterol concentration greater than 5.2 mmol/L, hypertension, or current smoking. Oval dots show differences of least-squares means between groups; horizontal bars show 95% CI.

Compared with the control group, patients in the bone-marrow-cell group had increased regional LVEF (p=0.04) and systolic wall motion in the border zone (p=0.03) at 6 months. By contrast, systolic wall motion in the infarct region was not significantly enhanced by transfer of bone-marrow-cells (table 3). Representative colour-coded images showing the effects of bone-marrow-cell transfer on left-ventricular function are shown in figure 4.

	Baseline		6 months		Change		BMC treatment	p
	Controls	BMC group	Controls	BMC group	Controls	BMC group	effect†	
Regional LVEF	47.8 (9.7)	46.3 (10.6)	48.9 (15.2)	53.0 (15.5)	1.1 (11.8)	6.7 (9.5)	5.7 (0.2 to 11.3)	0.04

(%)									
Systolic wall motion (mm),	3.9 (1.8)	4.4 (1.9)	4.9 (2.9)	5.9 (2.5)	1.0 (2.5)	1.5 (2.1)	0.6 (-0.6 to 1.8)	0.32	
infarct region									
Systolic wall motion (mm), border zone	6.8 (1.6)	7.0 (1.7)	6.8 (2.1)	8.0 (2.1)	-0.1 (2.2)	1.0 (1.9)	1.1 (0.1 to 2.1)	0.03	

BMC=bone-marrow cell. Data are mean (SD). Treatment effects are expressed as differences in least-squares means (ANCOVA model) and 95% CI. There were no differences between groups at baseline.

Table 3: Regional LVEF and systolic wall motion as determined by contrast-enhanced MRI at baseline and 6 months' follow-up

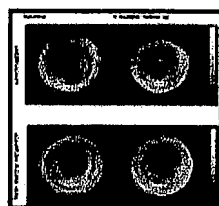


Figure 4: Representative colour-coded images showing systolic wall motion at baseline and 6 months' follow-up in two patients

Both patients had had an anterior acute myocardial infarction. Bright colours indicate good systolic wall motion, whereas dark colours indicate poor wall motion (expressed in mm). Note improved functional recovery in the patient treated with bone-marrow-cells.

No patient died or was lost to follow-up. There were no increases in troponin T concentrations in serum in any of the patients 24 h after intracoronary transfer of bone-marrow cells, indicating that the procedure did not inflict additional ischaemic damage to the myocardium. In 6 months of follow-up, three controls and one patient from the bone-marrow-cell group needed at least one hospital admission for worsening heart failure. One person from the bone-marrow-cell group developed a non ST-segment elevation myocardial infarction in the left circumflex territory 4 months after transfer of bone-marrow-cells into the left anterior descending coronary artery. This patient underwent PCI of the left circumflex coronary artery and completed the study.

There were no differences between the control and bone-marrow-cell groups with respect to the number of premature ventricular complexes per h and the occurrence of non-sustained or sustained ventricular tachycardias by Holter monitoring at 6 weeks', 3 months', and 6 months' follow-up. 28 (93%) controls and 27 (90%) patients who had bone-marrow-cell transfer agreed to undergo an electrophysiological study at 6 months' follow-up. A non-sustained ventricular tachycardia was inducible in one control patient and in one bone-marrow-cell transfer patient. Ventricular fibrillation was inducible in one control patient. In 29 (97%) controls and 28 (93%) patients who had bone-marrow-cell transfer, coronary angiograms were obtained at 6 months' follow-up. Mean in-stent restenosis in the infarct-related artery, expressed as a percentage of luminal diameter, was 32% (SD 20) in the control group and 33% (23) in the bone-marrow-cell group ($p=0.88$). Four

patients from the control group and seven from the bone-marrow-cell group presented with an in-stent restenosis of at least 50% ($p=0.28$). One patient from the control group developed total in-stent occlusion.

[▲ top](#)

Discussion

Our randomised controlled clinical trial addresses the effect of autologous bone-marrow-cell therapy on left-ventricular functional recovery after acute ST-segment elevation myocardial infarction. We have shown that infusion of autologous bone-marrow-cells into the infarct-related coronary artery during the early postinfarction period (4-8 days after symptom onset) improves recovery of global LVEF after 6 months.

In view of the size of our trial, subgroup analyses must be considered with caution. With this caveat in mind, it is noteworthy that the effects of bone-marrow-cell transfer on global LVEF change were consistent across all investigated subgroups. The effects of cell transfer were over and above benefits associated with established strategies to promote functional recovery after acute myocardial infarction, such as PCI with stent implantation, and postinfarction pharmacotherapy with ACE-inhibitors, angiotensin-receptor blockers and β blockers.^{11,12}

Global LVEF at baseline was 51% (SD 10) in our patient cohort, which is consistent with previous MRI studies in patients after myocardial infarction.^{14,15} In healthy adults, normal LVEF values of 67% (5) have been shown with MRI.¹⁶ Therefore, patients enrolled in our study had substantial functional impairment. Global LVEF increased by only 0.7 percentage points after 6 months' in the control group, emphasising the need for additional therapeutic strategies to enhance functional recovery in patients with acute myocardial infarction. Since 40% of patients had been transferred for rescue PCI from outside hospitals, the average time from symptom onset to PCI was quite long in our trial (median 8.5 h). Previous studies have shown that greater LVEF improvement (up to 4 percentage points) can be achieved when coronary patency is re-established within 4 h of symptom onset.^{17,18} Of note, however, is that in these studies baseline LVEF was measured within 24 h of PCI.^{17,18} By contrast, we assessed baseline LVEF 3.5 days (SD 1.5) after PCI, at a time when left-ventricular function is likely to have partly recovered from postischaemic myocardial dysfunction (ie, stunning).¹⁹ Similar to the results obtained in our control group, two MRI studies that used serial LVEF measurements in patients with reperfused myocardium after acute myocardial infarction have reported no significant improvement in LVEF from a baseline investigation at day 5-7, to follow-up at 3-6 months.^{14,15}

Improvement of global LVEF in the treatment group was due mostly to improved regional systolic wall motion in the infarct border zone. Left-ventricular end-diastolic volumes did not decrease, indicating that transfer of bone-marrow-cells did not improve left-ventricular remodelling at 6 months. Longer follow-up of our patients is required (and will be done) to assess the impact of bone-marrow-cell transfer on long-term left-ventricular structural adaptation after acute myocardial infarction.

Because of ethical considerations, we decided not to do bone-marrow aspiration and a sham left-heart catheterisation in patients randomised to the control group. Importantly, however, all MRI data were analysed by two investigators who were not aware of treatment assignments.

Our study was not designed to assess underlying mechanisms of treatment with bone-marrow-cells that promote functional recovery after acute myocardial infarction. Apparently, transdifferentiation of bone-marrow-derived haemopoietic stem cells to cardiomyocytes cannot account for the beneficial effects.^{20,21} Instead, recent papers have highlighted the potential of bone-marrow cells to promote paracrine effects in ischaemic tissues (eg, secretion of angiogenic factors), and suggest that paracrine signalling, rather than cell incorporation, promotes functional recovery.^{5,22-25}

Our experience suggests that intracoronary bone-marrow-cell transfer is safe; specifically, there was no evidence for an increased rate of in-stent restenosis or proarrhythmic effects. It should be noted that high rates of in-stent restenosis have been reported after intracoronary transfer of granulocyte colony-stimulating-factor mobilised peripheral-blood mononuclear cells.²⁶ Importantly, granulocyte colony-stimulating factor, which may promote in-stent restenosis by enhancing neutrophil recruitment at sites of tissue injury,²⁷ was not used in our study. Intracoronary injection of bone marrow-derived mesenchymal stromal cells has been shown to cause microinfarctions in dogs.²⁸ It should be noted that nucleated bone-marrow cells are significantly smaller than expanded mesenchymal stromal cells *ex vivo*,²⁸ which may explain why we, and others,¹⁰ did not observe infarctions (ie, increases in concentrations of troponin T in serum) after intracoronary transfer of bone-marrow cells.

Our results lend support to the concept that autologous bone-marrow cells can be used to enhance left-ventricular functional recovery in patients after acute myocardial infarction. Larger trials are needed to address the effect of bone-marrow cell transfer on clinical endpoints such as the incidence of heart failure and survival.

Contributors

K C Wollert contributed to study design, enrolment, and clinical follow-up of patients, aspiration and intracoronary transfer of bone marrow, and the writing of the manuscript. G P Meyer contributed to study design, enrolment of patients, MRI data acquisition, and intracoronary BMC transfer. J Lotz contributed to MRI data acquisition. C Breidenbach and S Fichtner analysed MRI data. S Ringes-Lichtenberg contributed to enrolment and clinical follow-up of patients. T Korte did electrophysiological studies. B Hornig did intracoronary transfer of bone-marrow cells. P Lippolt and D Messinger did statistical analyses. L Arseniev did bone-marrow-cell sedimentations. B Hertenstein and A Ganser contributed to study design and did bone-marrow aspirations. H Drexler contributed to study design and the writing of the manuscript.

Conflict of interest statement

L Arseniev is business unit leader of Cytonet Hannover, the company that did the bone-marrow-cell sedimentations during the trial. L Arseniev has not been involved in any way in MRI data collection or data analysis in this trial.

Acknowledgments

Kai C Wollert and Gerd P Meyer contributed equally to this work. We thank Alix Menke and Dieter Fischer for analysing the coronary angiograms; our colleagues and nurses at the Departments of Cardiology and Angiology and Diagnostic Radiology, and at Cytonet Hannover for their support during the trial. The trial was supported by internal funding from the Department of

Cardiology.

References

- 1 Keeley EC, Boura JA, Grines CL. Primary angioplasty versus intravenous thrombolytic therapy for acute myocardial infarction: a quantitative review of 23 randomised trials. *Lancet* 2003; **361**: 13-20. [Text]
- 2 Giugliano RP, Braunwald E. Selecting the best reperfusion strategy in ST-elevation myocardial infarction: it's all a matter of time. *Circulation* 2003; **108**: 2828-30. [PubMed]
- 3 Cohn JN, Ferrari R, Sharpe N. Cardiac remodeling: concepts and clinical implications: a consensus paper from an international forum on cardiac remodeling. *J Am Coll Cardiol* 2000; **35**: 569-82. [PubMed]
- 4 Orlic D, Kajstura J, Chimenti S, et al. Bone marrow cells regenerate infarcted myocardium. *Nature* 2001; **410**: 701-05. [PubMed]
- 5 Kamihata H, Matsubara H, Nishiue T, et al. Implantation of bone marrow mononuclear cells into ischemic myocardium enhances collateral perfusion and regional function via side supply of angioblasts, angiogenic ligands, and cytokines. *Circulation* 2001; **104**: 1046-52. [PubMed]
- 6 Blau HM, Brazelton TR, Weimann JM. The evolving concept of a stem cell: entity or function? *Cell* 2001; **105**: 829-41. [PubMed]
- 7 Korbly M, Estrov Z. Adult stem cells for tissue repair: a new therapeutic concept? *N Engl J Med* 2003; **349**: 570-82. [PubMed]
- 8 Perin EC, Geng YJ, Willerson JT. Adult stem cell therapy in perspective. *Circulation* 2003; **107**: 935-38. [PubMed]
- 9 Strauer BE, Brehm M, Zeus T, et al. Repair of infarcted myocardium by autologous intracoronary mononuclear bone marrow cell transplantation in humans. *Circulation* 2002; **106**: 1913-18. [PubMed]
- 10 Assmus B, Schachinger V, Teupe C, et al. Transplantation of Progenitor Cells and Regeneration Enhancement in Acute Myocardial Infarction (TOPCARE-AMI). *Circulation* 2002; **106**: 3009-17. [PubMed]
- 11 Ryan TJ, Antman EM, Brooks NH, et al. 1999 update: ACC/ AHA guidelines for the management of patients with acute myocardial infarction: executive summary and recommendations: a report of the American College of Cardiology/American Heart Association Task Force on Practice Guidelines (Committee on Management of Acute Myocardial Infarction). *Circulation* 1999; **100**: 1016-30. [PubMed]
- 12 Hunt SA, Baker DW, Chin MH, et al. ACC/AHA guidelines for the evaluation and management of chronic heart failure in the adult: executive summary a report of the American College of Cardiology/American Heart Association Task Force on Practice Guidelines (Committee to revise the 1995 guidelines for the evaluation and management of heart failure): developed in collaboration with the international society for heart and lung transplantation; endorsed by the Heart Failure Society of America. *Circulation* 2001; **104**: 2996-3007. [PubMed]
- 13 Wu E, Judd RM, Vargas JD, Klocke FJ, Bonow RO, Kim RJ.

Visualisation of presence, location, and transmural extent of healed Q-wave and non-Q-wave myocardial infarction. *Lancet* 2001; **357**: 21-28. [Text]

14 Schroeder AP, Houliand K, Pedersen EM, Nielsen TT, Egeblad H. Serial magnetic resonance imaging of global and regional left ventricular remodeling during 1 year after acute myocardial infarction. *Cardiology* 2001; **96**: 106-14. [PubMed]

15 Beek AM, Kuhl HP, Bondarenko O, et al. Delayed contrast-enhanced magnetic resonance imaging for the prediction of regional functional improvement after acute myocardial infarction. *J Am Coll Cardiol* 2003; **42**: 895-901. [PubMed]

16 Lorenz CH, Walker ES, Morgan VL, Klein SS, Graham TP Jr. Normal human right and left ventricular mass, systolic function, and gender differences by cine magnetic resonance imaging. *J Cardiovasc Magn Reson* 1999; **1**: 7-21. [PubMed]

17 Montalescot G, Barragan P, Wittenberg O, et al. Platelet glycoprotein IIb/IIIa inhibition with coronary stenting for acute myocardial infarction. *N Engl J Med* 2001; **344**: 1895-903. [PubMed]

18 Stone GW, Grines CL, Cox DA, et al. Comparison of angioplasty with stenting, with or without abciximab, in acute myocardial infarction. *N Engl J Med* 2002; **346**: 957-66. [PubMed]

19 Braunwald E, Kloner RA. The stunned myocardium: prolonged, postischemic ventricular dysfunction. *Circulation* 1982; **66**: 1146-49. [PubMed]

20 Murry CE, Soonpaa MH, Reinecke H, et al. Haematopoietic stem cells do not transdifferentiate into cardiac myocytes in myocardial infarcts. *Nature* 2004; **428**: 664-68. [PubMed]

21 Balsam LB, Wagers AJ, Christensen JL, Kofidis T, Weissman IL, Robbins RC. Haematopoietic stem cells adopt mature haematopoietic fates in ischaemic myocardium. *Nature* 2004; **428**: 668-73. [PubMed]

22 Tateishi-Yuyama E, Matsubara H, Murohara T, et al. Therapeutic angiogenesis for patients with limb ischaemia by autologous transplantation of bone-marrow cells: a pilot study and a randomised controlled trial. *Lancet* 2002; **360**: 427-35. [Text]

23 Ziegelhoeffer T, Fernandez B, Kostin S, et al. Bone marrow-derived cells do not incorporate into the adult growing vasculature. *Circ Res* 2004; **94**: 230-38. [PubMed]

24 Kinnaird T, Stabile E, Burnett MS, et al. Marrow-derived stromal cells express genes encoding a broad spectrum of arteriogenic cytokines and promote in vitro and in vivo arteriogenesis through paracrine mechanisms. *Circ Res* 2004; **94**: 678-85. [PubMed]

25 Heil M, Ziegelhoeffer T, Mees B, Schaper W. A different outlook on the role of bone marrow stem cells in vascular growth: bone marrow delivers software not hardware. *Circ Res* 2004; **94**: 573-74. [PubMed]

26 Kang HJ, Kim HS, Zhang SY, et al. Effects of intracoronary infusion of peripheral blood stem-cells mobilised with granulocyte-colony stimulating factor on left ventricular systolic function and restenosis after coronary

stenting in myocardial infarction: the MAGIC cell randomised clinical trial. *Lancet* 2004; **363**: 751-56. [[Text](#)]

27 Chakraborty A, Hentzen ER, Seo SM, Smith CW. Granulocyte colony-stimulating factor promotes adhesion of neutrophils. *Am J Physiol Cell Physiol* 2003; **284**: C103-10. [[PubMed](#)]

28 Vulliet PR, Greeley M, Halloran SM, MacDonald KA, Kittleson MD. Intracoronary arterial injection of mesenchymal stromal cells and microinfarction in dogs. *Lancet* 2004; **363**: 783-84. [[Text](#)]

▲ [top](#)



| [Home](#) | [Privacy](#) | [Terms & Conditions](#) Copyright © 2004 [Elsevier Limited](#). All rights reserved.

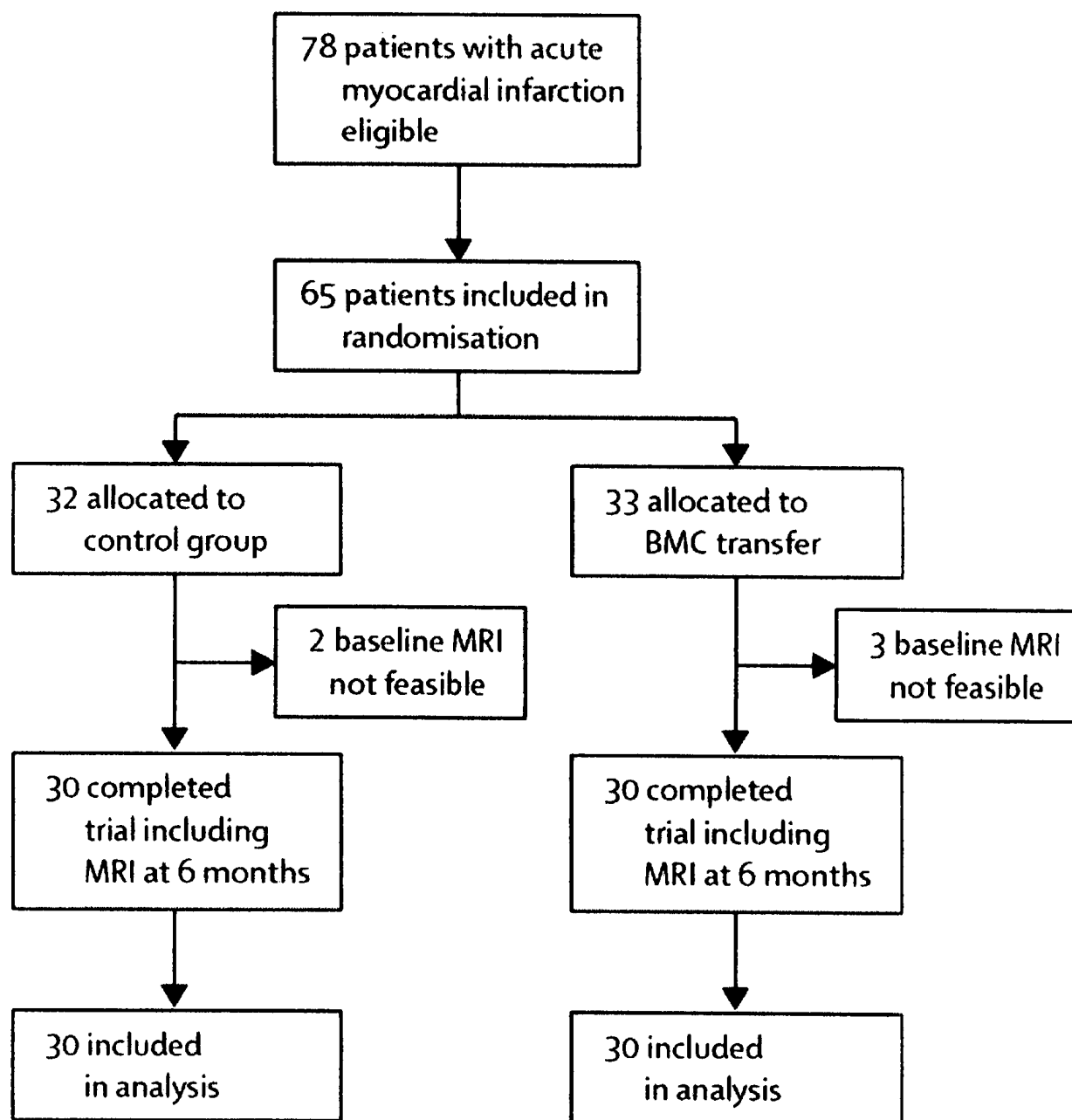


Figure1:Trial profile

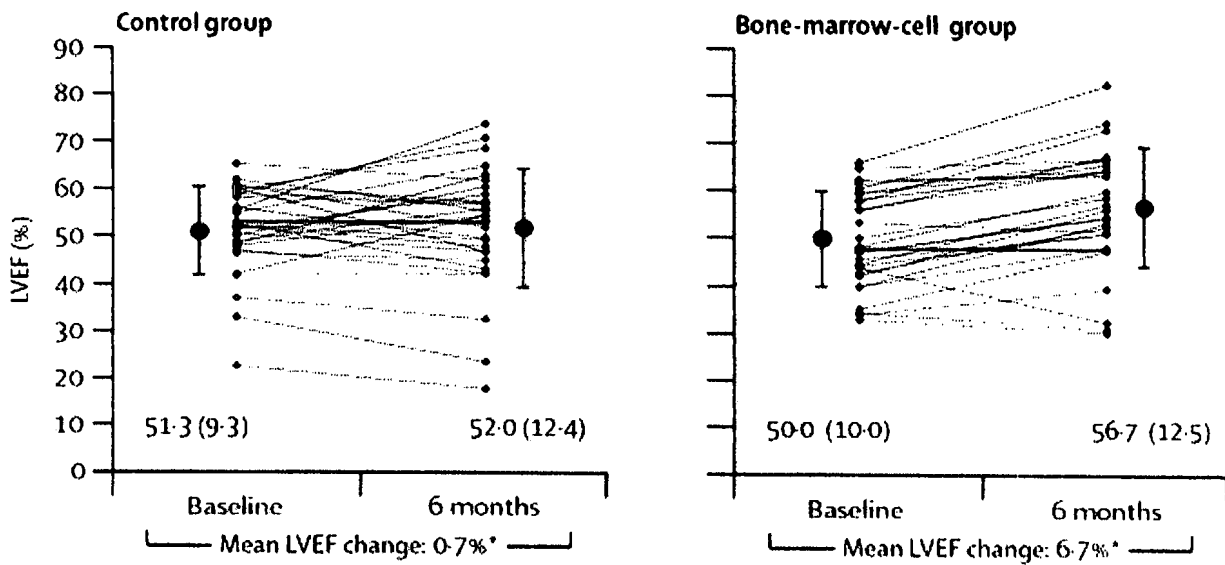


Figure2:Global LVEF at baseline and 6months' follow-up

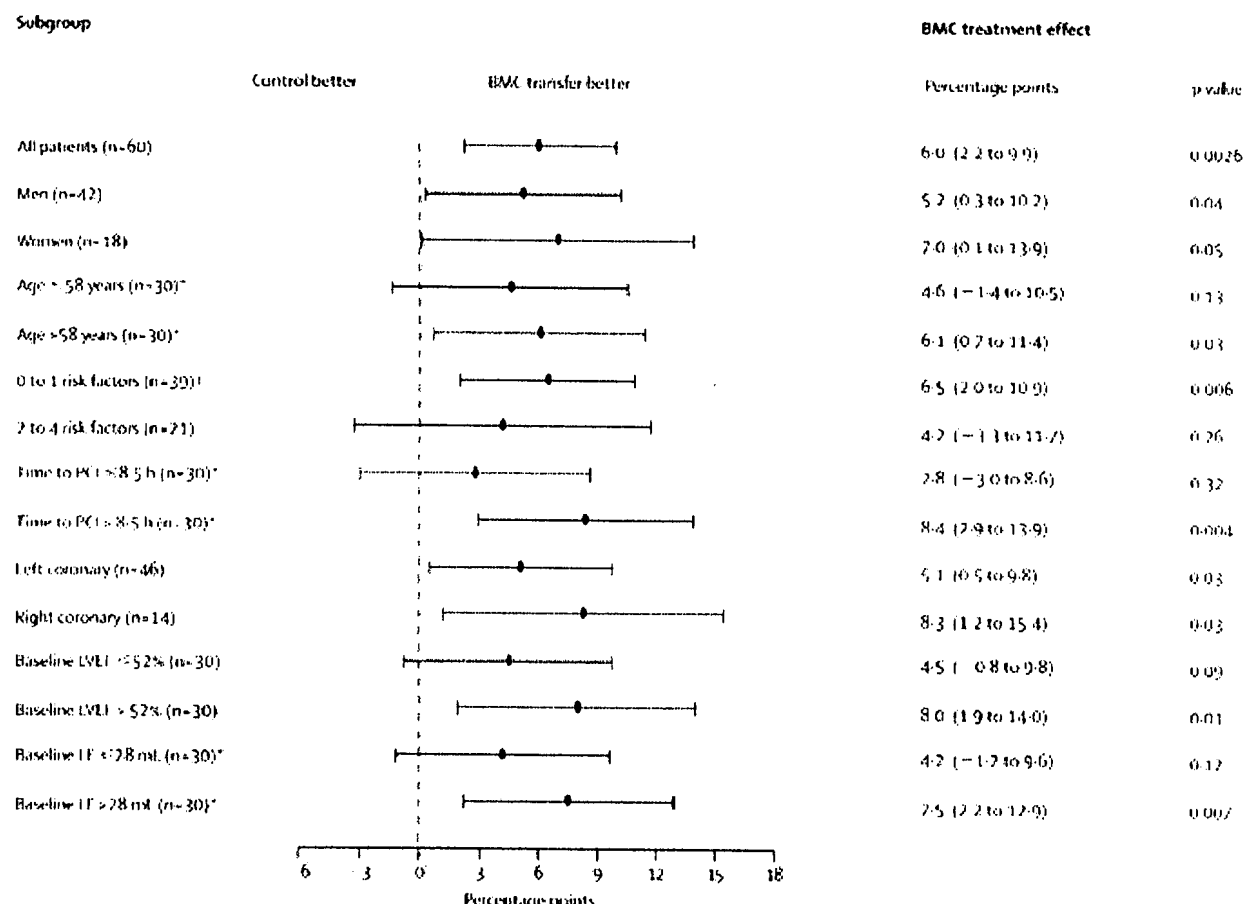


Figure3:Subgroup analyses of global LVEF changes from baseline at 6months' follow-up

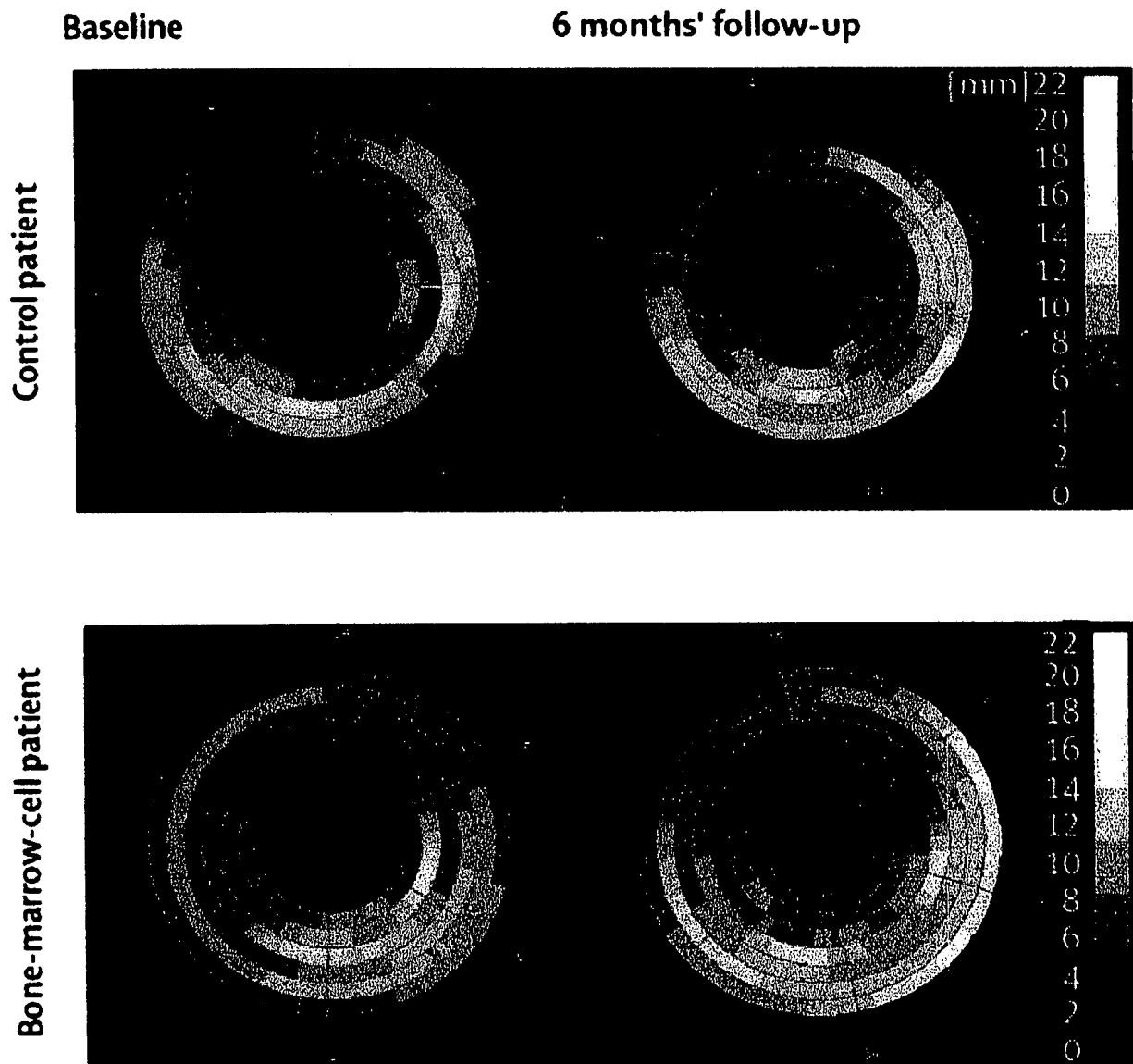


Figure4:Representative colour-coded images showing systolic wall motion at baseline and 6months' follow-up in two patients

EVIDENCE APPENDIX

Item 14

CERTIFICATE OF MAILING

RECEIVED
03 JUN 17 AM 12:31

I hereby certify that the attached DECLARATION OF RICHARD HEUSER, M.D. was delivered to the Assistant Commissioner for Patents by the undersigned from Arrow Intellectual Property Services, 2001, Jefferson Davis Highway, Suite 602, Arlington, Virginia 22202, by hand carrying said DECLARATION to Art Unit 1646, Crystal Plaza 1, Tenth Floor, Attention: Examiner Elizabeth C. Kemmerer this 17th day of June, 2003.

Dated: June 17, 2003

Ann Rutledge
Printed Name: Ann Rutledge

ARROW INTELLECTUAL PROPERTY SERVICE

Docket No. ~~XXXXXX~~: 1000-10-CO1
Serial No. ~~XXXXXX~~: 09/836,750
Filed/Registered ~~XXXXXX~~: 04/17/01
Due Date: - - -

The Patent Office acknowledges, and has stamped hereon, the date of receipt of the items check below:

- ☐ Transmittal Letter
- ☐ Application - Trademark
- ☐ Application - Patent Specification Total Pgs _____
- ☐ Total Claims _____ Ind. Claims _____ Total Pgs _____
- ☐ Fee: \$ _____
- ☐ Abstract Total Pgs _____
- ☐ Drawings: Formal _____ Informal _____ Total Pgs _____
- ☐ Declaration/Oath/Power of Attorney Total Pgs _____
- ☐ Assignment Fee: \$ _____
- ☐ Request for Non-Publication
- ☐ Information Disclosure Statement
- ☐ Form PTO-1449 References _____ Total No _____
- ☐ Request for Extension of Time Fee: \$ _____
- ☐ Amendment/Response
- ☒ ~~XXXXXX~~ Declaration of Richard Heuser
- ☐ Brief/Reply Brief/Notice of Appeal
- ☐ Fee-Base/Maintenance Fee: \$ _____
- ☐ Check No. _____ Fee: \$ _____
- ☐ _____
- ☐ _____

COPY

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

APPLICANT: James P. Elia)	
)	
SERIAL NO.: 09/836,750)	EXAMINER: E.C. Kemmerer, Ph.D.
)	
FILED: April 17, 2001)	
)	GROUP ART UNIT: 1646
FOR: METHOD AND APPARATUS)	
FOR INSTALLATION OF)	
DENTAL IMPLANT)	

DECLARATION OF RICHARD HEUSER, M.D.

I, Richard Heuser, declare as follows:

1. I have offices at 525 North 18th Street, Suite 504, Phoenix, Arizona 85006.
2. My Curriculum Vitae is attached hereto as Exhibit A.
3. I have read and understood the disclosures of the above-referenced patent application at page 20, line 10 through page 21, line 15; and page 44, line 19 through page 46, line 16. A copy of such disclosures is attached hereto as Exhibit B.
4. I note that the disclosures referenced in above Paragraph 3 relate to using a growth factor for promoting the growth of soft tissue and, more specifically, to a method of using a growth factor for growing muscle in a human heart.

5. I am aware of and have considered the definition of *growth factor* in the specification of the above-referenced patent application at page 20, line 10 through page 21, line 15. Such definition is set forth in Exhibit C along with a definition from the medical dictionary, MEDLINE plus: Merriam-Webster Medical Dictionary. A service of the U.S. NATIONAL LIBRARY OF MEDICINE and the NATIONAL INSTITUTES OF HEALTH. I find that the dictionary definition is consistent with that contained at page 20, line 10 through page 21, line 15 of the above-referenced patent application. I believe that both definitions are appropriate for use in the field of tissue growth and would be understood by one skilled in the medical arts. Accordingly, I am adopting and utilizing the definition contained in the patent application throughout this declaration.
6. I have read and understood the claims set forth in Exhibit D and have been informed that such claims will be presented to the Patent and Trademark Office in the near future.
7. The materials included in attached Exhibit E illustrate that placement of a growth factor in a human patient causes muscle growth in a heart. These materials report work performed by reputable, skilled scientists and reputable organizations in the medical arts. Consequently, I believe that these reports would be recognized as clearly valid by one of ordinary skill in the medical arts because they report the results of scientific tests conducted by competent, disinterested third parties with use of proper scientific controls.
8. Based upon above Paragraphs 3-7, it is my opinion that introducing a growth factor into a human patient will predictably cause new muscle growth in the heart of the patient.

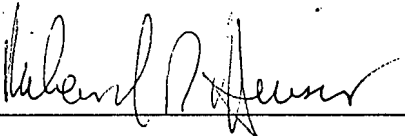
9. Based upon above Paragraphs 3-6, it is my opinion that one skilled in the medical arts, armed with the knowledge in such paragraphs, would be able to practice the method set forth in Exhibit D without need for resorting to undue experimentation.
10. Declarant states that the above opinion was reached independently.

Declarant understands that (1) any willful false statements and the like made herein are punishable by fine or imprisonment, or both (18 U.S.C. 1001) and may jeopardize the validity of the application or any patent issuing thereon, and (2) that all statements made of Declarant's own knowledge are true and that all statements made on information and belief are believed to be true.

Further Declarant sayeth not.

Date: _____

6/5/03



Richard Heuser

EXHIBIT A

CURRICULUM VITAE

Curriculum Vitae
Richard Ross Heuser, M.D., F.A.C.C., F.A.C.P.

ADDRESS:

525 North 18th Street, Suite 504
Phoenix, Arizona 85006
(602) 234-0004
(602) 234-0058 (fax)
phoenixheart@earthlink.net

EDUCATION:

1969 - 1972 University of Wisconsin
 Honors in Chemistry
 Phi Beta Kappa
 Evan Helfaer Scholarship in Chemistry

1972 - 1976 University of Wisconsin School of Medicine
 Graduation with Honors – May 1976
 Alpha Omega Alpha
 Evan Helfaer Scholarship in Medicine

POST GRADUATE TRAINING:

1976 - 1977 Internship in Medicine
 The Johns Hopkins Hospital
 Baltimore, Maryland

1977 - 1979 Residency in Medicine
 The Johns Hopkins Hospital
 Baltimore, Maryland

1979 - 1981 Fellowship in Cardiology
 The Johns Hopkins Hospital
 Baltimore, Maryland

LICENSURE:

State of Arizona, License #19703
State of New Mexico, License #83-220

EMPLOYMENT:

December 2002 - Present Director of Cardiovascular Research
 St. Joseph's Hospital and Medical Center
 Phoenix, Arizona

April 2001 - Present Cardiac Cath Lab Director
 St. Luke's Medical Center, Phoenix, Arizona

June 2000 - Present Medical Director
 Discovery Alliance, Phoenix, Arizona

1998 - June 2000 Director
 Phoenix Research Center, Phoenix, Arizona

April 1997 - Present	Medical Director Phoenix Heart Center, Phoenix, Arizona
December 1999 - Present	Director of Research St. Luke's Medical Center, Phoenix, Arizona
April 1997 - December 1999	Director of Research and Education Phoenix Regional Medical Center, Phoenix, Arizona
April 1990 - April 1997	Director of Research and Education Arizona Heart Institute, Phoenix, Arizona
July 1983 - April 1990	Private Practice New Mexico Heart Clinic, Albuquerque, New Mexico
July 1982 - June 1983	Private Practice Houston Cardiovascular Associates, Houston, Texas
June 1981 - July 1982	Instructor in Medicine, Cardiology The Johns Hopkins Hospital, Baltimore, Maryland

PROFESSIONAL APPOINTMENTS:

1981 - July 1982	Instructor in Medicine - Cardiology Division of Cardiology The Johns Hopkins Hospital, Baltimore, Maryland
July 1982 - June 1983	Instructor in Medicine, Cardiology Baylor College of Medicine, Houston, Texas
July 1983 - February 1990	Director, Interventional Cardiology New Mexico Heart Clinic, Albuquerque, New Mexico
April 1984 - June 1986	Clinical Assistant Professor of Medicine University of New Mexico, Albuquerque, New Mexico Director, Medical Residency Program New Mexico Heart Clinic, Albuquerque, New Mexico
June 1986 - April 1990	Clinical Associate Professor of Medicine University of New Mexico, Albuquerque, New Mexico
May 1996 - April 1997	Director, Interventional Cardiology Arizona Heart Institute Foundation, Phoenix, Arizona
Sept 1995 - December 1999	Medical Director - Cardiac Catheterization Laboratory Phoenix Regional Medical Center, Phoenix, Arizona
December 1990 - Present	Clinical Associate Professor of Medicine University of Louisville, Louisville, Kentucky
April 1990 - April 1997	Director of Research and Education Arizona Heart Institute Foundation, Phoenix, Arizona

April 1997 - December 1999 Director of Research and Education
Phoenix Regional Medical Center, Phoenix, Arizona

BOARD MEMBERSHIPS:

American Board of Internal Medicine
American Board of Cardiovascular Diseases, Diplomat
American Board of Interventional Cardiovascular Diseases, Diplomat

PROFESSIONAL MEMBERSHIPS:

Fellow, American College of Angiology
Fellow, American College of Cardiology
Fellow, American College of Physicians
Fellow, of the American Heart Association
Fellow, American Society of Cardiovascular Interventions
Fellow, International Society of Cardiovascular Interventions
Fellow, Society for Cardiac Angiography and Interventions
Member, American Association for the Advancement of Science
Member, American Heart Association
Member, American Medical Association
Member, Houston Cardiology Society
Member, Houston Society of Internal Medicine
Member, International Andreas Grüntzig Society
Member, International Network of Interventional Cardiology
Member, International Society for Carotid Artery Therapy
Member, International Society for Minimally Invasive Cardiac Surgery
Member, New Mexico Medical Society
Member, Harris County Medical Society
Member, Texas Medical Association
Member, National Register's Who's Who in Executives and Professionals
Member, Who's Who in Medicine and Healthcare 2002-2003

CLINICAL ADVISORY BOARDS:

Advanced Cardiovascular Systems
USCI
Mansfield Scientific Interventional Board
Medtronic Interventional Vascular
Scientific Advisory Board of International Society of Heart Failure

EDITORIAL BOARDS:

Catheterization and Cardiovascular Diagnosis
Journal of Endovascular Surgery
Cardiovascular Research Foundation/Society of Cardiac Angiography and Interventions
Abstract Grader TCT

DATA SAFETY BOARDS:

- ICEM Data Safety Monitoring Board

- Abbott Laboratories Data Safety Monitoring Board for Drug Coated Stent Program, PREFER, A Perspective STUDY to Evaluate the Safety and Efficacy of the ABT-578 coated BiodivYsio® Stent for the Reduction of Resethnosis

CONSULTANT TO:

Editors of the *Annals of Internal Medicine*
 Editors of *Catheterization and Cardiovascular Diagnosis*
 Editors of *Circulation*
 Editors of the *Journal of Invasive Cardiology*
 Editors of the *American Journal of Cardiology*
 Editors of *Web M.D.*
 Annual Scientific Session Program Committee of the American College of Cardiology
 Annual Scientific Session Program Committee of the American College of Cardiology
 Abstract Advisor for Angioplasty; Stents
 Annual International Symposium of Transcatheter Cardiovascular Therapeutics
 Abstract Grader

DEVICE RESEARCH:

Sub-Investigator	ACS Multi-Link Stent Trial Principal Investigator - ACS RX
Principal Investigator	ACT-One Trial Principal Investigator - Angio-Seal Trial
Principal Investigator	Balloon Expandable Intraluminal Stent for Subtotally Occluded Iliac Arteries
Principal Investigator	Bard® Memotherm Carotid Stent Study
Principal Investigator	BARRICADE Trial - The Barrier Approach to Restenosis: Restrict Intima and Curtail Adverse Events (JOMED JOSTENT)
Principal Investigator	BEST Trial
Principal Investigator	BetaCath System Trial
Principal Investigator	Boehringer Ingelheim Pharmaceuticals Protocol Comparing Micardis and COZAAR
Principal Investigator	CABERNET Clinical Trial - Carotid Artery Revascularization using the Boston Scientific EPI FiltreWire EX™ and the EndoTex™ NexStent™
Principal Investigator	CADILLAC Trial
Principal Investigator	CAPRICORN Trial
Principal Investigator	CAPTIVE - Cardioshield Application Protects During Transluminal Intervention of Vein Grafts by Reducing Emboli
Principal Investigator	CARDIOMETRICS
Principal Investigator	Carotid Wallstent Trial
Principal Investigator	CAVEAT II Trial
Principal Investigator	Clinical Investigation of the Magnum Wire vs. Standard Guide Wires during Total Occlusion Angioplasty
Principal Investigator	Cook GR II Trial
Principal Investigator	CORDIS Nitinol Carotid Stent And Delivery System for the Treatment of Obstructive Carotid Artery Disease
Principal Investigator	Cordis Carotid Randomized Sapphire
Principal Investigator	Cordis Bilateral AAA Device & Delivery System
Principal Investigator	(CATS) Safe-Steer™ Wire System Coronary Artery Total Occlusion Study
Principal Investigator	CREDO Trial
Principal Investigator	Novoste CUP Trial
Principal Investigator	CVD Accucath Infusion Catheter
Principal Investigator	Duett Closure Device
Principal Investigator	EndoSonics Cath scanner Oracle - PTCA Catheter

Principal Investigator EPI FilterWire EX™ System During Transluminal Intervention of Saphenous Vein Grafts

Principal Investigator Extra Stent

Principal Investigator GREAT - Guided Radio Frequency Energy Ablation of Total Occlusions Using the Safe Cross™ Radio Frequency Total Occlusion Crossing System

Principal Investigator GRIP - Guided Radio Frequency in Peripheral Total Occlusions using the Safe-Cross™ Radio Frequency (RF) Total Occlusion (TO) Crossing System

Principal Investigator HIPS Trial

Principal Investigator Human Percutaneous Laser Angioplasty of the Coronary Arteries

Principal Investigator Johnson & Johnson Intracoronary Stent Program Supplement #27 "New" Delivery System

Principal Investigator Kensey Nash Hemostatic Puncture Closure Device

Principal Investigator Mansfield-Boston Scientific Strecker Coronary Stent

Principal Investigator Medtronic AVE S7 with Discrete Technology Coronary Stent System

Principal Investigator Medtronic AVE S7 Coronary Stent Registry

Principal Investigator MOBILE Trial - More Patency with Beta for In-Stent Restenosis in the Lower Extremities Trial IDE #G010295; Protocol D00789 Rev B dated 12/01

Principal Investigator NIR Stent Trial

Principal Investigator Neurex/Elan Pharmaceuticals Trial

Principal Investigator PAMI Stent Trial

Principal Investigator Paragon Stent

Principal Investigator Paris Radiation Trial

Principal Investigator PaS Trial

Principal Investigator Percutaneous Coronary Angioscopy in Unstable Angina

Principal Investigator Percutaneous-Recanalization of Stenotic Human Coronary Arteries with Balloon Expandable Intracoronary Stents

Principal Investigator Percutaneous Recanalization of Stenotic Human Saphenous Vein Bypass Graft with Balloon Expandable Intraluminal Stents

Principal Investigator Percutaneous Thermal Balloon Angioplasty

Principal Investigator PMR Trial

Principal Investigator Pravastatin or Atorvastatin Evaluation and Infection Therapy (Prove It)

Principal Investigator Presto Trial

Principal Investigator RAVES Trial

Principal Investigator RESCUE Trial

Principal Investigator SAFER - Saphenous Vein Graft Angioplasty Free of Emboli Randomized Study Using the PercuSurge Guard Wire™ System

Principal Investigator SAVED Trial

Principal Investigator Schering-Plough Phase III Study of SCH 58235 in addition to Pravastatin compared to placebo in subjects with primary hypercholesterolemia

Principal Investigator Long-Term, Open-Label, Safety and Tolerability Study of SCH 58235 in Addition to Pravastatin in Patients with Primary Hypercholesterolemia

Principal Investigator Schneider WINS Trial

Principal Investigator SCORES Trial

Principal Investigator Sepracor Study of Norastemizole in Cardiac Compromised Subjects

Principal Investigator SMART Trial (National PI)

Principal Investigator SMART: Post-Approval Study

Principal Investigator SNAPIST - A Phase 2, Safety Study of Systemic Nanoparticle Paclitaxel (ABI-007) For In-Stent Restenosis; IND #63,082

Principal Investigator SOAR - Renal Stent

Principal Investigator Efficacy and Safety Study of the Oral Direct Thrombin Inhibitor H 376/95 Compared with Dose-Adjusted Warfarin (Coumadin) in the Prevention of Stroke and Systemic Embolic Events in Patients with Atrial Fibrillation (SPORTIF V)

Principal Investigator STARS Trial

Principal Investigator START Trial (National PI)

Principal Investigator STRATUS Trial

Principal Investigator STRESS III Trial

Principal Investigator	SUMO Trial
Principal Investigator	(SWING) Sound Wave Inhibition of Neointimal Growth
Principal Investigator	Talent Endoluminal Graft (High Risk & Low Risk)
Principal Investigator	Talent Endoluminal Spring Stent-Graft System
Principal Investigator	Tenax-XR Coronary Stent System
Principal Investigator	TITAN Trial
Principal Investigator	Trimedyn Excimer Laser Assisted Percutaneous Coronary Angioplasty
Sub-Investigator	Trimdyne Percutaneous Eclipse Holmium Laser Coronary Angioplasty
Principal Investigator	VeGAS 2 Trial
Principal Investigator	Velocity Trial Principal Investigator - Venus Stent
Co-Investigator	WALLSTENT Study
Principal Investigator	WIKTOR Coronary Stent

PHARMACOLOGY RESEARCH:

Principal Investigator	Abbott rUK Trial
Principal Investigator	Ajinimoto Pharmaceuticals Double-Blind Placebo-Controlled Study of AT-1015 in Patients with Intermittent Claudication due to peripheral arterial disease
Sub-Investigator	Amgen, Inc. Anakinra Trial for Rheumatoid Arthritis
Principal Investigator	Astra Zeneca Pharmaceutical Trial to Evaluate the Safety and Efficacy of XXXX and Atorvastatin
Principal Investigator	Astra Zeneca Trial Open Label Dose Comparison Study to Evaluate the Safety and Efficacy of Rosuvastatin versus Atorvastatin, Pravastatin, and Simvastatin in Subjects with Hypercholesterolemia
Principal Investigator	Parke-Davis and Pfizer Randomized Open-Label Study Comparing the Efficacy of Once Daily Atorvastatin to Simvastatin in Hypercholesterolemic Patients
Principal Investigator	Pilot Study to Evaluate Intracoronary Administration of Activase for the Treatment of Intracoronary Thrombus
Principal Investigator	Artistic Trial
Principal Investigator	AstraZeneca Trial of Niaspan versus New Generation Statin for the Treatment of Type IIB and Type IV Hyperlipidemia
Principal Investigator	AstraZeneca Multicenter Trial for drug (XXX) and Atorvastatin for the Treatment of Hypercholesterolemia
Principal Investigator	BRAVO Trial
Principal Investigator	BioVail Angina & Hypertension Trial
Principal Investigator	CAPRICORN Trial
Principal Investigator	Challenge Trial
Sub-Investigator	Comparison of Lopentol and Omnipaque in Adult Angiocardiology
Sub-Investigator	Comparison of Intravenous Adenosine to Intravenous Placebo in Termination of Spontaneous or Induced Paroxysmal Supraventricular Tachycardia
Principal Investigator	Centocor Chimeric 7E3 Fab
Principal Investigator	COR Therapeutics Randomized Placebo-Controlled Dose Ranging Study of drug (XXXX) in Patients with Atherosclerotic Cardiovascular, Peripheral Vascular, and/or Cerebrovascular Disease
Sub-Investigator	Dose Response Study of Bucindolol in Patients with Congestive Heart Failure
Principal Investigator	Effects of Recombinant Human Superoxide Dismutase in Patients with Acute Myocardial Infarction Subject to Coronary Artery Reperfusion
Sub-Investigator	Eli Lilly - Agitation/Alzheimer's Trial
Principal Investigator	EPILOG Trial
Principal Investigator	ERASER Trial
Principal Investigator	GUSTO Trial
Principal Investigator	A multi-center, randomized, double blind, placebo-and-active controlled Parallel Group Dose-ranging Study of the HMG CoA Reductase Inhibitor, BMS-423526, in the treatment of Hyperlipidemia

Principal Investigator Study Lovastatin XL with MEVACOR in patients with hypercholesterolemia

Sub-Investigator Lovastatin Multi-Center Trial

Principal Investigator Extended Trial of Lovastatin XL for the treatment of hypercholesterolemia

Principal Investigator Multicenter Double-Blind Placebo controlled trial of drug (XXXX) in patients with Type 2 Diabetes and Congestive Heart Failure

Principal Investigator Effect of LDL-Cholesterol Lowering Beyond Currently Recommended Minimum Targets on coronary heart disease (CHD) Recurrence in patients with Pre-Existing CHD

Principal Investigator A Double-Blind, Multi-Center, Randomized, Placebo-Controlled, Parallel Group Dosing Study Evaluating the Effects of Nebivolol on Blood Pressure in Patients with Mild to moderate Hypertension, NEB 302

Principal Investigator Parallel Group Extension Study to Determine the Safety and Efficacy of Long-Term Nebivolol Exposure in Patients with Mild to Moderate Hypertension NEB 306,

Sub-Investigator NeoTherapeutics Alzheimer's Disease 2000

Sub-Investigator NeoTherapeutics Alzheimer's Disease 2001

Principal Investigator OCTAVE Trial

Sub-Investigator OCTAVE Trial

Principal Investigator Pfizer Phase II Multicenter, double-blind placebo controlled randomized parallel group dose ranging study of the safety of CP529,414 soft-gel capsules

Principal Investigator PLAC Trial

Principal Investigator Protocol 073 Trial

Principal Investigator Knoll Pharmaceutical Double-Blind Randomized Clinical Trial of Slow Release Propafenone (Rythmol-SR®) in the Prevention of Symptomatic Recurrences of Atrial Fibrillation

Principal Investigator PREVAIL - A Phase 2 Multicenter, Double-Blind Placebo-Controlled, Dose-Ranging Study to Evaluate the Safety and Efficacy of BO-653 in Prevention of Post-Angioplasty Restenosis in Stented Lesions

Principal Investigator PROVE-IT TIMI 22 - Pravastatin or Atorvastatin Evaluation and Infection Therapy

Principal Investigator PURSUIT Trial

Principal Investigator QUIET Trial

Principal Investigator RAFT Trial

Principal Investigator REPLACE Randomized Evaluation in PCI Linking Angiomax to reduce Clinical Events

Sub-Investigator Safety and Efficacy Study of Burroughs - Wellcome Tissue Plasminogen Activator in Patients with Acute Myocardial Infarction

Principal Investigator A 6-week, open-label, dose-comparison study to evaluate the safety and Efficacy of Rosuvastatin versus Atorvastatin, Cerivastatin, pravastatin, and Simvastatin in subjects with hypercholesterolemia

Principal Investigator A 48-week, open-label, non-comparative, Multicentre, Phase IIIb study to evaluate the efficacy and safety of the Lipid-Regulating agent Rosuvastatin in the treatment of subjects with Fredrickson Type IIa and Type IIb Dyslipidemia, including Heterozygous Familial Hypercholesterolemia

Principal Investigator SAGE Trial

Sub Investigator Long Term Open Label Safety and Tolerability Study of SCH58235 in addition to Pravastatin in Patient With Primary Hypercholesterolemia

Principal Investigator Phase III Double-Blind Efficacy and Safety Study SCH58235 (10 mg) in Addition to Pravastatin Compared to Placebo in Subjects with Primary Hypercholesterolemia

Principal Investigator Phase III Open Label Efficacy and Safety Study SCH58235 (10 mg) in Addition to Pravastatin Compared to Placebo in Subjects with Primary Hypercholesterolemia

Principal Investigator Sepracor Protocol Study of Norastemizole in Cardiac Compromised Subjects

Principal Investigator SPORTIF V - Atrial Fibrillation Trial

Principal Investigator SWORD Trial

Principal Investigator Titration-to-Response Trial Comparing Micardis and COZAAR® in Patients with Mild-to-moderate Hypertension

Principal Investigator TNT Trial
 Principal Investigator TREND Trial
 Sub-Investigator VALDECOXIB Trial
 Principal Investigator An Open-Label, Multinational, Multicentre, Extension Trial to Assess the
 Long-Term Safety and Efficacy of ZD4522 in Subjects in the ZD4522 Clinical Trial Program

BASIC RESEARCH:

- 1990 - 1993 Systematic assessment of Medtronic balloons and guiding catheters in porcine and canine models. Sponsored by Medtronic, Inc.
- 1990 - 1993 Determination of radiopacity and torquability of Medtronic vascular catheters in porcine models. Sponsored by Medtronic, Inc.
- 1992 - 1996 Evaluation of Strecker stent in porcine and canine models.
Sponsored by Boston Scientific
- Evaluation of Wiktor stent and stent in porcine and canine models.
Sponsored by Medtronic, Inc.
- Evaluation of NIR stent in porcine models.
Sponsored by Cordis Corp.
- 1990 - 1994 Evaluation of Japan Crescent radiofrequency balloon in porcine model with emphasis on histopathology of heat-produced lesions. Abstract submitted at 1993 AHA Conference.
- 1993 Evaluation of radiofrequency wire for total coronary occlusions in porcine models: Determining energy limitations. Equipment subsequently licensed to Radius Medical.
- 1994 - 1997 Training courses for professionals (physicians, engineers, technicians) in techniques and strategies for placement of coronary stents. Five courses sponsored by Johnson & Johnson, Medtronic, Inc. and Cook, Inc.
- 1997 Efficacy of the Endotex Abdominal Aortic Aneurysm exclusion device in a porcine model gauging ability to exclude renal arteries, ease of placement and radiopacity. Sponsored by Endotex
- 1998 Use of percutaneous myocardial revascularization in a porcine model.
Sponsored by Cardiogenesis Corporation at Stanford University.
- 1998 - 1999 Utility of radiofrequency (RF) percutaneous myocardial revascularization in acute and chronic porcine model: Histopathology and angiogenesis related to use of RF alone and in combination with growth factor (VEGF). Results presented at Angiogenesis 1999, Washington, DC.
- 1999 Development and testing of embolic probe device in porcine model (patent pending). Performed at PRMC and separately at Columbia Presbyterian in New York.
- 1999 Evaluation of the Medtronic carotid and SVG stent in porcine carotid and saphenous vein graft lesions assessing ease of use and 30-day outcome.
Sponsored by Medtronic, Inc.
- 1999 Development and testing of Protector vascular embolic protection device in

porcine model at Mayo Clinic (device patent pending).

- 1999 Evaluation of ability of intramuscular growth factor to stimulate angiogenesis in rabbit hindlimb model at 30 and 60 days post-procedure. Sponsored by Sulzer-Medical.
- 1999 Use of *Vesseeal* device to close porcine peripheral artery tears (patent #6,159,197) Sponsored by Phoenix Heart Center.

PUBLICATIONS:

- Bayless TM, **Heuser RR**: Fulminant Colitis. Johns Hopkins Medical Journal 1979 May;144(5): 168-172.
- Heuser RR**, Achuff SC, Brinker JA: Inadvertent division of an anomalous left anterior descending coronary artery during complete repair of tetralogy of fallot. American Heart Journal 1982 Mar;103(3):430-432.
- Fuchs RM, **Heuser RR**, Yin FC, Brinker JA: Limitations of pulmonary wedge V-waves in diagnosing mitral regurgitation. The American Journal of Cardiology 1982 Mar;49(4): 849-854.
- Fuchs RM, Brin KP, Brinker JA, Guzman PA, **Heuser RR**, Yin FC: Augmentation of regional coronary blood flow by intraaortic balloon counterpulsation in patients with unstable angina. Circulation 1983 Jul;68(1):117-123.
- Alexander EL, Weiss JL, Firestein GS, **Heuser RR**, Leitzl G, Wagner Jr HN, Brinker JA, Cluff AA, Becker LC: Reversible cold-induced abnormalities in myocardial perfusion and function in systemic sclerosis. Annals of Internal Medical 1986 Nov;105(5):661-668.
- Heuser RR**: Cardiogenic shock treated by PTCA. Cardiology 1987;4(6):64-66.
- Heuser RR**, Maddoux GL, Goss JE, Ramo BW, Raff GL, Shadoff N: Coronary angioplasty for acute mitral regurgitation due to myocardial infarction: A nonsurgical treatment preserving mitral valve integrity. Annals of Internal Medical 1987 Dec;107(6):852-855.
- Maddoux GL, Goss JE, Ramo BW, Raff GL, **Heuser RR**, Shadoff N, Wilson JN, Deane WM, Hoyt TW, Fowler BN, Gerety RL, Hoffman AR: Left main coronary artery embolism: A case report. Catheterization and Cardiovascular Diagnosis 1987 Nov-Dec;13(6):394-397.
- Maddoux GL, Ramo BW, Goss JE, Raff GL, **Heuser RR**, Shadoff N, Leatherman GF, Blake K, Wilson JN, Deane WM, Hoyt TW, Fowler BN, Gerety RL, Sansonetti E: Angina and vasospasm at rest in a patient with an anomalous left coronary system. Catheterization and Cardiovascular Diagnosis 1989 Feb;16(2):95-98.
- Goss JE, Ramo BW, Raff GL, Maddoux GL, **Heuser RR**, Shadoff N, Leatherman GF, Blake K: Power injection of contrast media during percutaneous transluminal coronary artery angioplasty. Catheterization and Cardiovascular Diagnosis 1989 Mar;16(3):195-198.
- Heuser RR**, Mehta SS: Holmium laser angioplasty after failed coronary balloon dilation: Use of a new solid-state, infrared laser system. Catheterization and Cardiovascular Diagnosis 1991 Jul;23(3):187-189.

- Heuser RR, Mehta SS, Strumpf RK, Ponder R:** Intracoronary stent implantation via the brachial approach: A technique to reduce vascular bleeding complications. *Catheterization and Cardiovascular Diagnosis* 1992 Apr;25(4):300-303.
- Strumpf RK, Mehta SS, Ponder R, **Heuser RR:** Palmaz-Schatz stent implantation in stenosed saphenous vein grafts: Clinical and angiographic follow-up. *American Heart Journal* 1992 May;123(5):1329-1336.
- Segal J, Kern MJ, Scott NA, King SB, Doucette JW, **Heuser RR**, Ofili E, Siegel R: Alterations of phasic coronary artery flow velocity in humans during percutaneous coronary angioplasty. *The Journal of the American College of Cardiology* 1992 Aug;20(2):276-286.
- Heuser RR:** The use of the Holmium: YAG laser in coronary disease: The utility of a unique lensed fiber catheter. *The Journal of Interventional Cardiology* 1992 Dec;5(4):293-300.
- Heuser RR, Eagan JT, Strumpf RK:** Angioscopy in coronary interventions. *Cardiology Intervention* 1992;2(4):23-28.
- Santiago O, Diethrich EB, **Heuser RR**, Gustafson G: What is the next step for the application of the Palmaz stent – the abdominal aorta? *Angiology* 1992;42:267-268.
- Heuser RR, Mehta SS, Strumpf RK:** The ACS RX™ flow support catheter as a temporary stent for dissection or occlusion during balloon angioplasty: initial experience. *Catheterization and Cardiovascular Diagnosis* 1992 Sept;27(1):66-74.
- Diethrich EB, Santiago O, Gustafson G, **Heuser RR:** Preliminary observations on the use of the Palmaz stent in the distal portion of the abdominal aorta. *American Heart Journal* 1993 Feb;125(2):490-501.
- Heuser RR:** Lasers in coronary disease (commentary). *Catheterization and Cardiovascular Interventions* 1993;28:17.
- Eagan JT, Strumpf RK, **Heuser RR:** New treatment approach for chronic total occlusions of saphenous vein grafts: Thrombolysis and intravascular stents. *Catheterization and Cardiovascular Diagnosis* 1993 May;29(1):62-69.
- Heuser RR, Strumpf RK, Diethrich EB, Eagan JT, Hardigan KR:** Intraluminal diagnostics, the "guide" to the future. *Angiology* 1993;44:6.
- Heuser RR, Strumpf RK, Eagan JT, Hardigan KR:** Experience with four types of coronary stents. *Angiology* 1993;44:6-7.
- Heuser RR:** The use of a new wire in a 6-year-old coronary artery occlusion: The Jagwire™ recanalization guidewire. *Catheterization and Cardiovascular Diagnosis* 1993 Jun;29(2):173-176.
- Heuser RR, Strumpf RK, Hardigan K:** Use of the Doppler guidewire for intraluminal diagnosis to facilitate coronary intervention. *American Heart Journal* 1993 July; 126(1):213-218.
- Strumpf RK, **Heuser RR, Eagan Jr JT:** Angioscopy: A valuable tool in the deployment and evaluation of intracoronary stents. *American Heart Journal* 1993 Nov;126(5):1204-1210.
- Heuser RR:** Current status of coronary stents: Promises and disappointments. *Critical Issues* 1993;2:1-13.

- White CJ, Ramee SR, Collins TJ, Escobar AE, Karsan A, Shaw D, Jain SP, Bass TA, **Heuser RR**, Teirstein PS, Bonan R, Walter PD, Smalling RW: Coronary thrombi increase PTCA risk: Angioscopy as a clinical tool. *Circulation* 1996 Jan;93(2):253-258.
- Fischman DL, Leon MB, Baim D, Schatz RA, Penn I, Detre K, Savage MP, Veltri L, Ricci D, Nobuyoshi M, Cleman M, **Heuser RR**, Almond D, Teirstein P, Fish D, Colombo A, Brinker J, Moses J, Hirshfeld J, Bailey S, Ellis S, Rake R, Goldberg S: For the STRESS Trial Investigators: A randomized comparison of coronary stent placement and balloon angioplasty in the treatment of coronary artery disease. *The New England Journal of Medicine* 1994; 331:396-501.
- Aker UT, Kensey KR, **Heuser RR**, Sandza JG, Kussmaul III WG: Immediate arterial homostasis after cardiac catheterization: Initial experience with a new puncture closure device. *Catheterization and Cardiovascular Diagnosis* 1994 Mar;31(3):228-232.
- Flaherty JT, Pitt B, Gruber JW, Topol E, **Heuser RR**, Rothbaum DA, Burwell LR, George BS, Kereiakes DJ, Deitchman D, Gustafson N, Brinker JA, Becker LC, Mancini CBJ, Weisfeldt ML, Werns SW: Recombinant human superoxide dismutase (h-SOD) fails to improve recovery of ventricular function in patients undergoing coronary angioplasty for acute myocardial infarction. *Circulation* 1994 May;89(5):1982-1991.
- Fenton SH, Fischman DL, Savage MP, Schatz RA, Leon MB, Baim DS, King SB, **Heuser RR**, Curry C, Rake RC, Goldberg S: Long-term angiographic and clinical outcome after implantation of balloon-expandable stents in aortocoronary saphenous vein grafts. *The Journal of the American College of Cardiology* 1994; Dec 15;74(12):1187-1191.
- Heuser RR**: Breaking the barrier: Stenting in acute myocardial infarction (commentary). *Catheterization and Cardiovascular and Diagnosis* 1994;33:46.
- Heuser RR**: A wire in every lesion (commentary). *Catheterization and Cardiovascular Interventions* 1994;33:197.
- Heuser RR**: All the glitters in not gold (commentary). *Catheterization and Cardiovascular Interventions* 1994 Dec;33(4):330.
- Savage MP, Fischman DL, Schatz RA, Teirstein PS, Leon MB, Baim DS, Ellis SG, Topol EJ, Hirshfeld JW, Cleman MW, Buchbinder M, Bailey S, **Heuser RR**, Walker CM, Curry RC Jr, Gebhardt S, Rake R, Goldberg S: For the Palmaz-Schatz Study Group: Long-term angiographic and clinical outcome after implantation of a balloon-expandable stent in native coronary circulation. *The Journal of the American College of Cardiology* 1994;24:1207-1212.
- Heuser RR**, Diethrich EB, Papazoglou C, Reynolds GT: Endoluminal grafting for percutaneous aneurysm exclusion in an aortocoronary saphenous vein graft: the first clinical experience. *Journal of Endovascular Surgery* 1995 Feb;2(1):81-88.
- Diethrich EB, **Heuser RR**, Tarlian H, Eckert J, Cardenas JR,: Endovascular techniques in adult aortic coarctation: the use of stents for native and recurrent coarctation repair. *Journal of Endovascular Surgery* 1995 May;(2)2:183-188.
- Aguirre FV, Topol EJ, Ferguson JJ, Anderson K, Blankenship JC, **Heuser RR**, Sigmon K, Taylor M, Gottlieb R, Hanovich G, Rosenberg M, Donohue TJ, Weisman H, Califf RM for the EPIC Investigators: Bleeding complications with the chimeric antibody to platelet glycoprotein IIb/IIIa integrin in patients undergoing percutaneous coronary intervention. *Circulation* 1995; June 15;91(12):2882-2890.

- Heuser RR:** Treatment alternatives for chronically occluded saphenous vein grafts. The Journal of Invasive Cardiology 1995 May;7(4):94-96.
- Kussmaul III WG, Buchbinder M, Whitlow PL, Aker UT, **Heuser RR**, King SB, Kent KM, Leon MB, Kolansky DM, Sandza JG: Rapid arterial homeostasis and decreased access site complications after cardiac catheterization and angioplasty: Results of a randomized trial of a novel hemostatic device. The Journal of the American College of Cardiology 1995 June;25(7):1685-1692.
- Wong SC, Baim DS, Schatz RA, Teirstein PS, King SB 3rd, Curry RC Jr, **Heuser RR**, Ellis SG, Cleman MW, Overlie P, et al. Immediate results and late outcomes after stent implantation in saphenous vs lesions: the multicenter US Palmaz-Schatz stent experience. The Palmaz-Schatz Study Group. Journal of the American College of Cardiology 1995 Sept;26(3):704-712.
- Cardenas JR, Strumpf RK, **Heuser RR:** Rotational artherectomy in restenotic lesions at the distal saphenous vein graft anastomosis. Catheterization and Cardiovascular Interventions 1995 Sept;36(1):53-57.
- Cohen DJ, Krumholz HM, Williams C, Baim DS, Brinker J, Cabin HS, **Heuser RR**, Leon MB, Moses J, Savage MP, Cleman M: In-hospital and one-year economic outcomes after coronary stenting or balloon angioplasty. Results from a randomized clinical trial. The Stent Restenosis Study Investigators. Circulation 1995 Nov.;1;92(9):2480-2487.
- Heuser RR:** A new breed of drug pusher? (commentary). Catheterization and Cardiovascular Interventions 1995;34:271.
- Heuser RR:** Recanalization of occluded SVGs: Is there light at the end of the graft? (commentary). Catheterization and Cardiovascular Diagnosis 1995 Dec;36(4):333-334.
- Heuser RR:** Percutaneous revascularization using endoluminal prostheses: Coming of age (commentary). Catheterization and Cardiovascular Diagnosis 1996 Feb;37(2):213-214.
- Kussmaul WG, Buchbinder M, Whitlow PL, Aker UT, **Heuser RR**, King SB, Kent KM, Leon MB, Kolansky DM, Sandza JG: Femoral artery hemostasis using an implantable device (AngioSeal™) after coronary angioplasty. Catheterization and Cardiovascular Diagnosis 1996 Apr;37(4):362-365.
- Heuser RR:** Stents in jackets: The latest in endovascular haute couture. Catheterization and Cardiovascular Diagnosis 1996 Jun;38(2):179.
- Heuser RR:** "Homemade" stents: Penny-wise or pound-foolish? (commentary) Catheterization and Cardiovascular Diagnosis 1996 Oct;39(2):197.
- Schultz RD, **Heuser RR**, Hatler C, Frey D: Use of c7E3 Fab in conjunction with primary coronary stenting for acute myocardial infarctions complicated by cardiogenic shock. Catheterization and Cardiovascular Diagnosis 1996 Oct;39(2):143-148.
- Heuser RR:** The Multi-Link stent: Another good idea. (commentary) Catheterization and Cardiovascular Diagnosis 1996 Dec;39(4):420.
- Kussmaul WG, Sandza JG, Kolansky DM, Leon MB, Kent KM, King SB, **Heuser RR**, Aker UT, Whitlow PL, Buchbinder M: Femoral artery hemostasis using an implantable device (Angio-Seal™) after coronary angioplasty. Catheterization and Cardiovascular Interventions 1996;37:362-365.

- Heuser RR, Hartz RS:** Embolization of IMA side branch for post-CABG ischemia. *Ann Thorac Surgery* 1997 Jun;63(6):1765-1766.
- Heuser RR:** Unlimiting our resources. (commentary) *Catheterization and Cardiovascular Diagnosis* 1997 Aug;41(4):415.
- Krumholz HM, Cohen DJ, Williams C, Baim DS, Brinker J, Cabin HS, **Heuser RR**, Hirschfeld J, Leon MB, Moses J, Savage MP, Cleman M: Health after coronary stenting or balloon angioplasty: Results from the Stent Restenosis Study (STRESS). *The American Heart Journal* 1997 Sept;134(3):337-44.
- Heuser RR:** A Freudian stent? (commentary) *Catheterization and Cardiovascular Interventions* 1997;41:4.
- Savage MP, Douglas JS, Fischman DL, Pepine CJ, King III SB, Werner JA, Bailey SR, Overlie PA, Fenton SH, Brinker JA, Leon MB, Goldberg S, **Heuser RR:** Stent placement compared with balloon angioplasty for obstructed coronary bypass grafts. *The New England Journal of Medicine* 1997;337:740-747.
- Heuser RR, Lopez AN:** Stenting, angioplasty and adjunctive treatments in acute myocardial infarction. *Cardiology Rounds* 1998;15:9:8-14.
- Savage MP, Douglas Jr JS, Fischman DL, Pepine CJ, King SB, Werner JA, Bailey SR, Overlie PA, Fenton SH, Brinker JA, Leon MB, **Heuser RR**, Smalling R, Safian RD, Cleman M, Buchbinder M, Rake RC, Gebhardt S, Goldberg S for the Stent Restenosis Study (STRESS) Investigators: Efficacy of coronary stenting versus balloon angioplasty in small coronary arteries. *The Journal of the American College of Cardiology* 1998;31:307-311.
- Heuser RR:** Outpatient coronary angiography: Indications, safety, and complication rates. *Herz* 1998 Feb;23(1)21-26.
- Heuser RR:** Garbage in, garbage out. (commentary) *Catheterization and Cardiovascular Interventions* 1998 Apr;43(4):402.
- Heuser RR:** Success with carotid stenting: A stroke of good luck or the wave of the future? (commentary) *Catheterization and Cardiovascular Interventions* 1998 May;44(1)7-8.
- Heuser RR:** Stentus Noninterruptus. (commentary) *Catheterization and Cardiovascular Interventions* 1998;43:253.
- Woodfield SL, Lopez AN, **Heuser RR:** Fracture of Coronary Guidewire During Rotational Arterectomy with Coronary Perforation and Tamponade. *Catheterization and Cardiovascular Diagnosis* 1998 Jun;44(2)220-223.
- Heuser RR, Lopez AN:** Stenting, Angioplasty, and Adjunctive Treatments in Acute Myocardial Infarction. *Cardiology Review* 1998;15:8-14.
- Heuser RR:** Endovascular Haute Couture Revisited: Ready-to-Wear Comes of Age. (commentary) *Catheterization and Cardiovascular Interventions* 1998 Nov;45(3):314.
- Heuser RR:** The Cure for a Rainy Day. (commentary) *Catheterization and Cardiovascular Diagnosis* Dec 1998;45(4):421.
- Heuser RR, Lopez A:** Abdominal Aorta Aneurysm and ELG: A Review of a Treatment in Its Infancy. *Journal of Interventional Cardiology* 1998 December (II);6:591-602.

- Heuser RR**, Woodfield SL, Lopez AN: Obliteration of a Coronary Artery Aneurysm with a PTFE-Covered Stent: Endoluminal Graft for Coronary Disease Revisited. *Catheterization and Cardiovascular Interventions* 1999 Jan;46(1):113-116.
- Heuser RR**: The Right Place, the Right Time. *Catheterization and Cardiovascular Interventions* 1999 Apr;46(4):450-451.
- Heuser RR**: The growing role of stents in coronary artery disease. *Patient Care* May 1999; 43-58.
- Teirstein PS, Mann III JT, Cundey PE, Schechter E, Jacobs WC, Grines CL, Stagaman DJ, Lansky AJ, Hultquist MA, Kusnick BA, **Heuser RR**, Kleinert HD, Popma J: Low versus high-dose recombinant Urokinase for the treatment of chronic saphenous vein graft occlusion. *The American Journal of Cardiology* 1999 June 15;83(12) 1623-1628.
- Heuser RR**: Warming up to the reptilian heart (commentary). *Catheterization and Cardiovascular Interventions* 1999 Jul;47(3):360.
- Heuser RR**: Of Baling Wire and Bubble Gum. *Catheterization and Cardiovascular Interventions* 1999 August;47(4):495.
- Heuser RR**: Innovative Interventions. *Catheterization and Cardiovascular Interventions* 1999 Sept;48(1):113.
- Cordero-Yordan H, Lopez AN., **Heuser RR**: Saphenous Vein Graft Intervention, *The Journal of Interventional Cardiology* 1999;12(6):495-497.
- Cordero-Yordan H., Lopez AN, **Heuser RR**: Carotid Artery Percutaneous Transluminal Angioplasty and Stenting; *The Journal of Interventional Cardiology* 1999;12:499-504.
- Heuser RR**, Cordero-Yordan H: Carotid Artery Percutaneous Transluminal Angioplasty and Stenting: Indications, Technical Approach, and Complications. *The Journal of Interventional Cardiology*. 1999;12(6): 499-504.
- Heuser RR**: At Arms' Length: Should We Teach Radial and Brachial Access Techniques? *Catheterization and Cardiovascular Interventions* 2000 Jan;49(1):38.
- Heuser RR**: Coronary Lesions: The Good, the Bad, and the Ugly, *Catheterization and Cardiovascular Interventions* 2000 Jan;49(1):112.
- Oesterle SN, Sanborn TA, Ali N, Resar J, Ramee SR, **Heuser RR**, Dean L, Knopf W, Schofield P, Schaer GL, Reeder G, Masden R, Yeung AC, Burkoff D: Percutaneous transmyocardial laser revascularization for severe angina: the PACIFIC randomized trial. *The Lancet* (356) November 28, 2000;1705-1710.
- Heuser RR**, Houser F., Culler SD., Becker ER, Battaglia SL, Tarkington L, Simon AW: A Retrospective Study of 6,671 Patients comparing Coronary Stenting and Balloon Angioplasty; *The Journal of Invasive Cardiology* 2000;July 2000.
- Heuser RR**: Delivery Systems for Endoluminal Grafting of Abdominal Aortic Aneurysms: A Square Peg for a Round Hole? (commentary). *Catheterization and Cardiovascular Interventions* 2000 Jul;50(3):361.
- Heuser RR**, Cordero H, Gupta N, Underwood PL, Gogte ST: Intracoronary Autologous Blood to Seal a Coronary Perforation; *Herz* 2001; March 26;(2):157-160.

Heuser RR, Lopez A, Kuntz R, Reduto L, Badger R, Coleman P, Whitlow P, Iannone LA, Safian R, Yeung A, Moses J: SMART: The MicroStent's Ability to Limit Restenosis Trial. Catheterization and Cardiovascular Interventions 52:269-277 2001

Cordero H, Warburton KD, Underwood PL, **Heuser RR**: Initial Experience and Safety in the Treatment of Chronic Total Occlusions with Fiberoptic Guidance Technology: Optical Coherent Reflectometry. Catheterization and Cardiovascular Interventions Oct 2001; 54(2):180-187.

Heuser RR: If It Can Keep Eggs Off the Frying Pan... (commentary) Catheterization and Cardiovascular Interventions May, 2001; 53:5.

Heuser RR, Cordero H, Schroeder W, Weirick BA, Hatler CW, Underwood PL: Fiberoptic Guidance System for Treatment of Chronic Total Occlusions: Single Center Experience. Journal of the American College of Cardiology; February 2001 (37) Supplement A: 1158-38.

Heuser RR, Morales PA: Across the Great Divide (commentary) Catheterization and Cardiovascular Interventions. October 2001; 54(2):264.

Ellis SG, Mooney M, Talley DJ, Silber S, Teirstein PS, Rodriguez AR, Sanborn TA, Feldman T, Leon M, Collins TJ, Wilentz JR, Saucedo JF, Leimgruber PP, Pichard A, Cohen DJ, Ramee SR, **Heuser RR**, Smalling RW, Kuntz RE, Gershony G: Assessment of the Safety and Efficacy of the Duett™ Vascular Hemostasis Device - Final Results of the Seal Trial. American Heart Journal April 2002; 143(4):612-9.

Morales PA, **Heuser RR**: Chronic Total Occlusions: Experience with Fiber-Optic Guidance Technology - Optical Coherence Reflectometry. Journal of Interventional Cardiology; December 2001; 14:6:611-616.

Morales PA, **Heuser RR**: Peripheral Vascular Disease: Perspectives on Aortoiliac, Renal, and Femoral Treatments Using Catheter-Based Techniques. Journal of Interventional Cardiology; December 2001; 14:6:629-637.

Heuser RR, Laskey W, Lansky A, Popma J, Bonan R: Beta Radiation in Lesions > 15 mm: A START Subgroup. Journal of the American College of Cardiology. (In Press)

Philip A. Morales, M.D., **RR Heuser**, Embolic Protection Devices. Journal of Interventional Cardiology; December 2002; 15:485-490.

Philip A. Morales, M.D., and **RR Heuser**, Guiding Catheter Aspiration to Prevent Embolic Events During Saphenous Vein Graft Intervention. Journal of Interventional Cardiology; December 2002; 15:491-498.

Jeffrey J. Popma, Mohan Suntharalingam, Alexandra J. Lansky, **Richard R. Heuser**, Burton Speiser, Paul S. Teirstein, Vincent Massullo, Theodore Bass, Randall Henderson, Sigmund Silber, Peter von Rottkay, Raoul Bonan, Kalon K.L. Ho, Alison Osattin, MPH; Richard E. Kuntz, for the Stents And Radiation Therapy (START) Investigators, Randomized Trial of 90 SR/90 Y B-Radiation Versus Placebo Control for Treatment of In-Stent Restenosis. Circulation; August 2002; 106:1090-1096.

EDITED TEXTBOOKS:

Heuser RR: *Peripheral Vascular Stenting for the Cardiologist*, Martin Dunitz, Ltd Publishing, 1999

Heuser RR, Henry M: *Peripheral Vascular Interventions*, Martin Dunitz, Ltd Publishing; March 2002

TEXTBOOK CHAPTERS:

- Heuser RR:** Coronary angioplasty for acute mitral regurgitation due to myocardial infarction. In: *Ischemic Mitral Incompetence*. Vetter HO, Hetzer R, Schumtizer H, eds. New York, Springer Verlag, 1991:99-114.
- Murphy-Chutorian D, Knopf W, Moses J, **Heuser RR:** Selected clinical cases using the eclipse laser angioplasty system. In: *Primer of Laser Angioplasty*, 2nd ed., Ginsburg R, Geschwind H, eds. Futura Publishing Co., 1991:309-324.
- Heuser RR:** Experience with different coronary stents. In: *Endoluminal Stenting*. Sigwart U, ed. London, W.B. Saunders Company Ltd., 1996:338-345.
- Heuser RR:** Peripheral stenting for the cardiologist. In: *Frontiers in Interventional Cardiology*. Beyar, Keren, Leon M, Serruys P, eds. London, Martin Dunitz Limited, 1997:377-382.
- Erbel R, O'Neil W, **Heuser RR:** Long Lesion. In: *Tough Calls in Interventional Cardiology*. Safian RD, Freed M, eds. Birmingham, MI, Physician's Press, 1997:48-53.
- Heuser RR, Schatz R, Kimura T:** Tortuous RCA. In: *Tough Calls in Interventional Cardiology*. Safian RD, Freed M, eds. Birmingham, MI, Physician's Press, 1997:76-80.
- Heuser RR, Schatz R, Kimura T:** Tortuous Circumflex. In: *Tough Calls in Interventional Cardiology*. Safian RD, Freed M, eds. Birmingham, MI, Physician's Press, 1997:81-87.
- Heuser RR, Kern M:** Residual stenosis: Vein Graft. In: *Tough Calls in Interventional Cardiology*. Safian RD, Freed M, eds. Birmingham, MI, Physician's Press, 1997:549-551.
- Nobuyoshi M, **Heuser RR, Morice MC:** New filling defects. In: *Tough Calls in Interventional Cardiology*. Safian RD, Freed M, eds. Birmingham, MI, Physician's Press, 1997:555-557.
- Heuser RR, Schatz R, Nobuyoshi M:** Stent: Filling Defects. In: *Tough Calls in Interventional Cardiology*. Safian RD, Freed M, eds. Birmingham, MI, Physician's Press, 1997:599-600.
- Heuser RR, Holmes D, Roubin G:** Free Perforation. In: *Tough Calls in Interventional Cardiology*. Safian RD, Freed M, eds. Birmingham, MI, Physician's Press, 1997:615-617.
- Heuser RR, Holmes D, Roubin G:** Contained Perforation. In: *Tough Calls in Interventional Cardiology*. Safian RD, Freed M, eds. Birmingham, MI, Physician's Press, 1997:618-620.
- Heuser RR, Lass, T, Prebil B, Reid DB:** Computerized three-dimensional intravascular reconstruction. In: *Coronary Stenosis Morphology: Analysis and Implication*. Klein LW, ed. Chicago, IL, Kluwer Academic Publishers, 1997:103-120.
- Heuser RR, Lopez AN:** Introduction. In: *Peripheral Vascular Stenting for Cardiologists* Martin Dunitz Ltd. 1999:1-4.
- Woodfield SL, **Heuser RR:** Angiographic anatomy of the peripheral vasculature and the noninvasive assessment of peripheral vascular disease. In: *Peripheral Vascular Stenting for Cardiologists* Martin Dunitz Ltd. 1999:5-16.
- Heuser, RR, Gupta N, Lopez AN:** AVE Stents. In: *Strategic Approaches in Coronary Interventions, Second Edition*, Lippincott Williams & Wilkins Publishers, 2000.

Heuser RR, Cordero-Yordan H: Carotid Artery Interventions. In: *Practical Handbook of Advanced Interventional Cardiology*, Nguyen T, Saito S, Hu D, Feldman T, Armonk, New York, Futura Publishing Company; In press.

Heuser RR: *PTFE-Covered Stents*. In: *Handbook of Local Drug Delivery*, Martin Dunitz Ltd. Publishing; in press.

Heuser RR, Underwood PL: *Covered Stents in Peripheral Vascular Diseases*. In: *Textbook of Vascular Stenting Interventions*; in press.

Heuser RR: *Certification and Training in Peripheral Vascular Intervention*. In: *Textbook of Vascular Stenting Interventions*; in press.

Cordero H, **Heuser RR**: *The cardiologist and peripheral intervention*. In: *Practical Interventional Cardiology 2nd Edition*, Martin Dunitz Ltd, Publishing: 2002:349-359.

White CW, **Heuser RR**: *The Covered Stent-Graft: Treatment of an Arteriovenous Fistula*. In: *Fifty Cases in Peripheral Intervention*. Martin Dunitz Ltd. Publishing: 2002:71-73.

ABSTRACTS:

Fuchs RM, **Heuser RR**, Brinker JA: Limitations of pulmonary capillary wedge V waves in diagnosing mitral regurgitation. Presented at XI InterAmerican Congress of Cardiology; September 1980.

Heuser RR, Brinker JA: Acute effects of protamine on cardiac function. *Circulation* 1980; 62(3): I-II.

Heuser RR, Brinker JA: Cardiac catheterization in aortic stenosis: "A Necessary Evil". *Clinical Research* 1981; 29:206A.

Alexander E, **Heuser RR**, Weiss JL, Firestein GS, Leitl G, Wagner Jr HN, Stevens MB, Becker LC: Scleroderma heart disease: Evidence for cold induced abnormalities of myocardial perfusion and function. *Circulation* 1981; 64(4): IV-24.

Heuser RR, Becker LC, Flaherty J: Extensive myocardial infarction associated with minor electrocardiographic abnormalities: Value of admission scintigraphy. *Circulation* 1981; 64(4):IV-85.

Heuser RR, Goss JE, Ramo BW, Raff GL, Maddoux GL: Coronary angioplasty in Albuquerque: Report of the first large series and preliminary follow-up. Presented at the American College of Physicians, Albuquerque, New Mexico; December 1984.

Heuser RR, Goss JE, Ramo BW, Raff GL, Maddoux GL: Coronary angioplasty in patients with acute mitral regurgitation due to myocardial infarction: A nonsurgical treatment to preserve mitral valve integrity. Presented at the American Heart Association Meeting, Washington, DC; November 1985.

Heuser RR, Maddoux GL, Goss JE, Ramo BW, Raff GL: Treatment of cardiogenic shock with percutaneous coronary angioplasty: The therapy of choice. Presented at the American College of Cardiology Meeting, Atlanta, Georgia; March 1986.

Heuser RR, Maddoux GL, Goss JE, Ramo BW, Raff GL, Shadoff N: Coronary angioplasty in the elderly: A low risk option. Presented at the World Congress of Cardiology, Washington, DC; September 1986.

- Heuser RR**, Goss JE, Ramo BW, Raff GL, Maddoux GL, Shadoff N: Repeat coronary angioplasty: A highly successful and low risk procedure. Presented at the World Congress of Cardiology, Washington, DC; September 1986.
- Raff GL, **Heuser RR**, Maddoux GL, Goss JE, Ramo BW, Archuleta B: Thallium treadmill testing accurately predicts restenosis following percutaneous transluminal coronary angioplasty. Presented at the World Congress of Cardiology, Washington, DC; September 1986.
- Leatherman GF, Goss JE, Ramo BW, Raff GL, Maddoux GI, **Heuser RR**, Shadoff N, Blake K: Bucindolol improves exercise tolerance and cardiac function in patients with congestive heart failure. Presented at American College of Physicians Meeting, Albuquerque, New Mexico; December 1989.
- Heuser RR**, Goss JE, Ramo BW, Raff GL, Maddoux GI, Shadoff N, Leatherman GF, Blake K: A Randomized, double-blind trial of recombinant human superoxide dismutase (h-SOD) in patients undergoing PTCA for acute MI. Presented at the American College of Physicians Meeting, Albuquerque, New Mexico; December 1989.
- Heuser RR**, Goss JE, Ramo BW, Raff GL, Maddoux GI, Shadoff N, Leatherman GF, Blake K: Laser-assisted coronary angioplasty: Report of initial experience. Presented at the American College of Physicians Meeting, Albuquerque, New Mexico; December 1989.
- Blake K, Goss JE, Ramo BW, Raff GL, Maddoux GL, **Heuser RR**, Shadoff N, Leatherman GF: Adenosine for acute treatment of SVT. Presented at the American College of Physicians Meeting, Albuquerque, New Mexico; December 1989.
- Werns S, Brinker JA, Gruber J, Rothbaum D, **Heuser RR**, George B, Burwell L, Kereiakes D, Mancini GB, Flaherty J: A Randomized double-blind trial of recombinant human superoxide dismutase (h-SOD) in patients undergoing PTCA for acute MI. *Circulation* 1990 (IV):11-113.
- Heuser RR**, Schatz RA, Diethrich EB, Ponder R, Waters DB, Skeete R: Placement of the Palmaz balloon-expandable intraluminal stent in iliac arteries: A single center experience. *Radiology* 1991;181 (supple):295.
- Heuser RR**, Diethrich EB, Waters DB: Abdominal aortic stenosis: Application of Endovascular stents. *Radiology* 1991;181(supple):295.
- Leon MB, Ellis SG, Pichard AD, Baim DS, **Heuser RR**, Schatz RA: Stents may be the preferred treatment for focal aortocoronary vein graft disease. *Circulation* 1991;84 (supple II): II-249.
- Heuser RR**, Dorsey R, Mehta S: The "Smart Needle" - a new device for vascular access. Presented at Congres Internationale d'Angiologie, Paris, France; April 1991.
- Heuser RR**, Mehta S, Ponder R: Stent implantation for stenosis in saphenous vein bypass grafts. Presented at Congres Internationale d'Angiologie, Paris, France; April 1991.
- Ravi R, **Heuser RR**, Diethrich EB, Mehta S, Ponder R: Application of endovascular stents in the abdominal aorta. Presented at Congres Internationale d'Angiologie, Paris, France; April 1991.
- Mehta S, **Heuser RR**, Siegel RM, Willoughby S: Initial report of a new, pulsed, solid-state laser system for percutaneous coronary angioplasty. Presented at Congres Internationale d'Angiologie, Paris, France; April 1991.
- Mehta S, **Heuser RR**, Ponder R: Use of the Palmaz-Schatz balloon-expandable intracoronary stent for the treatment of coronary artery disease. Presented at the American College

of Chest Physicians, Arizona Chapter, Tucson, Arizona; September 1991.

Morton JK, Segal J, Ofili E, **Heuser RR**, Scott N, Aguirre F, St. Vrain J, Labovitz A, FLOMAP Study Group, SLU: A new method for quantitating coronary collateral flow in patients during coronary angioplasty using the Doppler guidewire. (poster) Presented at the American Heart Association Meeting, Anaheim, California; November 1991.

Heuser RR: Use of Holmium: YAG laser in coronary disease. Presented at the International Society for Optical Engineering - Biomedical Optics 1991. Los Angeles, California; January 1992.

Heuser RR, Strumpf RK, Walker CM, Cleman MW, Schatz RA: Coronary stenting in unstable angina: No greater risk than in stable angina. The Journal of the American College of Cardiology 1992;19:178A.

Whitlow PL, Gaspard P, Kent KM, Baim DS, Chapman JB, **Heuser RR**, Knopf WK: Improvement of coronary dissection with a removable flow support catheter: Acute results. The Journal of the American College of Cardiology 1992; 19:217A.

Teirstein PS, Schatz RA, Rocha-Singh KJ, Wong SC, Strumpf RK, **Heuser RR**: Coronary stenting with angioscopic guidance. The Journal of the American College of Cardiology 1992; 19:223A.

Donohue TJ, Kern MJ, Aguirre FV, Bell C, Penick D, Segal J, Ofili E, **Heuser RR**: Determination of the hemodynamic significance of angiographically intermediate coronary stenosis by intracoronary Doppler flow velocity. The Journal of the American College of Cardiology 1992; 19:242A.

Leon MB, Kent KM, Baim DS, Walker CM, Cleman MW, Buchbinder M, **Heuser RR**, Curry C, Schatz RA, JJIS Stent Investigators: Comparison of stent implantation in native coronaries and saphenous vein grafts. The Journal of the American College of Cardiology 1992; 19-263A.

Heuser RR, Diethrich EB, Strumpf RK, Ponder R: Chronic iliac occlusions: Palmaz stents are the treatment of choice. Circulation 1992; 86(supple):I-634.

Heuser RR, Strumpf RK: Coronary stenting of ostial coronary artery stenosis: Intravascular ultrasound guidance. Presented at Cardiostim 1992, Nice, France; June 1992.

Heuser RR, Strumpf RK: The use of angioscopy with coronary stenting. Presented at Cardiostim 1992, Nice, France; June 1992.

Heuser RR, Diethrich EB, Strumpf RK, Ponder R: Chronic iliac occlusions: Intraluminal stents are the treatment of choice. Presented at XIV Congress of European Society of Cardiology, Barcelona, Spain; August 1992.

Heuser RR, Strumpf RK: Angioscopy with coronary stenting: Is it helpful? Presented at XIV Congress of European Society of Cardiology, Barcelona, Spain; August 1992.

Heuser RR: Intravascular coronary artery stents. Presented at the American College of Surgeons, postgraduate courses, New Orleans, Louisiana; October 1992.

Heuser RR, Brinker JA, Weiner BH for Strecker Stent Investigators: Strecker coronary stent: Initial multicenter experience. The Journal of the American College of Cardiology 1993; 21:293A.

- Fischman D, Savage M, Leon M, Schatz RA, **Heuser RR**, Goldberg S: Acute and long-term results of coronary stenting for complex intimal dissections following balloon angioplasty. The Journal of the American College of Cardiology 1993; 21:178A.
- Schatz RA, Penn IM, Baim DS, Nobuyoshi M, Colombo A, Ricci DR, Cleman MW, Goldberg S, **Heuser RR**, Almond D, Fish D, Moses J, Gallup D, Detre K, Leon MB for the STRESS Investigators: Stent Restenosis Study (STRESS) analysis of in-hospital results. Circulation 1993; 88:I-594.
- Heuser RR**, Hardigan K, Strumpf R, Eagan J, Aultman R, Prebil B, Waters K: Single Center Palmaz "Biliary" Stent Experience in Coronary Arteries and Saphenous Vein Grafts. Circulation 1993; 88:(4) Part II.
- White CJ, Ramee SR, Collins TJ, Jain SP, Escobar A, Soni D, Bass TA, **Heuser RR**, Teirstein PS, Bonan R, Nesto RW, Walter PD, Smalling RW, Knopf WD: Angioscopically detected coronary thrombus correlates with adverse PTCA outcome. Circulation 1993; 88:I-596.
- Kussmaul WG, Buchbinder M, Whitlow PL, Aker UT, **Heuser RR**, King SB, Sandza JG, Kensey KR: Randomized trial of a new hemostatic puncture closure device. Circulation 1993; 88:I-72.
- Heuser RR**, Hardigan KR, Strumpf RK, Eagan JT: Single-center Palmaz biliary stent experience in coronary arteries and saphenous veins. Circulation 1993; 88:I-308.
- Fortuna R, **Heuser RR**, Garratt KN, Schwartz R, Buchbinder M: Wiktor intracoronary stent: Experience in the first 101 vein graft patients. Circulation 1993; 88:I-505.
- Teirstein PS, Schatz RA, Rocha-Singh KJ, DeNardo SJ, Morris N, Strumpf RK, **Heuser RR**: Angioscopy of coronary stents at follow-up. The Journal of the American College of Cardiology 1993; 21:134A.
- Thompson MA, Brinker JA, **Heuser RR**, Weiner BH, Hess K, Ross AM, Segal J, The George Washington University Quantitative Analysis Core Laboratory for the Strecker Stent Investigators: Balloon-expandable stent size versus inflation pressure: The Strecker experience. The Journal of the American College of Cardiology 1993; 21:293A.
- Heuser RR**, Hardigan K, Strumpf R, Eagan J, Aultman R, Prebil B, Waters K: Single Center Palmaz "Biliary" Stent Experience in Coronary Arteries and Saphenous Vein Grafts. Presented at the American Heart Association Scientific Session; October 1993.
- Heuser RR**, Strumpf RK, Ramirez-Cardenas J, Schmidt J: Experience with seven stent models in the coronary arteries and vein grafts. Journal Interventional Cardiology 1994; 7:90.
- Fischman D, Savage M, Leon MB, Schatz R, Baim D, Penn I, Detre K, **Heuser RR**, Ricci D, Fish D, Rake R, Gebhardt S, Goldberg S for the STRESS Investigators, Jefferson Medical College, Philadelphia, PA: Acute and late angiographic results of the Stent Restenosis Study (STRESS). The Journal of the American College of Cardiology 1994 (special issue): 60A.
- Segal J, Reiner JS, Thompson MA, Brinker JA, **Heuser RR**, Weiner BH, Ross AM, The George Washington University Quantitative Analysis Core Laboratory for the Strecker Stent Investigators: Residual stenosis following coronary stenting do not predict restenosis in the Strecker stent. The Journal of the American College of Cardiology 1994 (special issue):134A.
- Carrozza JP, Kuntz RE, Leon MB, Schatz RA, Brinker J, Chuang YC, **Heuser RR**, Baim DS: Demographic, angiographic and procedural factors associated with late angiographic outcome in the STRESS Trial: A multivariable analysis. Circulation 1994; 90:I-18.

- Garratt KN, White CJ, Buchbinder M, Whitlow PK, **Heuser RR** on behalf of the American Wiktor Stent Investigators: Wiktor stent placement for unsuccessful coronary angioplasty. *Circulation* 1994; 90:I-279.
- Heuser RR**, Cleman M, Cabin H, Wong SC, Chuang YC, Bailey S, Nobuyoshi M, Almond D, Schatz R: The LAD subgroup in the Stent Restenosis Study: Early and late angiographic and clinical outcomes. *Circulation* 1994; 90:I-323.
- Savage M, Fischman D, Rake R, Hirshfeld J, Penn I, **Heuser RR**, Shakhovich A, Moses J, Ricci D, Goldberg S: Effect of lesion morphology on angiographic outcome after balloon angioplasty and coronary stenting: Results from the STRESS Trial. *Circulation* 1994; 90:I-324.
- Kussmaul WG, Sandza JG, **Heuser RR** for the Angio-Seal™ in patients at high risk of bleeding complications. *Catheterization and Cardiovascular Interventions* 1994; 32:98.
- Sketch Jr. MH, Wong SC, Chuang YC, Phillips HR, **Heuser RR**, Savage M, Stack RS, Balm DS, Schatz RA, Leon MB for the JJIS Stent Investigators: Progressive deterioration in late (2-Year) clinical outcomes after stent implantation in saphenous vein grafts: The multicenter JJIS experience. *The Journal of the American College of Cardiology* 1995 (special issue): 79A.
- Savage M, Douglas J, Fischman D, Fenton S, King S, Pepine C, Bailey S, Overlie P, Werner J, Leon M, **Heuser RR**, Brinker J, Buchbinder M, Smalling R, Snead D, Rake R, Gebhardt S, Kerensky R, Wargovich T, Goldberg S, SAVED Trial Investigators: Coronary stents versus balloon angioplasty for aortocoronary saphenous vein bypass graft disease: Interim results of a randomized trial. *The Journal of the American College of Cardiology* 1995 (special issue):79A.
- Wong SC, Chuang YC, Detre K, Pichard AD, Schatz RA, **Heuser RR**, Kent KM, Leon MB: Is the anti-restenosis effect of stent (vs PTCA) greater in LAD vessels? A subgroup analysis of the Stent Restenosis Study. *The Journal of the American College of Cardiology* 1995 (special issue):374A.
- Primack DS, **Heuser RR**, Kuntz RE, Waters K, Swiderski K: Coronary stenting performed via brachial access is more economical. Presented at the American College of Cardiology 44th Annual Scientific Session, New Orleans, Louisiana; March 1995.
- Primack DS, **Heuser RR**, Kuntz RE, Waters K, Swiderski K: Coronary stenting performed via brachial access is more economical. Presented at the 18th Annual Meeting of the Society for Cardiac Angiography and Interventions, Orlando, Florida; May 1995.
- Heuser RR**, Reynolds GT, Papazoglou C, Diethrich EB: Endoluminal grafting for percutaneous aneurysm in an aortocoronary saphenous vein graft: The first human experience. Presented at the 18th Annual Meeting of the Society for Cardiac Angiography and Interventions, Orlando, Florida; May 1995.
- Garratt KN, Schwartz RS, Holmes Jr DR, White CJ, Buchbinder M, Whitlow PL, **Heuser RR** on behalf of the American Wiktor Stent Investigators: Placement of Wiktor coronary stents for management of unsuccessful coronary angioplasty: The American Experience. Presented at the 1st International Meeting on Interventional Cardiology, Jerusalem, Israel; June 1995. *The Journal of Invasive Cardiology* 1995; 7 (supple)35C.
- Heuser RR**, Reynolds GT, Papazoglou C, Diethrich EB, Mukherjee R: Compassionate endoluminal grafting for percutaneous aneurysm exclusion in aortocoronary saphenous vein

grafts. Presented at the XVIIth Congress of the European Society of Cardiology, Amsterdam, The Netherlands; August 1995. *Journal of Endovascular Surgery* 1995; 2:81-88.

Heuser RR, Wong SC, Chuang YC, Detre K, Pichard AD, Schatz RA, Kent KM, Leon MB for the STRESS investigators: The LAD subgroup in the Stent REStenosis Study (STRESS): The most pronounced anti-restenosis effect of stenting. Presented at the XVIIth Congress of the European Society of Cardiology, Amsterdam, The Netherlands; August 1995.

Heuser RR, Reynolds GT, Papazoglou C, Diethrich EB, Mukherjee R: Compassionate endoluminal grafting for percutaneous treatment of aortocoronary SVG disease. *The Journal of the American College of Cardiology* 1996; 27:179A.

Savage M, Fischman D, Rake R, Schatz R, Penn I, Nobuyoshi M, Moses J, **Heuser RR**, Gebhardt S, Goldberg S for the STRESS Trial Investigators: Elective coronary stenting versus balloon angioplasty in smaller native coronary arteries: Results from STRESS. *The Journal of the American College of Cardiology* 1996; 27:253A.

Leon MB, Ellis SG, Moses J, King SB, **Heuser R**, Kent KM, Goldbert S, Fischman D, Senerchia C, Lanoue AS, Kuntz RE: Interim Report from the Reduced Anticoagulation Vein Graft Stent (RAVES) Study. Presented at the American Heart Association 69th Scientific Session, New Orleans, Louisiana; November 1996.

Savage M, Fischman D, Slota P, Rake R, Leon M, Schatz R, Moses J, Penn I, Nobuyoshi M, **Heuser RR**, Goldberg S: Coronary intervention in the diabetic patient: Improved outcome following stent implantation versus balloon angioplasty. Presented at the American College of Cardiology 46th Annual Scientific Session, Anaheim, California; March 1997.

Dean LS, **Heuser RR**, Moore PB, Rogers EW, Roubin GS, Leon MB, Voorhees WD, Holmes Jr DR, O'Shoughnessy CD, Zidar JP: Stenting in small vessels: A re-evaluation using the GR-II intracoronary stent, a Cook Registry Study. Presented at the American College of Cardiology 46th Annual Scientific Session, Anaheim, California; March 1997.

Heuser RR: Live Demonstrations. Presented at the 1997 Japanese Society of Interventional Cardiology Live Demonstration Symposium, Kokura, Japan; May 1997.

Heuser RR: Influence of minimally invasive coronary artery bypass surgery. Presented at the XIXth Congress of the European Society of Cardiology, Stockholm, Sweden; August 1997.

Heuser RR, Kuntz RE, Lansky AJ, Whitlow PL, Sanfian RD, Yeung AC, Senerchia C, Cutlip DE, Pino-Mauch B, Pedan A: The SMART Trial: Acute outcome indicates superior efficacy with the AVE Stent. Presented at the XIXth Congress of the European Society of Cardiology, Stockholm, Sweden; August 1997.

Heuser RR, Kuntz R, Popma J, Pino-Mauch B, Miller R, Yeung A, Badger R: The SMART Trial: 30-day outcome indicates superior efficacy with the AVE Stent. Presented at the Transcatheter Cardiovascular Therapeutics Symposium, Washington, DC; September 1997.

Heuser RR: Influence of modern bypass surgery on PTCA. Presented at 20 Years of PTCA - Back to the Cradle, Zurich, Switzerland; September 1997.

Heuser RR: An overview of new stent designs: Lesion specific stenting. Presented at Coronary Interventions 1997, LaJolla, California; October 1997.

Heuser RR: Covered stents: Clinical needs, future prospects. Presented at Coronary Interventions 1997, LaJolla, California; October 1997.

- Heuser RR**, Kuntz RE, Lansky AJ, Whitlow PL, Safian RD, Yeung AC, Senerchia C, Cutlip DE, Pino-Mauch B, Pedan A: The SMART Trial: Acute outcome indicates superior efficacy with the AVE Stent. American Heart Association 70th Scientific Sessions, Orlando, Florida; November 1997.
- Heuser RR**: Is carotid artery PTA superior to surgery? Presented at the International Course on Interventional Cardiology, Frankfurt, Germany; December 1997.
- Heuser RR**, Kuntz R, Popma J, Pino-Mauch B, Miller R, Yeung A, Badger R: The SMART Trial: 30-day outcome indicates superior efficacy with the AVE Stent. Presented at the American College of Cardiology 47th Annual Scientific Session, Atlanta, Georgia; April 1998.
- Heuser RR**, Kuntz R, Lansky A, Kalon KLH, Reduto L, Badger R, Iannone LA, Whitlow P, Vlietstra R: A comparison of the Long AVE Micro™ Stent II and the Palmaz-Schatz® Stent: A SMART trial registry. Presented at the American College of Cardiology 47th Annual Scientific Session, Atlanta, Georgia; April 1998.
- Heuser RR**, Houser F, Battaglia S, Tarkington L, Simon A, Culler S, Becker E: Repeat procedure rates in percutaneous interventions: Is there added benefit of atherectomy or stenting? Presented at Transcather Cardiovascular Therapeutics, Washington, DC; October 1998.
- Heuser RR**, Lopez AN, Stoerger H, Reifart N: Coronary artery application of an endoluminal polytetrafluorethylene stent graft: Two center experience with the JOMED® JoStent®. Presented at American Heart Association, Dallas, Texas; November 1998.
- Ramee SR, Baim DS, Popma JJ, Ho KKL, Cutlip DE, Lanoue AS, Dandreo KJ, Schatz RA, Sharma SK, Kuntz RE, **Heuser RR**: A randomized, prospective, multi-center study comparing intracoronary Urokinase to Rheolytic thrombectomy with the POSSIS AngioJet Catheter for intracoronary thrombus: Final results of the VeGAS 2 Trial. Presented at American Heart Association, Dallas, Texas; November 1998.
- Heuser RR**, Lopez AN: Coronary Artery Disease Revascularization with the JOMED JoStent®: A Two Center Experience. Presented at American College of Cardiology, New Orleans, Louisiana; March 1999.
- Lansky AJ, Popma JJ, Mehran R, Hanzel G, Cutlip D, Ho KK, Proctor B, Happe JA, Kuntz RE, Baim DS, **Heuser RR**, Midei M, O'Shaughnessy C, Leon MB: Tubular Slotted Stents: A "Breakthrough Therapy" for Women Undergoing Coronary Interventions. Pooled Results from the STARS, ACSSENT, SMART, and NIRVANA Randomized Clinical Trials. Presented at American College of Cardiology, New Orleans, Louisiana; March 1999.
- Oesterle SN, Yeung A, Ali N, Dean LS, **Heuser RR**, Knopf WD, Masden R, Ramee SR, Reeder GS, Resar J, Sanborn TA, Schaer GL, Schofield PM: The CardioGenesis Percutaneous Myocardial Revascularization (PMR) Randomized Trial: Initial Clinical Results. Presented at American College of Cardiology, New Orleans, Louisiana; March 1999.
- Heuser RR**: New Findings of PMR and Gene Therapy. Presented at 4th Asian-Pacific Society of Interventional Cardiology Conference and 8th Live Demonstration in Kokura, Kokura, Japan; May 1999.
- Heuser RR**: The Use of Covered Coronary Stents: A new Therapy in Evolution. Presented at 4th Asian-Pacific Society of Interventional Cardiology Conference and 8th Live Demonstration in Kokura, Kokura, Japan; May 1999.
- Heuser RR**: Radiofrequency Enhancement of Intramyocardial Gene Therapy. Presented at the Second Annual Symposium Angiogenesis and DMR Agenda, Washington, DC; June 1999.

- Heuser RR:** Covered Stents for Coronary Artery Disease: Early Results of Clinical Trials. Presented at the Sixth Annual Mayo Interventional Cardiology Symposium, Napa Valley, California; July 1999.
- Heuser RR:** Peripheral Vascular Disease Management: New Devices, New Drugs, New Approaches. Presented at the Sixth Annual Mayo Intervention Cardiology Symposium, Napa Valley, California; July 1999.
- Heuser RR:** Radiofrequency PMR: A Unique System for Simultaneous Radiofrequency PMR and Gene Therapy. Presented at the XXist Congress of the European Society of Cardiology, Barcelona, Spain; August 1999; 20;2828.
- Heuser RR:** Do Coronary Stents Improve Clinical Outcomes? A retrospective study comparing stent device utilization and adverse outcomes. Presented at the XXist Congress of the European Society of Cardiology, Barcelona, Spain; August 1999.
- Oesterle SN, Sanborn TA, Ali N, Resar J, Ramee SR, **Heuser RR**, Dean L, Knopf W, Schofield P, Schaer GL, Reeder G, Masden R, Yeung AC, Burkhoof D: Percutaneous transmyocardial laser revascularization for severe angina: the PACIFIC randomized trial. Presented at the 72nd Scientific Session American Heart Association, Atlanta, Georgia, November 1999.
- Sameer Mehta, **Heuser RR**, Salvatore BL, Simon AW, Culler SD, Becker ER: Evolving Patterns of Coronary Stenting: Report from a Large National Database. Presented at the 49th Annual Scientific Session American College of Cardiology, March 2000.
- Heuser RR:** Future of Covered Stents and Endoluminal Grafts. Presented at the GET International Endovascular Peripheral Course in Monaco; May 2000.
- Heuser RR**, Heilbrunn S, Fogarty TJ, Motarjeme A: Establishing an Integrated Vascular Center. Presented at the Eighth New Devices Seminar for the Cardiac and Peripheral Vascular Laboratories in Orlando, Florida; July 2000.
- Heuser RR**, Covered Stents: New Options & Strategies. Presented at the Eighth New Devices Seminar for the Cardiac and Peripheral Vascular Laboratories in Orlando, Florida; July 2000.
- Heuser RR**, Waksman R, White CJ: Advanced Coronary Workshop: The experts discuss techniques and equipment on the Progress Occurring with Stent Grafts & Brachytherapy for the Coronary and Bypass Graft Circulation. Presented at the Eight New Devices Seminar for the Cardiac and Peripheral Vascular Laboratories in Orlando, Florida; July 2000.
- Heuser RR**, Cordero H, Underwood PL, Schroeder WS, Weirick BA, Hatler CW: Single Center Experience with OCR Guidance in the Treatment of Chronic Total Occlusions (CTO). Submitted to Transcatheter Cardiovascular Therapeutics; September 2001.
- Cordero H, Underwood PL, Schroeder WS, Weirick BA, Hatler CW, **Heuser RR**: Fiber Optic Guidance Technology in the Treatment of Chronic Total Occlusions: Initial Experience. Presented at 12th Annual Symposium Transcatheter Cardiovascular Therapeutics; October 2000.
- Laskey WK, **Heuser RR**, Suntharalingam M, Bass TA, Silber S, Rutherford BD, Wilmer C, Berke AD, Snell RJ, Popma J, Kuntz RE: Effects of ⁹⁰Sr/⁹⁰Beta Radiation on Diabetic Patients with In-Stent Restenosis. Presented at AHA November 2000.
- Heuser RR**, Cordero H, Underwood PL, Schroeder WS, Weirick BA, Hatler CW: OCR Can Be Used to Safely Treat Chronic Total Occlusions. Presented at AHA; November 2000.

Heuser RR, Cordero H, Underwood PL, Schroeder WS, Weirick BA, Hatler CW: Single Center Experience with OCR Guidance in the Treatment of Chronic Total Occlusions (CTO). *Circulation* October 23, 2001; Vol 104 (17) 1978.

Heuser RR, Cordero H, Schroeder W, Weirick BA, Hatler CW, Underwood PL: Fiberoptic Guidance System for Treatment of Chronic Total Occlusions: Single Center Experience. Presented at ACC 2001; March 2001.

Heuser RR, Lasky W, Bonan R, Lansky A: B-Radiation in Lesions Longer Than 20 Millimeters: A START 40/20 Subgroup Analysis. TCT 2001: September 2001.

Heuser RR, Underwood PL, Schroeder W, Weirick BA, Hatler CW: Experience with OCR Guidance in the Treatment of Chronic Total Occlusions. TCT 2001: September 2001.

PA Morales, **RR Heuser**, W Laskey, R Bonan, AJ Lansky, JJ Popma: Treating In-Stent Restenosis in Long Coronary Lesions. Presented at the 51st Annual Scientific Session American College of Cardiology; March 2002.

PA Morales, **RR Heuser**, EA Weirick, CW Hatler: Predicting Success in Crossing Chronic Total Occlusions With a New Guidewire. Presented at the 51st Annual Scientific Session American College of Cardiology; March 2002.

WK Laskey, M Suntharalingam, J Popma, **R Heuser**, B Rutherford, P Teirstein, A Lansky, R Kuntz, R Bonan: Efficacy of SR-90 Beta Radiation for the Treatment of In-Stent Restenosis: 24-Month Clinical Outcomes from the Stents And Radiation Therapy Trial (START). Presented at the 51st Annual Scientific Session American College of Cardiology; March 2002.

P.A. Morales, A. Reese, W. Schroeder, **R.R. Heuser**, New Treatment for Chronic Total Occlusions of Femoral Arteries. Presented at the Fourteenth Annual Symposium Transcatheter Cardiovascular Therapeutics; September, 2002

AWARDS & HONORS:

Columbia/HCA Cardiovascular Management Network - 1998 Cardiologist of the Year

PATENTS:

1. Method and Apparatus for Treating Body Tissues and Bodily Fluids; Patent granted December 12, 2000 Number: 6,159,197
2. Hot Tip Catheter; Patent granted February 20, 2001 Number: 6,190,379
3. Embolism Prevention Device; Patent granted April 2, 2002 Number: 6,364,900
4. Catheter apparatus and Method for Arterializing a Vein; Patent granted October 15, 2002 Number 6,464,665
5. Methods and apparatus for treating body tissues and bodily fluid vessels; Patent granted October 15, 2002 Number: 6,464,681
6. Catheter for Thermal Evaluation of Arteriosclerotic Plaque; Patent granted March 25, 2003 Number: 6,536,949
7. Small Diameter Snare; Patent granted April 29, 2003 Number: 6,554,842

**EXHIBIT
B**

DISCLOSURES

**APPLICATION
SERIAL NO. 09/836,750**

EXHIBIT B
DISCLOSURES
APPLICATION SERIAL NO. 09/836,750

PAGE 20, LINE 10 – PAGE 21, LINE 15

Growth factors can be utilized to induce the growth of “hard tissue” or bone and “soft tissues” like ectodermal and mesodermal tissues. As used herein, the term growth factor encompasses compositions and living organisms which promote the growth of hard tissue, such as bone, or soft tissue, in the body of a patient. The compositions include organic and inorganic matter. The compositions can be genetically produced or manipulated. The living organisms can be bacteria, viruses, or any other living organism which promote tissue growth. By way of example and not limitation, growth factors can include platelet-derived growth factor (PDGF), epidermal growth factor (EGF), fibroblast growth factor (acidic/basic (FGF a,b), interleukins (IL’s), tumor necrosis factor (TNF), transforming growth factor (TGF-B), colony-stimulating factor (CSF), osteopontin (Eta-1 OPN), platelet-derived growth factor (PDGF), interferon (INF), bone morphogenic protein 1 (BMP-1), and insulin growth factor (IGF). Recombinant and non-recombinant growth factors can be utilized as desired. Bacteria or viruses can, when appropriate, be utilized as growth factors. For example, there is a bacterial hydrophilic polypeptide that self-assembles into a nanometer internal diameter pore to build a selective lipid body. Various enzymes can be utilized for the synthesis of peptides which contain amino acids that control three-dimensional protein structure and growth. Growth factors can be applied in gels or other carriers which regulate the rate of release of the growth factors and help maintain the growth factors and the carrier, at a desired location in the body. Time release capsules, granules, or other carriers containing growth factor can be activated by tissue pH, by enzymes, by ultrasound,

by electricity, by heat, by selected *in vivo* chemicals or by any other selected means to release the growth factor. The carrier can be resorbable or non-resorbable. Or, the growth factor itself can be activated by similar means. Either the carrier or the growth factor can mimic extracellular fluid to control cell growth, migration, and function. The growth factor can be administered orally, systemically, in a carrier, by hypodermic needle, through the respiratory tract, or by any other desired method. The growth factor can also be administered into a capsule or other man-made composition or structure placed in the body. While administration of the growth factor is presently usually localized in the patient's body, circumstances may arise where it is advantageous to distribute a growth factor throughout the patient's body in uniform or non-uniform concentrations. An advantage to growth factors is that they can often, especially when in capsule form or in some other containment system, be inserted to a desired site in the body by simply making a small incision and inserting the growth factor. The making of such small incision comprises minor surgery which can often be accomplished on an out-patient basis. The growth factors can be multifactorial and nonspecific.

PAGE 44, LINE 19 – PAGE 46, LINE 16

Genetic material comprising a portion of a gene, a gene, genes, a gene product (i.e., a composition a gene causes to be produced like, for example, an organ-producing growth factor), growth factor, or an ECM (extracellular matrix) can be used in or on the body to grow an organ to tissue. For example, the vascular epithelial growth factor gene (VEGF) or its growth factor equivalent can be inserted into the body to cause an artery to grow. When insertion of a gene, portion of a gene, gene product, growth factor, or ECM *in vivo* or *ex vivo* is referred to herein in connection with any of the implant techniques of the invention, it is understood that a cell

nutrient culture(s), physiological nutrient culture(s), carrier (s), enhancer(s), promoter(s), or any other desired auxiliary component(s) can be inserted with the gene or at the same location as the gene, growth factor, ECM, etc.

An artery is an organ from the circulatory system. An artery can be grown in the heart, legs, or other areas by injecting a gene or other genetic material into muscle at a desired site. Size, vascularity, simplicity of access, ease of exploitation, and any other desired factors can be utilized in selecting a desired site. The gene is one of several known VEGF genes which cause the production of vascular endothelial growth factors. Several VEGF genes which produce vascular endothelial growth factors are believed to exist because nature intends for there to be several pathways (i.e., genes) which enable the production of necessary growth factors. The existence of several pathways is believed important because if one of the genes is damaged or inoperative, other similar genes can still orchestrate the production of necessary growth factors. VEGF genes are used by the body to promote blood vessel growth. VEGF genes are assimilated (taken in) by muscle cells. The genes cause the muscle cells to make a VEGF protein which promotes the growth of new arteries. VEGF proteins can be made in a lab and injected into a patient intravenously, intraluminally, or intramuscularly to promote the growth of an artery. Or, the genes (or other genetic material) can be applied with an angioplasty balloon, with the assistance of a vector, or by any other method.

It is not always desirable to grow a completely new organ. Sometimes growing a portion of an organ is desirable. For example, in some heart attacks or strokes, a portion of the heart or brain remains viable and a portion dies. An injection of a gene to form cardiac muscle and/or an injection of a gene to form an artery can be utilized to revive or replace the dead portion of the heart. The dead portion of the heart may (or may not) be used as a matrix while the new muscles

and vessels grow. Thus, in this example, a partial new organ is grown in a pre-existing organ. A pacemaker may (or may not) be necessary. A second injection of a gene may (or may not) be necessary to stop cardiac muscle growth once it is completed. Portions of organs throughout the body can similarly be repaired or replaced. It may be necessary to provide gene(s) or growth factor(s) sequentially. For instance, one or more blood vessels are grown by inserting an appropriate gene or other genetic material into a selected area. Second, an appropriate gene or other genetic material is inserted in the selected area to grow a bone or other organ.

The size and shape limitation of the desired structure can come from a containment and boundary contact inhibition phenomenon or by a chemical inhibition.

A variation on the theme of growing a portion of an organ is as follows: a portion of a heart dies. The pericardium is utilized as a scaffold and seeded with cells and/or genes to grow new muscle, and genes (or other genetic material) to grow new arteries. Immediately adjacent the dead cardiac muscle, onto or into the pericardium, the appropriate cells, genes, and/or growth factors (or other genetic material) are placed. Once the new muscle and blood vessels have grown, the function specific tissue can be applied to the damaged portion of the heart and paced, if necessary, to augment cardiac action. If the surgeon desires, the dead muscle can be removed and the new muscle and blood vessels can be surgically rotated into the excised region and secured. This probably can be done endoscopically. In essence, the pericardium is utilized to allow the new muscle wall to grow. The new muscle wall is then transplanted into the damaged heart wall. This procedure utilizes the body as a factor to grow an organ and/or tissue, after which the organ and/or tissue is transplanted to a desired region. On the other hand, the new muscle wall may integrate itself into the old wall and not require transplantation.

EXHIBIT C

DEFINITIONS

EXHIBIT C

DEFINITIONS

PAGE 20, LINE 10 – PAGE 21, LINE 15

Growth factors can be utilized to induce the growth of “hard tissue” or bone and “soft tissues” like ectodermal and mesodermal tissues. As used herein, the term growth factor encompasses compositions and living organisms which promote the growth of hard tissue, such as bone, or soft tissue, in the body of a patient. The compositions include organic and inorganic matter. The compositions can be genetically produced or manipulated. The living organisms can be bacteria, viruses, or any other living organism which promote tissue growth. By way of example and not limitation, growth factors can include platelet-derived growth factor (PDGF), epidermal growth factor (EGF), fibroblast growth factor (acidic/basic (FGF a,b), interleukins (IL's), tumor necrosis factor (TNF), transforming growth factor (TGF-B), colony-stimulating factor (CSF), osteopontin (Eta-1 OPN), platelet-derived growth factor (PDGF), interferon (INF), bone morphogenic protein 1 (BMP-1), and insulin growth factor (IGF). Recombinant and non-recombinant growth factors can be utilized as desired. Bacteria or viruses can, when appropriate, be utilized as growth factors. For example, there is a bacterial hydrophilic polypeptide that self-assembles into a nanometer internal diameter pore to build a selective lipid body. Various enzymes can be utilized for the synthesis of peptides which contain amino acids that control three-dimensional protein structure and growth. Growth factors can be applied in gels or other carriers which regulate the rate of release of the growth factors and help maintain the growth factors and the carrier, at a desired location in the body. Time release capsules, granules, or other carriers containing growth factor can be activated by tissue pH, by enzymes, by ultrasound,

by electricity, by heat, by selected *in vivo* chemicals or by any other selected means to release the growth factor. The carrier can be resorbable or non-resorbable. Or, the growth factor itself can be activated by similar means. Either the carrier or the growth factor can mimic extracellular fluid to control cell growth, migration, and function. The growth factor can be administered orally, systemically, in a carrier, by hypodermic needle, through the respiratory tract, or by any other desired method. The growth factor can also be administered into a capsule or other man-made composition or structure placed in the body. While administration of the growth factor is presently usually localized in the patient's body, circumstances may arise where it is advantageous to distribute a growth factor throughout the patient's body in uniform or non-uniform concentrations. An advantage to growth factors is that they can often, especially when in capsule form or in some other containment system, be inserted to a desired site in the body by simply making a small incision and inserting the growth factor. The making of such small incision comprises minor surgery which can often be accomplished on an out-patient basis. The growth factors can be multifactorial and nonspecific.

MEDLINE PLUS: MERRIAM-WEBSTER MEDICAL DICTIONARY
A SERVICE OF THE U.S. NATIONAL LIBRARY OF MEDICINE AND THE
NATIONAL INSTITUTES OF HEALTH

Growth factor: a substance (as a vitamin B₁₂ or an interleukin)
that promotes growth and especially cellular growth

EXHIBIT D

CLAIMS

EXHIBIT D

CLAIMS

Claim X: A method for growing a new portion of a pre-existing heart comprising the steps of: placing a growth factor in a body of a human patient and growing new muscle in said heart.

EXHIBIT E

PUBLICATIONS

EXHIBIT E

PUBLICATION INFORMATION SUMMARY

TITLE	AUTHOR	CITATION	DATE	AUTHOR COUNTRY	ROUTE OF ADMINISTRATION	GROWTH FACTOR ADMINISTERED	RESULT
Left Ventricular Electromechanical Mapping to Assess Efficacy of phVEGF165 Gene Transfer for Therapeutic Angiogenesis in Chronic Myocardial Ischemia	Vale	Circulation. 2000; 102:965-974	08/29/00	U.S.	Small incision (minithoracotomy) with syringe injection	VEGF (Gene form)	Repair of damaged portion of heart – Also pertains to new muscle growth
Repair of Infarcted Myocardium by Autologous Intracoronary Mononuclear Bone Marrow Cell Transplantation in Humans	Strauer	Circulation. 2002; 106:1913-1918	10/08/02	Germany	Balloon catheter with injection	Bone Marrow Cells	Repair of dead portion of heart – also pertains to new muscle growth

TITLE	AUTHOR	CITATION	DATE	AUTHOR COUNTRY	ROUTE OF ADMINISTRATION	GROWTH FACTOR ADMINISTERED	RESULT
Viability and differentiation of autologous skeletal myoblast grafts in ischemic cardiomyopathy	Hagege	Lancet 2003 Feb 8; 361 (9356):491-492	2003	France	Injection	Skeletal Muscle Cells	Repair of dead portion of heart; Histological Proof (muscle)
Autologous Cell Transplant Helpful in Ischemic Heart or Legs	Barclay	Medscape Medical News 2000 – Abstract from American Heart Association's 75 th Scientific Sessions on 11/18/02, Chicago	11/18/02	U.S.	Surgery with syringe injection	Bone Marrow Cells	Repair of damaged portion of heart – also pertains to new muscle growth
Autologous skeletal myoblasts transplanted to ischemia-damaged myocardium in humans. Histological analysis of cell survival and differentiation	Pagani	J Am Coll Cardiol 2003 Mar 5; 41(5): 879-888	2003	U.S.	Surgery with syringe injection	Skeletal Muscle Cells	Repair of dead portion of heart; Histological Proof (muscle and blood vessels)

Clinical Investigation and Reports

Repair of Infarcted Myocardium by Autologous Intracoronary Mononuclear Bone Marrow Cell Transplantation in Humans

Bodo E. Strauer, MD; Michael Brehm, MD; Tobias Zeus, MD; Matthias Köstering, MD; Anna Hernandez, PhD; Rüdiger V. Sorg, PhD; Gesine Kögler, PhD; Peter Wernet, MD

Background—Experimental data suggest that bone marrow–derived cells may contribute to the healing of myocardial infarction (MI). For this reason, we analyzed 10 patients who were treated by intracoronary transplantation of autologous, mononuclear bone marrow cells (BMCs) in addition to standard therapy after MI.

Methods and Results—After standard therapy for acute MI, 10 patients were transplanted with autologous mononuclear BMCs via a balloon catheter placed into the infarct-related artery during balloon dilatation (percutaneous transluminal coronary angioplasty). Another 10 patients with acute MI were treated by standard therapy alone. After 3 months of follow-up, the infarct region (determined by left ventriculography) had decreased significantly within the cell therapy group (from 30 ± 13 to $12 \pm 7\%$, $P=0.005$) and was also significantly smaller compared with the standard therapy group ($P=0.04$). Likewise, infarction wall movement velocity increased significantly only in the cell therapy group (from 2.0 ± 1.1 to 4.0 ± 2.6 cm/s, $P=0.028$). Further cardiac examinations (dobutamine stress echocardiography, radionuclide ventriculography, and catheterization of the right heart) were performed for the cell therapy group and showed significant improvement in stroke volume index, left ventricular end-systolic volume and contractility (ratio of systolic pressure and end-systolic volume), and myocardial perfusion of the infarct region.

Conclusions—These results demonstrate for the first time that selective intracoronary transplantation of autologous, mononuclear BMCs is safe and seems to be effective under clinical conditions. The marked therapeutic effect may be attributed to BMC-associated myocardial regeneration and neovascularization. (*Circulation*. 2002;106:1913-1918.)

Key Words: myocardial infarction ■ cell transplantation, intracoronary ■ angiogenesis ■ bone marrow ■ myogenesis

Remodeling of the left ventricle after myocardial infarction (MI) represents a major cause of infarct-related heart failure and death. This process depends on acute and chronic transformation of both the necrotic infarct region and the non-necrotic, peri-infarct tissue.^{1,2} Despite application of pharmacotherapeutics and mechanical interventions, the cardiomyocytes lost during MI cannot be regenerated. The recent finding that a small population of cardiac muscle cells is able to replicate itself is encouraging but is still consistent with the concept that such regeneration is restricted to viable myocardium.³

In animal experiments, attempts to replace the necrotic zone by transplanting other cells (eg, fetal cardiomyocytes or skeletal myoblasts) have invariably succeeded in reconstituting heart muscle structures, ie, myocardium and coronary vessels. However, these cells fail to integrate structurally and do not display characteristic physiological functions.⁴⁻⁷ Another approach to reverse myocardial remodeling is to repair myocardial tissue by using bone marrow–derived cells. Bone

marrow contains multipotent adult stem cells that show a high capacity for differentiation.⁸⁻¹⁰ Experimental studies have shown that bone marrow cells (BMCs) are capable of regenerating infarcted myocardium and inducing myogenesis and angiogenesis; this leads in turn to amelioration of cardiac function in mice and pigs.¹¹⁻¹⁴ However, procedures based on this phenomenon remain largely uninvestigated in a human clinical setting.

An investigation of one patient receiving autologous skeletal myoblasts into a postinfarction scar during coronary artery bypass grafting revealed improvement of contraction and viability 5 months afterward.¹⁵ Autologous mononuclear BMCs transplanted in a similar surgical setting showed long-term improvement of myocardial perfusion in 3 of 5 patients and no change in 2 patients.¹⁶ However, such studies entail a surgical approach and are therefore associated with well-known perioperative risks. Moreover, this surgical procedure cannot be used with MI. We therefore looked for a nonsurgical, safer mode for transplanting autologous cells

Received August 2, 2002; accepted August 2, 2002.

From the Department of Medicine, Division of Cardiology (B.E.S., M.B., T.Z., M.K.) and Institute for Transplantation Diagnostics and Cell Therapeutics (A.H., R.V.S., G.K., P.W.), Heinrich-Heine-University of Düsseldorf, Germany.

Correspondence to Professor Dr Bodo E. Strauer, Department of Medicine, Division of Cardiology, Heinrich-Heine-University, Moorenstr 5, 40225 Düsseldorf, Germany. E-mail Strauer@med.uni-duesseldorf.de

© 2002 American Heart Association, Inc.

Circulation is available at <http://www.circulationaha.org>

DOI: 10.1161/01.CIR.0000034046.87607.1C

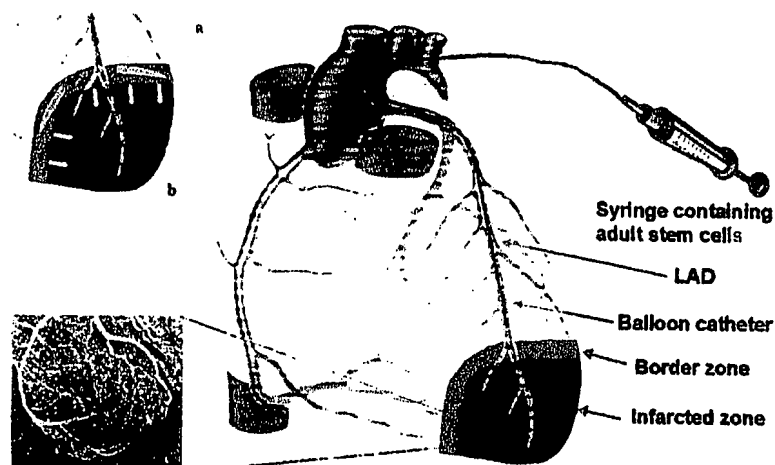


Figure 1. Procedure of cell transplantation into infarcted myocardium in humans. *a*, The balloon catheter enters the infarct-related artery and is placed above the border zone of the infarction. It is then inflated and the cell suspension is infused at high pressure under stop-flow conditions. *b*, In this way, cells are transplanted into the infarcted zone via the infarct-related vasculature (red dots). Cells infiltrate the infarcted zone. Blue and white arrows suggest the possible route of migration. *c*, A supply of blood flow exists within the infarcted zone.³⁵ The cells are therefore able to reach both the border and the infarcted zone.

into postinfarction tissue. A pilot study from our group demonstrated that intracoronary transplantation of autologous mononuclear BMCs 6 days after MI was associated with a marked decrease in infarct area and an increase in left ventricular (LV) function after 3 and 6 months of follow-up.¹⁷ To confirm these results and validate this promising new therapy for MI, we established a clinical trial involving 20 patients for comparing the safety and bioefficacy of autologous BMC transplantation. All 20 patients underwent standard therapy, and 10 patients received additional intracoronary cell transplantation. All 20 patients were followed up for 3 months.

Methods

Patient Population

All 20 patients had suffered transmural infarction according to World Health Organization criteria with the involvement of the left anterior descending coronary artery ($n=4$), left circumflex coronary artery ($n=3$), or right coronary artery ($n=13$). Mean duration of infarct pain was 12 ± 10 hours before invasive diagnostics and therapy. Patients had to be <70 years old and were excluded if one of the following criteria were met: screening >72 hours after infarction, cardiac shock, severe comorbidity, alcohol or drug dependency, or excessive travel distance to the study center.

After right and left heart catheterization, coronary angiography, and left ventriculography, mechanical treatment was initiated with recanalization of the infarct-related artery by balloon angioplasty ($n=20$) and subsequent stent implantation ($n=19$). All patients were monitored in our intensive care unit, and no arrhythmogenic events or hemodynamic impairments were recorded in either patient group.

All 20 patients were briefed in detail about the procedure of BMC transplantation. Informed consent was obtained from 10 patients, who formed the cell therapy group, whereas 10 patients who refused additional cell therapy served as controls. The local ethics committee of the Heinrich-Heine-University, Düsseldorf, approved the study protocol. All procedures conformed to institutional guidelines.

Before taking part in rehabilitation programs, all patients left the hospital with standard medication consisting of acetylsalicylic acid, an ACE inhibitor, a β -blocker, and a statin.

Bone Marrow Aspiration, Isolation, and Cultivation

Seven (± 2) days after acute coronary angiography, bone marrow (~ 40 mL) was aspirated under local anesthesia from ilium of cell therapy patients ($n=10$). Mononuclear BMCs were isolated by Ficoll density separation on Lymphocyte Separation Medium (BioWhittaker) before the erythrocytes were lysed with H_2O . For overnight

cultivation, 1×10^6 BMCs/mL were placed in Teflon bags (Vuelife, Cell Genix) and cultivated in X-Vivo 15 Medium (BioWhittaker) supplemented with 2% heat-inactivated autologous plasma. The next day, BMCs were harvested and washed 3 times with heparinized saline before final resuspension in heparinized saline. Viability was $93 \pm 3\%$. Heparinization and filtration (cell strainer, FALCON) was carried out to prevent cell clotting and microembolization during intracoronary transplantation. The mean number of mononuclear cells harvested after overnight culture was 2.8×10^7 ; this consisted of $0.65 \pm 0.4\%$ AC133-positive cells and $2.1 \pm 0.28\%$ CD34-positive cells. All microbiological tests of the clinically used cell preparations proved negative. As a viability and quality ex vivo control, 1×10^5 cells grown in H5100 medium (Stem Cell Technology) were found to be able to generate mesenchymal cells in culture.

Intracoronary Transplantation of BMCs

Five to nine days after onset of acute infarction, cells were directly transplanted into the infarcted zone (Figure 1). This was accomplished with the use of a balloon catheter, which was placed within the infarct-related artery. After exact positioning of the balloon at the site of the former infarct-vessel occlusion, percutaneous transluminal coronary angioplasty (PTCA) was performed 6 to 7 times for 2 to 4 minutes each. During this time, intracoronary cell transplantation via the balloon catheter was performed, using 6 to 7 fractional high-pressure infusions of 2 to 3 mL cell suspension, each of which contained 1.5 to 4×10^6 mononuclear cells. PTCA thoroughly prevented the backflow of cells and at the same time produced a stop-flow beyond the site of the balloon inflation to facilitate high-pressure infusion of cells into the infarcted zone. Thus, prolonged contact time for cellular migration was allowed.¹⁸

Functional Assessment of Hemodynamics

After 3 months, all 20 patients were followed up by left heart catheterization, left ventriculography, and coronary angiography. Ejection fraction, infarct region, and regional wall movement of the infarcted zone during ejection were determined by left ventriculography. Ejection fraction was measured with Quantcor software (Siemens). To quantify infarction wall movement velocity, 5 axes were placed perpendicular to the long axis in the main akinetic or dyskinetic segment of the ventricular wall. Relative systolic and diastolic lengths were measured, and the mean difference was divided by the systolic duration (in seconds). To quantify the infarct region, the centerline method according to Sheehan was used.¹⁹ All hemodynamic investigations were obtained by two independent observers.

In the cell therapy group before and 3 months after cell transplantation, additional examinations for measuring hemodynamics and myocardial perfusion included dobutamine stress echocardiography, radionuclide ventriculography, catheterization of the right heart, and

TABLE 1. Baseline Characteristics of the Patients

Clinical Data	Cell Therapy	Standard Therapy	P
Characteristics			
No. of patients	10	10	...
Age, y	49±10	50±6	NS
Sex	Male	Male	...
Onset of infarction before angioplasty, h	10±8	13±11	NS
Coronary angiography			
No. of diseased vessels	1.7±0.9	2.1±0.7	NS
No. of patients with LAD/LCX/RCA as the affected vessel	4/1/5	0/2/8	...
No. of patients with stent implantation	9	10	...
Laboratory parameters			
Creatinine kinase, U/L	1138±1170	1308±1187	NS
Creatinine kinase-MB, U/L	106±72	124±92	NS
Bone marrow puncture after angioplasty, d	7±2
Mononuclear bone marrow cells, n (×10 ³)	2.8±2.2

Values are mean±SD or number of patients.

NS indicates not significant; LAD, left anterior descending coronary artery; LCX, left circumflex coronary artery; and RCA, right coronary artery.

stress-redistribution-reinjection ²⁰¹thallium scintigraphy. The contractility index P_{mv}/ESV was calculated by dividing LV systolic pressure (P_{mv}) by end-systolic volume (ESV). Perfusion defect was calculated by scintigraphic bull's-eye technique. Each examination was performed according to standard protocols.

There were no complications or side effects determined in any patient throughout the diagnostic or therapeutic procedure or within the 3-month follow-up period.

Statistical Analysis

All data are presented as mean±SD. Statistical significance was accepted when P was <0.05. Discrete variables were compared as rates, and comparisons were made by χ^2 analysis. Intra-individual comparison of baseline versus follow-up continuous variables was performed with a paired t test. Comparison of nonparametric data between the two groups was performed with Wilcoxon test and Mann-Whitney test. Statistical analysis was performed with SPSS for Windows (version 10.1).

Results

Clinical data between the two groups did not differ significantly. The range of creatinine kinase levels was slightly but not significantly higher in the standard therapy group than it was in the cell therapy group (Table 1).

Comparison of the 2 groups 3 months after cell or standard therapy showed several significant differences in LV dynamics, according to the global and regional analysis of left ventriculogram. The infarct region as a percentage of hypokinetic, akinetic, or dyskinetic segments of the circumference of the left ventricle decreased significantly in the cell therapy group (from 30±13 to 12±7%, $P=0.005$). It was also significantly smaller compared with the standard therapy group after 3 months ($P=0.04$). Within the standard therapy group, only a statistically nonsignificant decrease from 25±8 to 20±11% could be seen. Wall movement velocity over the infarct region rose significantly in the cell therapy group (from 2.0±1.1 to 4.0±2.6 cm/s, $P=0.028$) but not in the standard therapy group (from 1.8±1.3 to 2.3±1.6 cm/s, $P=NS$). No significant difference was observed between the

two groups. Ejection fraction increased in both groups, albeit nonsignificantly (from 57±8 to 62±10% in the cell therapy group and from 60±7 to 64±7% in the standard therapy group) (Table 2).

Further significant improvement could also be seen on additional analysis of the cell therapy group alone. Perfusion defect was considerably decreased by 26% in the cell therapy group (from 174±99 to 128±71 cm², $P=0.016$, assessed by ²⁰¹thallium scintigraphy) (Figure 2). Parallel to the reduction in perfusion defect, improvement (Table 3) could also be seen in:

- (1) Cardiac function, as revealed by increase in stroke volume index (from 49±7 to 56±7 mL/m², $P=0.010$) and ejection fraction (from 51±14 to 53±13%, $P=NS$).
- (2) Cardiac geometry, as shown by decreases in both end-diastolic (from 158±20 to 143±30 mL, $P=NS$) and end-systolic volume (from 82±26 to 67±21 mL, $P=0.011$). Radionuclide ventriculography was used to acquire the data.
- (3) Contractility as evaluated by an increase in the velocity of circumferential fiber shortening (from 20.5±4.2 to 24.4±7.7 mm/s, $P=NS$, assessed by stress echocardiography) and by a marked increase in the ratio of systolic pressure to end-systolic volume (from 1.81±1.44 to 2.27±1.72 mm Hg/mL, $P=0.005$).

Discussion

The present report describes the first clinical trial of intracoronary, autologous, mononuclear BMC transplantation for improving heart function and myocardial perfusion in patients after acute MI. The results demonstrate that transplanted autologous BMCs may lead to repair of infarcted tissue when applied during the immediate postinfarction period. These results also show that the intracoronary approach of BMC transplantation seems to represent a novel

TABLE 2. Comparison of Cell Therapy and Standard Therapy Groups

	Cell Therapy	Standard Therapy	P
No. of patients	10	10	...
Infarct region as functional defect			
Hypokinetic, akinetic, or dyskinetic region at 0 mo, %	30±13	25±8	NS
Hypokinetic, akinetic, or dyskinetic region at 3 mo, %	12±7	20±11	0.04
P	0.005	NS	...
Contractility indices			
Infarction wall movement velocity at 0 mo, cm/s	2.0±1.1	1.8±1.3	NS
Infarction wall movement velocity at 3 mo, cm/s	4.0±2.6	2.3±1.6	NS
P	0.028	NS	...
Hemodynamic data			
LV ejection fraction at 0 mo, %	57±8	60±7	NS
LV ejection fraction at 3 mo, %	62±10	64±7	NS
P	NS	NS	...

NS indicates not significant; 0 mo, zero months, which means the time of infarction; 3 mo, 3 months, which means the time of the follow-up examinations. All data were obtained according to analysis of left ventriculogram.

and effective therapeutic procedure for concentrating and/or depositing infused cells within the region of interest.

Neogenesis of both cardiomyocytes and coronary capillaries with some functional improvement has been shown recently by several investigators using bone marrow-derived cells in experimental infarction.^{11–14,18,20–23} Moreover, trans-endothelial migration from the coronary capillaries and incorporation of cells into heart muscle has been observed experimentally.^{3,12,24–26} Until now, clinical data only existed for the cell therapy of surgically treated chronic ischemic heart disease.^{15,16} Our aim was to transform the encouraging results from animal models to a safe clinical setting. The most crucial questions we had to address while designing and

realizing this trial were: (1) What cell population should we deliver? (2) Which application method is the most efficient? (3) When should the cells be transplanted?

In recent years, several laboratories have shown that environmentally dictated changes of fate (transdetermination) are not restricted to stem cells but may also involve progenitor cells at different steps of a given differentiation pathway (transdifferentiation). Moreover, mesenchymal stem cells may represent an ideal cell source for treating different diseases.²⁷ Adult, mononuclear BMCs contain such stem and progenitor cells (≤1%), eg, mesodermal progenitor cells, hematopoietic progenitor cells, and endothelial progenitor cells. In several animal infarction models it has been shown that: (1) Bone marrow hemangioblasts contribute to the formation of new vessels; (2) bone marrow hematopoietic stem cells differentiate into cardiomyocytes, endothelium,

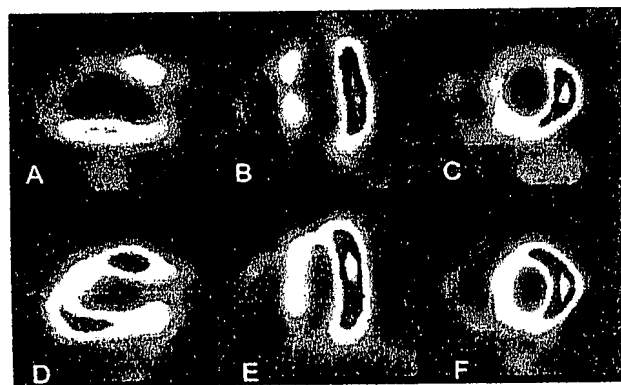


Figure 2. Improved myocardial perfusion of infarcted anterior wall 3 months after intracoronary cell transplantation subsequent to an acute anterior wall infarction detected by ²⁰¹thallium scintigraphy. The images on the left (A, D, sagittal) and in the middle (B, E) show the long axis, whereas those on the right (C, F, frontal) show the short axis of the heart. Initially the anterior wall, with green-colored apical and anterior regions, had reduced myocardial perfusion (A, B, C). Three months after cell transplantation the same anterior wall, now yellow in color, revealed a significant improvement in myocardial perfusion (D, E, F). All illustrations depict the exercise phase.

TABLE 3. Cardiac Function Analysis at 3-Month Follow-Up

	Before Cell Therapy	3 Months After Cell Therapy	P
No. of patients	10	10	...
Hemodynamic data			
LV ejection fraction, %	51±14	53±13	NS
Stroke volume index, mL/m ²	49±7	56±7	0.010
Cardiac geometry			
LV end-diastolic volume, mL	158±20	143±30	NS
LV end-systolic volume, mL	82±26	67±21	0.011
Contractility indices			
Circumferential fiber shortening, mm/s	20.5±4.2	24.4±7.7	NS
P _{sys} /ESV, mm Hg/mL	1.81±1.44	2.27±1.72	0.005
Infarct region as perfusion defect			
²⁰¹ Thallium scintigraphy, cm ²	174±99	128±71	0.016

NS indicates not significant.

and smooth muscle cells⁸⁻¹³; (3) BMCs give rise to mesodermal progenitor cells that differentiate to endothelial cells²⁸; and (4) endothelial progenitors can transdifferentiate into beating cardiomyocytes.²⁹ Thus, several different fractions of mononuclear BMCs may contribute to the regeneration of necrotic myocardium and vessels. In order to utilize this large and perhaps heterogeneous regenerative potential, we decided to use all mononuclear cells from the bone marrow aspirate as a whole, rather than a subpopulation. No further expansion was performed because experimental data have revealed a dramatic decline in the homing capacity of in vitro amplified hematopoietic stem or progenitor cells.³⁰

The second question was how to deliver the cells most efficiently. When given intravenously, only a very small fraction of infused cells can reach the infarct region after the following injection: assuming a normal coronary blood flow of 80 mL/min per 100 g of LV weight, a quantity of 160 mL per left ventricle (assuming a regular LV mass of ≈ 200 g) will flow per minute.^{31,32} This corresponds to only about 3% of cardiac output (assuming a cardiac output of 5000 mL/min).³¹ Therefore, intravenous application would require many circulation passages to enable infused cells to come into contact with the infarct-related artery. Throughout this long circulation and recirculation time, homing of cells to other organs could considerably reduce the numbers of cells dedicated to cell repair in the infarcted zone. Thus, supplying the entire complement of cells by intracoronary administration obviously seems to be advantageous for the tissue repair of infarcted heart muscle and may also be superior to intraventricular injection,³³ because all cells are able to flow through the infarcted and peri-infarcted tissue during the immediate first passage. Accordingly, by this intracoronary procedure the infarct tissue and the peri-infarct zone can be enriched with the maximum available amount of cells at all times.

As stem cells differentiate into more mature types of progenitor cells, it is thought that a special microenvironment in so-called niches regulates cell activity by providing specific combinations of cytokines and by establishing direct cellular contact. For successful long-term engraftment, at least some stem cells have to reach their niches, a process referred to as homing. Mouse experiments have shown that significant numbers of BMCs appear in liver, spleen, and bone marrow after intravenous injection.³⁴ To offer the BMCs the best chance of finding their niche within the myocardium, a selective intracoronary delivery route was chosen. Presumably, therefore, fewer cells were lost by extraction toward organs of secondary interest by this first pass-like effect. To facilitate transendothelial passage and migration into the infarcted zone, cells were infused by high-pressure injection directly into the necrotic area, and the balloon was kept inflated for 2 to 3 minutes; the cells were not washed away immediately under these conditions.

The time point for delivery was chosen as 7 to 8 days after infarction onset for the following reasons:

- (1) In dogs, infarcted territory becomes rich in capillaries and contains enlarged, pericyte-poor "mother vessels" and endothelial bridges 7 days after myocardial ischemia and reperfusion. Twenty-eight days later, a significant muscular vessel wall has already formed.³⁵ Thus, with such timing, cells may be able to reach the worst

damaged parts and at the same time salvage tissue. Transendothelial cell migration may also be enhanced because an adequate muscular coat is not yet formed.

- (2) Until now, only one animal study has attempted to determine the optimum time for cardiomyocyte transplantation to maximize myocardial function after LV injury. Adult rat hearts were cryoinjured and fetal rat cardiomyocytes were transplanted immediately, 2 weeks later, and 4 weeks later. The authors discussed the inflammatory process, which is strongest in the first days after infarction, as being responsible for the negative results after immediate cell transplantation, and they assumed that the best results seen after 2 weeks may have been due to transplantation before scar expansion.³⁶ Until now, however, no systematic experiments have been performed with BMCs to correlate the results of transplantation with the length of such a time delay.
- (3) Another important variable is the inflammatory response in MI, which seems to be a superbly orchestrated interaction of cells, cytokines, growth factors and extracellular matrix proteins mediating myocardial repair. In the first 48 hours, debridement and formation of a fibrin-based provisional matrix predominates before a healing phase ensues.³⁷⁻⁴⁰ Moreover, vascular endothelial growth factor is at its peak concentration 7 days after MI, and the decline of adhesion molecules (intercellular adhesion molecules, vascular cell adhesion molecules) does not take place before days 3 to 4 after MI. We assumed that transplantation of mononuclear BMCs within the "hot" phase of post-MI inflammation might lead them to take part in the inflammation cascade rather than the formation of functional myocardium and vessels.

Taking all of this into account, we can conclude that cell transplantation within the first 5 days after acute infarction is not possible for logistical reasons and is not advisable because of the inflammatory process. On the other hand, transplantation 2 weeks after infarction scar formation seems to reduce the benefit of cell transplantation. Although the ideal time point for transplantation remains to be defined, it is most likely between days 7 and 14 after the onset of MI, as in the present study.

This trial was designed as a phase I safety and feasibility trial, meaning that no control group is necessarily required. However, to validate the results, we correlated them with those obtained from 10 patients who refused to get additional cell therapy and thus received standard therapy alone. We are aware of the fact that such a comparison does not reach the power of a randomly allocated, blinded control group. However, the significant improvement with regard to infarct region, hemodynamics (stroke volume index), cardiac geometry (LV end-systolic volume), and contractility ($P_{1/2}/ESV$ and infarction wall movement velocity) did confirm a positive effect of the additional cell therapy because the changes observed in the standard therapy group failed to reach significance.

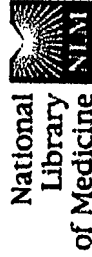
Another important factor for interpreting the results is time interval between onset of symptoms and revascularization of the infarct-related artery by angioplasty; this represents a crucial determinant of LV recovery. For patients with acute MI, it has

been shown that if the time interval is >4 hours, no significant changes in ejection fraction, regional wall motion, or ESV are observed after 6-month follow-up by echocardiography and angiography.⁴¹ None of our 20 patients was treated by angioplasty within 4 hours after onset of symptoms. Our average time interval was 12 ± 10 hours. Thus, PTCA-induced improvement of LV function can be nearly excluded; indeed, the only mild and nonsignificant changes within the standard therapy group are consistent with the above-mentioned data.⁴¹ In contrast, the cell therapy group showed considerable and significant improvement in the same parameters, which may be attributed to BMC-mediated coronary angiogenesis and cardiomyoneogenesis.

These results show that transplantation of autologous BMCs, as well as the intracoronary approach, represent a novel and effective therapeutic procedure for the repair of infarcted myocardium. For this method of therapy, no ethical problems exist, and no side effects were observed at any point of time. The therapeutic benefit for the patient's heart seems to prevail. However, further experimental studies, controlled prospective clinical trials, and variations of cell preparations are required to define the role of this new approach for the therapy of acute MI in humans.

References

- Pfeffer MA, Braunwald E. Ventricular remodeling after myocardial infarction: experimental observations and clinical implications. *Circulation*. 1990;81:1161-1172.
- Ertl G, Gaudron P, Hu K. Ventricular remodeling after myocardial infarction: experimental and clinical studies. *Basic Res Cardiol*. 1993;88:125-137.
- Quaini F, Urbancsek K, Beltrami AP, et al. Chimerism of the transplanted heart. *N Engl J Med*. 2002;346:5-15.
- Leor J, Patterson M, Quinones MJ, et al. Transplantation of fetal myocardial tissue into infarcted myocardium of rat: a potential method for repair of infarcted myocardium? *Circulation*. 1996;94(suppl II):332-336.
- Murry CE, Wiseman RW, Schwartz SM, et al. Skeletal myoblast transplantation for repair of myocardial necrosis. *J Clin Invest*. 1996;98:2512-2523.
- Taylor DA, Atkins BZ, Hungspreugs P, et al. Regenerating functional myocardium: improved performance after skeletal myoblast transplantation. *Nat Med*. 1998;4:929-933.
- Tomita S, Li RK, Weisel RD, et al. Autologous transplantation of bone marrow cells improves damaged heart function. *Circulation*. 1999;100(suppl II):247-256.
- Blau HM, Brazelton TR, Weimann JM. The evolving concept of a stem cell: entity or function? *Cell*. 2001;105:829-841.
- Krause DS, Theise ND, Collector MI, et al. Multi-organ, multi-lineage engraftment by a single bone marrow-derived stem cell. *Cell*. 2001;105:369-377.
- Goodell MA, Jackson KA, Majka SM, et al. Stem cell plasticity in muscle and bone marrow. *Ann NY Acad Sci*. 2001;938:208-218.
- Orlic D, Kajstura J, Chimenti S, et al. Bone marrow cells regenerate infarcted myocardium. *Nature*. 2001;410:701-705.
- Orlic D, Kajstura J, Chimenti S, et al. Mobilized bone marrow cells repair the infarcted heart, improving function and survival. *Proc Natl Acad Sci U S A*. 2001;98:10344-10349.
- Kocher AA, Schuster MD, Szabo MJ, et al. Neovascularization of ischemic myocardium by human bone-marrow-derived angioblasts prevents cardiomyocyte apoptosis, reduces remodeling and improves cardiac function. *Nat Med*. 2001;7:430-436.
- Tomita S, Mickle DA, Weisel RD, et al. Improved heart function with myogenesis and angiogenesis after autologous porcine bone marrow stromal cell transplantation. *J Thorac Cardiovasc Surg*. 2002;123:1132-1135.
- Menasche P, Hagege AA, Scorsin M, et al. Myoblast transplantation for heart failure. *Lancet*. 2001;357:279-280.
- Hamano K, Nishida M, Hirata K, et al. Local implantation of autologous bone marrow cells for therapeutic angiogenesis in patients with ischemic heart disease: clinical trial and preliminary results. *Jpn Circ J*. 2001;65:845-847.
- Strauer BE, Brehm M, Zeus T, et al. Myocardial regeneration after intracoronary transplantation of human autologous stem cells following acute myocardial infarction. *Disch med Wschr*. 2001;126:932-938.
- Wang JS, Shum-Tim D, Chedrawy E, et al. The coronary delivery of marrow stromal cells for myocardial regeneration: pathophysiologic and therapeutic implications. *J Thorac Cardiovasc Surg*. 2001;122:699-705.
- Sheehan FH, Bolson EL, Dodge HT, et al. Advantages and applications of the Centerline method for characterizing regional ventricular function. *Circulation*. 1986;74:293-305.
- Sussman M. Cardiovascular biology: hearts and bones. *Nature*. 2001;410:640-641.
- Toma C, Pittenger MF, Cahill KS, et al. Human mesenchymal stem cells differentiate to a cardiomyocyte phenotype in the adult murine heart. *Circulation*. 2002;105:93-98.
- Kamihata H, Matsubara H, Nishiue T, et al. Implantation of bone marrow mononuclear cells into ischemic myocardium enhances collateral perfusion and regional function via side supply of angioblasts, angiogenic ligands, and cytokines. *Circulation*. 2001;104:1046-1052.
- Ferrari G, Cusella-De Angelis G, Coletta M, et al. Muscle regeneration by bone marrow-derived myogenic progenitors. *Science*. 1998;279:1528-1530.
- Kawamoto A, Gwon HC, Iwaguro H, et al. Therapeutic potential of ex vivo expanded endothelial progenitor cells for myocardial ischemia. *Circulation*. 2001;103:634-637.
- Robinson SW, Cho PW, Levitsky HI, et al. Arterial delivery of genetically labeled skeletal myoblasts to the murine heart: long-term survival and phenotypic modification of implanted myoblasts. *Cell Transplant*. 1996;5:77-91.
- Bittner RE, Schofer C, Weipoltshammer K, et al. Recruitment of bone-marrow-derived cells by skeletal and cardiac muscle in adult dystrophic mdx mice. *Anat Embryol (Berl)*. 1999;199:391-396.
- Jiang Y, Jahagirdar B, Reinhardt RL, et al. Pluripotency of mesenchymal stem cells derived from adult marrow. *Nature*. 2002;20:1-12.
- Reyes M, Lund T, Lenvik T, et al. Purification and ex vivo expansion of postnatal human marrow mesodermal progenitor cells. *Blood*. 2001;98:2615-2625.
- Condorelli G, Borello U, De Angelis L, et al. Cardiomyocytes induce endothelial cells to trans-differentiate into cardiac muscle: implications for myocardium regeneration. *Proc Natl Acad Sci U S A*. 2001;98:10733-10738.
- Szilvassy SJ, Bass MJ, Van Zant G, et al. Organ-selective homing defines engraftment kinetics of murine hematopoietic stem cells and is compromised by ex vivo expansion. *Blood*. 1999;93:1557-1566.
- Gregg DE, Fisher LC. Blood supply to the heart. In: *Handbook of Physiology*. Vol 2. Washington, DC: American Physiological Society; 1963:chap 44, 1517-1584.
- Strauer BE. Myocardial oxygen consumption in chronic heart disease: role of wall stress, hypertrophy and coronary reserve. *Am J Cardiol*. 1979;43:730-740.
- Toma C, Pittenger MF, Byrne BJ, et al. Adult human mesenchymal stem cells differentiate to a striated muscle phenotype following arterial delivery to the murine heart. *Circulation*. 2000;102(suppl II):II-683. Abstract.
- Hendriks PJ, Martens CM, Hagenbeek A, et al. Homing of fluorescently labeled murine hematopoietic stem cells. *Exp Hematol*. 1996;24:129-140.
- Ren G, Michael LH, Entman ML, et al. Morphological characteristics of the microvasculature in healing myocardial infarcts. *J Histochem Cytochem*. 2002;50:71-79.
- Li RK, Mickle DA, Weisel RD, et al. Optimal time for cardiomyocyte transplantation to maximize myocardial function after left ventricular injury. *Ann Thorac Surg*. 2001;72:1957-1963.
- Frangogiannis NG, Smith CW, Entman ML. The inflammatory response in myocardial infarction. *Cardiovasc Res*. 2002;53:31-47.
- Allgöwer M. *The Cellular Basis of Wound Repair*. Springfield, Ill: Charles C. Thomas; 1956.
- Xie Y, Zhou T, Shen W, et al. Soluble cell adhesion molecules in patients with acute coronary syndrome. *Chin Med J*. 2000;113:286-288.
- Soeki T, Tamura Y, Shinohara H, et al. Serial changes in serum VEGF and HGF in patients with acute myocardial infarction. *Cardiology*. 2000;93:168-174.
- Sheiban I, Frangasso G, Rosano GMC, et al. Time course and determinants of left ventricular function recovery after primary angioplasty in patients with acute myocardial infarction. *J Am Coll Cardiol*. 2001;38:464-471.



PubMed Nucleotide Protein Genome Structure PMC Taxonomy OMIM Books

Search PubMed for

☒ Limits ☐ Preview/Index ☐ History ☐ Details

About Entrez

Text Version

Entrez PubMed
Overview
Help | FAQ
Tutorial
New/Noteworthy
E-Utilities

PubMed Services
Journals Database
MeSH Database
Single Citation Matcher
Batch Citation Matcher

Clinical Queries
LinkOut
Cubby

Related Resources
Order Documents
NLM Gateway
TOXNET
Consumer Health
Clinical Alerts
ClinicalTrials.gov
PubMed Central

Privacy Policy

☐ 1: Lancet 2003 Feb 8;361(9356):491-2

[ELSEVIER SCIENCE
FULL-TEXT ARTICLE](#)

Viability and differentiation of autologous skeletal myoblast grafts in ischaemic cardiomyopathy.

Hagege AA, Carrion C, Menasche P, Vilquin JT, Duboc D, Marolleau JP, Desnos M, Bruneval P.

Assistance Publique-Hopitaux de Paris, Department of Cardiology, Hopital European Georges Pompidou and INSERM EMI-16, Necker-Paris V University, Paris, France. hagege@club-internet.fr

Autologous skeletal myoblast transplantation might improve postinfarction ventricular function, but graft viability and differentiation (ie, proof of concept) has not been shown. A 72-year-old man had autologous cultured myoblasts from his vastus lateralis injected to an area of transmural inferior myocardial infarction in non-reperfused scar tissue. He showed improvement in symptoms and left-ventricular ejection fraction. When he died 17.5 months after the procedure, the grafted post-infarction scar showed well developed skeletal myotubes with a preserved contractile apparatus. 65% of myotubes expressed the slow myosin isoform and 33% coexpressed the slow and fast isoforms (vs 44% and 0.6%, respectively, in skeletal muscle). Myoblast grafts can survive and show a switch to slow-twitch fibres, which might allow sustained improvement in cardiac function.

PMID: 12583951 [PubMed - indexed for MEDLINE]

Related Articles, Links

myocardial tissue. These results establish the feasibility of myoblast transplants for myocardial repair in humans.

Publication Types:

- Clinical Trial
- Clinical Trial, Phase I

PMID: 12628737 [PubMed - indexed for MEDLINE]

Display	Abstract	Show:	20	Sort	Send to	Text
---------	----------	-------	----	------	---------	------

Write to the Help Desk
[NCBI](#) | [NLM](#) | [NIH](#)
 Department of Health & Human Services
[Freedom of Information Act](#) | [Disclaimer](#)

May 2 2003 16:34:23

This is Google's cache of http://www.medscape.com/viewarticle/444727_print.

Google's cache is the snapshot that we took of the page as we crawled the web.

The page may have changed since that time. Click here for the current page without highlighting.

To link to or bookmark this page, use the following url: http://www.google.com/search?q=cache:cxRoFu2EUxYC:www.medscape.com/viewarticle/444727_print+regenerate+dead+heart+tissue+after+myocardial+infarction&hl=en&ie=UTF-8

Google is not affiliated with the authors of this page nor responsible for its content.

These search terms have been

regenerate dead heart tissue after myocardial infarction



www.medscape.com

To Print: Click your browser's PRINT button.

NOTE: To view the article with Web enhancements, go to:

<http://www.medscape.com/viewarticle/444727>



Autologous Cell Transplant Helpful in Ischemic Heart or Legs

Laurie Barclay, MD

Medscape Medical News 2002. © 2002 Medscape

Nov. 18, 2002 — Autologous cell transplantation may benefit ischemic hearts and legs, according to three presentations on Nov. 18 at the American Heart Association's 75th Scientific Sessions held in Chicago, Illinois. Two studies focused on injecting autologous bone marrow cells or autologous skeletal myoblasts into the scarred area of an infarcted heart. In another study, injecting autologous bone marrow into ischemic limbs led to new vessel growth, reducing the need for amputation.

"Bone marrow not only can differentiate into heart cells, but also smooth muscle cells, connective tissue cells and other types of cells to reconstitute the entire structure of a tissue," presenter Manuel Galinanes, MD, from the University of Leicester in the U.K., says in a news release. "The benefit [of transplanting bone marrow into scar tissue of the heart] could be seen only six weeks after injection."

In 14 patients with low ejection fraction post-myocardial infarction (MI), autologous bone marrow from the sternum was injected into scarred myocardium during nonemergency coronary artery bypass surgery. Heart wall motion measured with echocardiography improved within weeks of treatment, and improvements persisted for at least 10 months after treatment.

The regional wall motion score decreased significantly, reflecting less movement abnormality, from a mean score of 2.41 at baseline to 2.16 six weeks after treatment and 2.09 ten months after treatment. The global wall motion score also decreased significantly from 1.96 before surgery to 1.64 at six weeks, and stabilized at 1.65 after 10 months.

Although it is still unproven that bone marrow creates a new cellular infrastructure in heart scar tissue, "that is the only possible explanation," Galinanes says. "The ability to confirm the presence of scar tissue with dobutamine stress echo before surgery, and then confirm it again during surgery, told us that the affected area was dysfunctional and the abnormality was irreversible. We wanted to make sure that we were injecting the marrow into dead tissue to help ensure that the injection would not pose any serious risk to the patient."

If additional studies confirm safety and efficacy, Galinanes says that this treatment would be a welcome addition to the post-MI arsenal, which also includes gene therapy, growth factor therapy, and laser treatments.

In a multicenter trial supervised by the U.S. Food and Drug Administration, investigators safely transplanted 16 patients with autologous skeletal myoblasts injected into hearts severely damaged by MI or heart failure. Baseline left-ventricular ejection fraction was less than 30%. Eleven patients were undergoing coronary artery bypass surgery and five were having implantation of a left ventricular assist device. Myoblasts extracted from thigh muscle were grown in large quantities in vitro using a controlled cell expansion manufacturing process, and were injected in doses ranging from 10 million to 300 million cells.

"We have been able to regenerate dead heart muscle, or scar tissue, in the area of heart attack without increasing risk of death. Our findings will allow us to move forward with testing if the procedure can improve the contractility of the heart," says lead author Nabil Dib, MD, from the Arizona Heart Institute in Phoenix. "We found that the transplanted myoblasts survived and thrived in patients. Areas damaged by heart attack and cardiovascular disease showed evidence of repair and viability."

Twelve weeks after transplant, mean ejection fraction rates improved from 22.7% to 35.8%, or a 58% increase. Echocardiogram, magnetic resonance imaging, and positron emission tomography showed evidence of regeneration in the area of the graft. There were no significant adverse events related to the cell transplant procedure at nine-month follow-up.

The third study showed that bone marrow cells implanted into ischemic legs in patients with peripheral arterial disease (PAD) formed new blood vessels, increased blood flow, and prevented amputation.

"This is the first multicenter and double-blind clinical study to prove the clinical efficacy of growing new blood vessels (angiogenesis) using bone marrow cell transplantation," says lead author Hiroya Masaki, MD, PhD, from Kansai Medical University in Osaka, Japan.

In this randomized trial, 45 patients with PAD received injections of autologous bone marrow mononuclear cells into the calf muscles. Compared with controls who received saline injections, patients who received bone marrow mononuclear cell transplants had a "striking" increase in new capillary formation and in newly visible collateral vessels.

Of 45 treated patients, 31 had an increase in ankle-brachial pressure index in the treated limbs, and 39 had decreased rest pain with improved treadmill endurance. Ischemic ulcers or gangrene healed in 21 of 28 treated limbs.

CD34-cells, which can develop into endothelial progenitor cells, expressed angiogenic growth factors including basic fibroblast growth factor, vascular endothelial growth factor, and angiopoietin-1. Although more research is needed to determine long-term efficacy and safety, "this new angiogenesis therapy using bone marrow cell transplantation may help many patients suffering with ischemic limbs," Masaki says.

AHA 75th Scientific Sessions: Abstracts 111623, 101758, 109801. Presented Nov. 18, 2002.

Reviewed by Gary D. Vogin, MD



PubMed

Nucleotide

Protein

Genome

Structure

PMC

Taxonomy

OMIM

Books

Search PubMed for

☒ Limits

Preview/Index

History

Clipboard

Details

About Entrez

Text Version

Display Abstract

Go

Clear

Send to

Text

1: J Am Coll Cardiol 2003 Mar 5;41(5):879-88

ELSEVIER SCIENCE
FULL-TEXT ARTICLE

Entrez PubMed
Overview
Help | FAQ
Tutorial
New/Noteworthy
E-Utilities

PubMed Services
Journals Database
MeSH Database
Single Citation Matcher
Batch Citation Matcher
Clinical Queries
LinkOut
Cubby

Related Resources
Order Documents
NLM Gateway
TOXNET
Consumer Health
Clinical Alerts
ClinicalTrials.gov
PubMed Central

Privacy Policy

Related Articles, Links

Autologous skeletal myoblasts transplanted to ischemia-damaged myocardium in humans. Histological analysis of cell survival and differentiation.

Pagani FD, DerSimonian H, Zawadzka A, Wetzel K, Edge AS, Jacoby DB, Dinsmore JH, Wright S, Aretz TH, Eisen HJ, Aaronson KD.

Section of Cardiac Surgery, University of Michigan, Ann Arbor, MI 48109, USA. fpagani@umich.edu

OBJECTIVES: We report histological analysis of hearts from patients with end-stage heart disease who were transplanted with autologous skeletal myoblasts concurrent with left ventricular assist device (LVAD) implantation. **BACKGROUND:** Autologous skeletal myoblast transplantation is under investigation as a means to repair infarcted myocardium. To date, there is only indirect evidence to suggest survival of skeletal muscle in humans. **METHODS:** Five patients (all male; median age 60 years) with ischemic cardiomyopathy, refractory heart failure, and listed for heart transplantation underwent muscle biopsy from the quadriceps muscle. The muscle specimen was shipped to a cell isolation facility where myoblasts were isolated and grown. Patients received a transplant of 300 million cells concomitant with LVAD implantation. Four patients underwent LVAD explant after 68, 91, 141, and 191 days of LVAD support (three transplant, one LVAD death), respectively. One patient remains alive on LVAD support awaiting heart transplantation. **RESULTS:** Skeletal muscle cell survival and differentiation into mature myofibers were directly demonstrated in scarred myocardium from three of the four explanted hearts using an antibody against skeletal muscle-specific myosin heavy chain. An increase in small vessel formation was observed in one of three patients at the site of surviving myotubes, but not in adjacent tissue devoid of engrafted cells. **CONCLUSIONS:** These findings represent demonstration of autologous myoblast cell survival in human heart. The implanted skeletal myoblasts formed viable grafts in heavily scarred human

EVIDENCE APPENDIX

Item 15

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

APPLICANT: James P. Elia)	
)	
SERIAL NO.: 09/836,750)	EXAMINER: E.C. Kemmerer, Ph.D.
)	
FILED: April 17, 2001)	
)	GROUP ART UNIT: 1646
FOR: METHOD FOR GROWING)	
MUSCLE IN A HUMAN HEART)	

SUPPLEMENTAL DECLARATION OF RICHARD HEUSER, M.D.

I, Richard Heuser, declare as follows:

1. I have offices at 500 West Thomas Road, Suite 900, Phoenix, Arizona 85013.
2. This Supplemental Declaration is submitted in addition to my previously submitted Declaration in this application, dated June 5, 2003, and makes no changes to such previous Declaration.
3. My Curriculum Vitae is attached as Exhibit A to my previous Declaration.
4. I have read and understood the disclosures of the above-referenced patent application at page 20, line 10 through page 21, line 15; and page 44, line 19 through page 46, line 16. Such disclosures are the same as read and understood by me in my previous Declaration. A copy of such disclosures is attached hereto as Supplemental Exhibit A.

5. I note that the disclosures referenced in above Paragraph 4 relate to using a growth factor for promoting the growth of soft tissue and, more specifically, to a method which may use such growth factors for growing a new portion of a human heart by growing new muscle in the heart.
6. I have read and understood the claims set forth in Supplemental Exhibit B and have been informed that such claims will be presented to the Patent and Trademark Office in the near future.
7. Based upon above Paragraphs 4-6 and Paragraph 7 of my previous Declaration, it is my opinion that introducing a growth factor into a human patient will predictably cause new muscle growth in the heart of the patient.
8. Based upon above Paragraphs 4-6, it is my opinion that one skilled in the medical arts, armed with the knowledge in such paragraphs, would be able to practice the method set forth in Supplemental Exhibit B without need for resorting to undue experimentation. I have been informed that the Examiner has questioned the fact that dosages are not recited in the specification of the above-identified application in connection with the administration of cell growth factors to a human patient with use of intravenous or intraluminal techniques. Such techniques are the subject of claims 248-249 in above-mentioned Supplemental Exhibit B. In my opinion, dosages of cellular growth factors to achieve the above-mentioned heart muscle growth are a matter of routine medical practice, requiring only a reasonable degree of experimentation, depending upon such factors as extent of prior heart condition, size of patient, age of patient, health of patient, etc. Consequently, it is my opinion that the disclosure mentioned in Supplemental Exhibit A would enable a person skilled in the medical arts to practice the invention of claims 248-249 and predictably anticipate the results defined therein without need for resorting to undue experimentation.

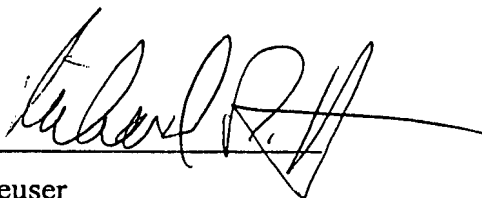
9. Declarant states that the above opinion was reached independently.

Declarant understands that (1) any willful false statements and the like made herein are punishable by fine or imprisonment, or both (18 U.S.C. 1001) and may jeopardize the validity of the application or any patent issuing thereon, and (2) that all statements made of Declarant's own knowledge are true and that all statements made on information and belief are believed to be true.

Further Declarant sayeth not.

Date:

2/4/04



Richard Heuser

SUPPLEMENTAL EXHIBIT A
DISCLOSURES
APPLICATION SERIAL NO. 09/836,750

PAGE 20, LINE 10 – PAGE 21, LINE 15

Growth factors can be utilized to induce the growth of “hard tissue” or bone and “soft tissues” like ectodermal and mesodermal tissues. As used herein, the term growth factor encompasses compositions and living organisms which promote the growth of hard tissue, such as bone, or soft tissue, in the body of a patient. The compositions include organic and inorganic matter. The compositions can be genetically produced or manipulated. The living organisms can be bacteria, viruses, or any other living organism which promote tissue growth. By way of example and not limitation, growth factors can include platelet-derived growth factor (PDGF), epidermal growth factor (EGF), fibroblast growth factor (acidic/basic (FGF a,b), interleukins (IL's), tumor necrosis factor (TNF), transforming growth factor (TGF-B), colony-stimulating factor (CSF), osteopontin (Eta-1 OPN), platelet-derived growth factor (PDGF), interferon (INF), bone morphogenic protein 1 (BMP-1), and insulin growth factor (IGF). Recombinant and non-recombinant growth factors can be utilized as desired. Bacteria or viruses can, when appropriate, be utilized as growth factors. For example, there is a bacterial hydrophilic polypeptide that self-assembles into a nanometer internal diameter pore to build a selective lipid body. Various enzymes can be utilized for the synthesis of peptides which contain amino acids that control three-dimensional protein structure and growth. Growth factors can be applied in gels or other carriers which regulate the rate of release of the growth factors and help maintain the growth factors and the carrier, at a desired location in the body. Time release capsules, granules, or other carriers containing growth factor can be activated by tissue pH, by enzymes, by ultrasound,

by electricity, by heat, by selected *in vivo* chemicals or by any other selected means to release the growth factor. The carrier can be resorbable or non-resorbable. Or, the growth factor itself can be activated by similar means. Either the carrier or the growth factor can mimic extracellular fluid to control cell growth, migration, and function. The growth factor can be administered orally, systemically, in a carrier, by hypodermic needle, through the respiratory tract, or by any other desired method. The growth factor can also be administered into a capsule or other man-made composition or structure placed in the body. While administration of the growth factor is presently usually localized in the patient's body, circumstances may arise where it is advantageous to distribute a growth factor throughout the patient's body in uniform or non-uniform concentrations. An advantage to growth factors is that they can often, especially when in capsule form or in some other containment system, be inserted to a desired site in the body by simply making a small incision and inserting the growth factor. The making of such small incision comprises minor surgery which can often be accomplished on an out-patient basis. The growth factors can be multifactorial and nonspecific.

PAGE 44, LINE 19 – PAGE 46, LINE 16

Genetic material comprising a portion of a gene, a gene, genes, a gene product (i.e., a composition a gene causes to be produced like, for example, an organ-producing growth factor), growth factor, or an ECM (extracellular matrix) can be used in or on the body to grow an organ to tissue. For example, the vascular epithelial growth factor gene (VEGF) or its growth factor equivalent can be inserted into the body to cause an artery to grow. When insertion of a gene, portion of a gene, gene product, growth factor, or ECM *in vivo* or *ex vivo* is referred to herein in connection with any of the implant techniques of the invention, it is understood that a cell

nutrient culture(s), physiological nutrient culture(s), carrier (s), enhancer(s), promoter(s), or any other desired auxiliary component(s) can be inserted with the gene or at the same location as the gene, growth factor, ECM, etc.

An artery is an organ from the circulatory system. An artery can be grown in the heart, legs, or other areas by injecting a gene or other genetic material into muscle at a desired site. Size, vascularity, simplicity of access, ease of exploitation, and any other desired factors can be utilized in selecting a desired site. The gene is one of several known VEGF genes which cause the production of vascular endothelial growth factors. Several VEGF genes which produce vascular endothelial growth factors are believed to exist because nature intends for there to be several pathways (i.e., genes) which enable the production of necessary growth factors. The existence of several pathways is believed important because if one of the genes is damaged or inoperative, other similar genes can still orchestrate the production of necessary growth factors. VEGF genes are used by the body to promote blood vessel growth. VEGF genes are assimilated (taken in) by muscle cells. The genes cause the muscle cells to make a VEGF protein which promotes the growth of new arteries. VEGF proteins can be made in a lab and injected into a patient intravenously, intraluminally, or intramuscularly to promote the growth of an artery. Or, the genes (or other genetic material) can be applied with an angioplasty balloon, with the assistance of a vector, or by any other method.

It is not always desirable to grow a completely new organ. Sometimes growing a portion of an organ is desirable. For example, in some heart attacks or strokes, a portion of the heart or brain remains viable and a portion dies. An injection of a gene to form cardiac muscle and/or an injection of a gene to form an artery can be utilized to revive or replace the dead portion of the heart. The dead portion of the heart may (or may not) be used as a matrix while the new muscles

and vessels grow. Thus, in this example, a partial new organ is grown in a pre-existing organ. A pacemaker may (or may not) be necessary. A second injection of a gene may (or may not) be necessary to stop cardiac muscle growth once it is completed. Portions of organs throughout the body can similarly be repaired or replaced. It may be necessary to provide gene(s) or growth factor(s) sequentially. For instance, one or more blood vessels are grown by inserting an appropriate gene or other genetic material into a selected area. Second, an appropriate gene or other genetic material is inserted in the selected area to grow a bone or other organ.

The size and shape limitation of the desired structure can come from a containment and boundary contact inhibition phenomenon or by a chemical inhibition.

A variation on the theme of growing a portion of an organ is as follows: a portion of a heart dies. The pericardium is utilized as a scaffold and seeded with cells and/or genes to grow new muscle, and genes (or other genetic material) to grow new arteries. Immediately adjacent the dead cardiac muscle, onto or into the pericardium, the appropriate cells, genes, and/or growth factors (or other genetic material) are placed. Once the new muscle and blood vessels have grown, the function specific tissue can be applied to the damaged portion of the heart and paced, if necessary, to augment cardiac action. If the surgeon desires, the dead muscle can be removed and the new muscle and blood vessels can be surgically rotated into the excised region and secured. This probably can be done endoscopically. In essence, the pericardium is utilized to allow the new muscle wall to grow. The new muscle wall is then transplanted into the damaged heart wall. This procedure utilizes the body as a factor to grow an organ and/or tissue, after which the organ and/or tissue is transplanted to a desired region. On the other hand, the new muscle wall may integrate itself into the old wall and not require transplantation.

SUPPLEMENTAL EXHIBIT B
CLAIMS
APPLICATION SERIAL NO. 09/836,750

236. A method of growing a new portion of a pre-existing heart comprising the steps of placing a growth factor in a body of a human patient and growing new muscle and growing a new artery in said heart.
238. The method of claim 236, further comprising repairing a dead portion of said heart.
239. The method of claim 236, further comprising repairing a damaged portion of said heart.
240. The method of claim 236, wherein said growth factor comprises genetic material selected from the group consisting of a portion of a gene, a gene, a gene product, and an extracellular matrix.
241. The method of claim 240, wherein said genetic material comprises a gene.
242. The method of claim 241, wherein said gene comprises VEGF.
243. The method of claim 236, wherein said growth factor comprises a member selected from the group consisting of cells, cellular products, and derivatives of cellular products.
244. The method of claim 243, wherein said growth factor comprises a cell
245. The method of claim 244, wherein said cell is multifactorial and non-specific.
246. The method of claim 245, wherein said cell comprises a stem cell.

- 247. The method of claim 236, wherein said growth factor is placed in said patient by injection.
- 248. The method of claim 247, wherein said injection is intravenous.
- 249. The method of claim 247, wherein said injection is intraluminal.
- 250. The method of claim 247, wherein said injection is intramuscular.
- 251. The method of claim 236, wherein said growth factor is placed in said patient by a carrier.
- 252. The method of claim 251, wherein said carrier comprises an angioplasty balloon.
- 253. The method of claim 236, wherein said growth factor comprises a gene and a cell.

EVIDENCE APPENDIX

Item 16

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

APPLICANT: James P. Elia)	
)	
SERIAL NO.: 09/836,750)	EXAMINER: E.C. Kemmerer, Ph.D.
)	
FILED: April 17, 2001)	
)	
FOR: METHOD FOR GROWING)	GROUP ART UNIT: 1646
MUSCLE IN A HUMAN HEART)	

**SECOND SUPPLEMENTAL DECLARATION
OF RICHARD HEUSER, M.D., F.A.C.C., F.A.C.P.**

I, Richard Heuser, declare as follows:

1. I have offices at 500 West Thomas Road, Suite 900, Phoenix, Arizona 85013.
2. This Second Supplemental Declaration is submitted in addition to my previous Declaration, dated June 5, 2003 and my Supplemental Declaration dated February 4, 2004. No changes are made to either of such previous Declarations.
3. My Curriculum Vitae (hereinafter "CV") is attached as Exhibit A to my Declaration of June 5, 2003.
4. It is my understanding that the Examiner in charge of the above-identified patent

application, in an Office Action dated June 1, 2004 for related patent application Serial No. 09/794,456, questioned my qualification, for the first time, to render my previous opinions mentioned in above Paragraph 2. It is my further understanding that the basis for such questioning was that the Examiner noted that I did not report experience with cellular therapy. I desire to provide the information contained in following paragraph 5 so that the Examiner can consider such information in this application, as well.

5. I am currently Director of Cardiovascular Research at St. Joseph's Hospital and Medicine Center, and I serve as Clinical Professor of Medicine at University of Arizona College of Medicine. Over the past six years, I have worked in gene therapy, as well as muscle regeneration for the treatment of cardiomyopathy.

In my CV, you will note reference to work that was done with Sulzer Medical involving a rabbit hind limb model to stimulate peripheral vascular disease. I injected a growth mixture that included FGF, etc. into the hind limb model.

In my U.S. Patent No. 6,190,379 entitled "Hot Tip Catheter," I developed a technique to deliver radiofrequency (PMR). In the full embodiment of the patent, I discuss delivery of protein and/or muscle cells in the myocardium using the inventive technique.

I have been involved as a member of the scientific advisory board with the world leader in cardiomyocyte regeneration, Bioheart, Miami Lakes, Florida. This company has been involved with laboratory and clinical trials using skeletal muscle cultured and modified. The sample is then delivered into the myocardium via a surgical or catheter approach.

6. I have read and understood the disclosures of the above-referenced patent application at page 20, line 10 through page 21, line 17; and page 44, line 19 through page 46, line 16. Such disclosures are the same as I read and understood in my previous Declaration and

Supplemental Declaration. A copy of such disclosures is attached hereto as Second Supplemental Declaration Exhibit A.

7. I note that the disclosures referenced in above Paragraph 6 relate to using a growth factor for promoting the growth of soft tissue and, more specifically, to a method which may use such growth factors for growing a new portion of a human heart by growing new cardiac muscle in the heart.
8. I have read and understood the claims set forth in Second Supplemental Declaration Exhibit B and have been informed that such claims are currently presented in this application.
9. Based upon above Paragraphs 6-8, it is my opinion that one skilled in the medical arts, armed with the knowledge in such paragraphs, would be enabled to practice the method set forth in Second Supplemental Declaration Exhibit B and to predictably anticipate the results defined therein without need for resorting to undue experimentation.
10. I believe that one skilled in the medical arts, upon reading the disclosures in above , such as multifactorial and non-specific cells, Paragraph 6, would understand that cellular growth factors are included in such disclosures. Moreover, such skilled person would understand the disclosure on page 45 to be authored as an illustration of various modes of delivery of growth factors, whether they are genes or other genetic material; and that such skilled person would further understand that the disclosures on pages 45 and 46 describe genetic material to include appropriate cells and genes.

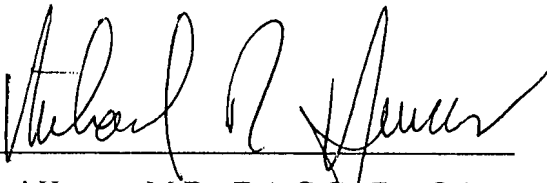
11. Declarant states that the above opinion was reached independently.

Declarant understands that (1) any willful false statements and the like made herein are punishable by fine or imprisonment, or both (18 U.S.C. 1001) and may jeopardize the validity of the application or any patent issuing thereon, and (2) that all statements made of Declarant's own knowledge are true and that all statements made on information and belief are believed to be true.

Further Declarant sayeth not.

Date:

7/18/04


Richard Heuser, M.D., F.A.C.C., F.A.C.P.

SECOND SUPPLEMENTAL DECLARATION

EXHIBIT A

DISCLOSURES

**SECOND SUPPLEMENTAL DECLARATION
EXHIBIT A**

**DISCLOSURES
APPLICATION SERIAL NO. 09/836,750**

PAGE 20, LINE 10 – PAGE 21, LINE 15

Growth factors can be utilized to induce the growth of “hard tissue” or bone and “soft tissues” like ectodermal and mesodermal tissues. As used herein, the term growth factor encompasses compositions and living organisms which promote the growth of hard tissue, such as bone, or soft tissue, in the body of a patient. The compositions include organic and inorganic matter. The compositions can be genetically produced or manipulated. The living organisms can be bacteria, viruses, or any other living organism which promote tissue growth. By way of example and not limitation, growth factors can include platelet-derived growth factor (PDGF), epidermal growth factor (EGF), fibroblast growth factor (acidic/basic (FGF a,b), interleukins (IL’s), tumor necrosis factor (TNF), transforming growth factor (TGF-B), colony-stimulating factor (CSF), osteopontin (Eta-1 OPN), platelet-derived growth factor (PDGF), interferon (INF), bone morphogenic protein 1 (BMP-1), and insulin growth factor (IGF). Recombinant and non-recombinant growth factors can be utilized as desired. Bacteria or viruses can, when appropriate, be utilized as growth factors. For example, there is a bacterial hydrophilic polypeptide that self-assembles into a nanometer internal diameter pore to build a selective lipid body. Various enzymes can be utilized for the synthesis of peptides which contain amino acids that control three-dimensional protein structure and growth. Growth factors can be applied in gels or other carriers which regulate the rate of release of the growth factors and help maintain the growth factors and the carrier, at a desired location in the body. Time release capsules, granules, or

other carriers containing growth factor can be activated by tissue pH, by enzymes, by ultrasound, by electricity, by heat, by selected *in vivo* chemicals or by any other selected means to release the growth factor. The carrier can be resorbable or non-resorbable. Or, the growth factor itself can be activated by similar means. Either the carrier or the growth factor can mimic extracellular fluid to control cell growth, migration, and function. The growth factor can be administered orally, systemically, in a carrier, by hypodermic needle, through the respiratory tract, or by any other desired method. The growth factor can also be administered into a capsule or other man-made composition or structure placed in the body. While administration of the growth factor is presently usually localized in the patient's body, circumstances may arise where it is advantageous to distribute a growth factor throughout the patient's body in uniform or non-uniform concentrations. An advantage to growth factors is that they can often, especially when in capsule form or in some other containment system, be inserted to a desired site in the body by simply making a small incision and inserting the growth factor. The making of such small incision comprises minor surgery which can often be accomplished on an out-patient basis. The growth factors can be multifactorial and nonspecific.

PAGE 44, LINE 19 – PAGE 46, LINE 16

Genetic material comprising a portion of a gene, a gene, genes, a gene product (i.e., a composition a gene causes to be produced like, for example, an organ-producing growth factor), growth factor, or an ECM (extracellular matrix) can be used in or on the body to grow an organ to tissue. For example, the vascular epithelial growth factor gene (VEGF) or its growth factor equivalent can be inserted into the body to cause an artery to grow. When insertion of a gene, portion of a gene, gene product, growth factor, or ECM *in vivo* or *ex vivo* is referred to herein in

connection with any of the implant techniques of the invention, it is understood that a cell nutrient culture(s), physiological nutrient culture(s), carrier (s), enhancer(s), promoter(s), or any other desired auxiliary component(s) can be inserted with the gene or at the same location as the gene, growth factor, ECM, etc.

An artery is an organ from the circulatory system. An artery can be grown in the heart, legs, or other areas by injecting a gene or other genetic material into muscle at a desired site. Size, vascularity, simplicity of access, ease of exploitation, and any other desired factors can be utilized in selecting a desired site. The gene is one of several known VEGF genes which cause the production of vascular endothelial growth factors. Several VEGF genes which produce vascular endothelial growth factors are believed to exist because nature intends for there to be several pathways (i.e., genes) which enable the production of necessary growth factors. The existence of several pathways is believed important because if one of the genes is damaged or inoperative, other similar genes can still orchestrate the production of necessary growth factors. VEGF genes are used by the body to promote blood vessel growth. VEGF genes are assimilated (taken in) by muscle cells. The genes cause the muscle cells to make a VEGF protein which promotes the growth of new arteries. VEGF proteins can be made in a lab and injected into a patient intravenously, intraluminally, or intramuscularly to promote the growth of an artery. Or, the genes (or other genetic material) can be applied with an angioplasty balloon, with the assistance of a vector, or by any other method.

It is not always desirable to grow a completely new organ. Sometimes growing a portion of an organ is desirable. For example, in some heart attacks or strokes, a portion of the heart or brain remains viable and a portion dies. An injection of a gene to form cardiac muscle and/or an injection of a gene to form an artery can be utilized to revive or replace the dead portion of the

heart. The dead portion of the heart may (or may not) be used as a matrix while the new muscles and vessels grow. Thus, in this example, a partial new organ is grown in a pre-existing organ. A pacemaker may (or may not) be necessary. A second injection of a gene may (or may not) be necessary to stop cardiac muscle growth once it is completed. Portions of organs throughout the body can similarly be repaired or replaced. It may be necessary to provide gene(s) or growth factor(s) sequentially. For instance, one or more blood vessels are grown by inserting an appropriate gene or other genetic material into a selected area. Second, an appropriate gene or other genetic material is inserted in the selected area to grow a bone or other organ.

The size and shape limitation of the desired structure can come from a containment and boundary contact inhibition phenomenon or by a chemical inhibition.

A variation on the theme of growing a portion of an organ is as follows: a portion of a heart dies. The pericardium is utilized as a scaffold and seeded with cells and/or genes to grow new muscle, and genes (or other genetic material) to grow new arteries. Immediately adjacent the dead cardiac muscle, onto or into the pericardium, the appropriate cells, genes, and/or growth factors (or other genetic material) are placed. Once the new muscle and blood vessels have grown, the function specific tissue can be applied to the damaged portion of the heart and paced, if necessary, to augment cardiac action. If the surgeon desires, the dead muscle can be removed and the new muscle and blood vessels can be surgically rotated into the excised region and secured. This probably can be done endoscopically. In essence, the pericardium is utilized to allow the new muscle wall to grow. The new muscle wall is then transplanted into the damaged heart wall. This procedure utilizes the body as a factor to grow an organ and/or tissue, after which the organ and/or tissue is transplanted to a desired region. On the other hand, the new muscle wall may integrate itself into the old wall and not require transplantation.

SECOND SUPPLEMENTAL DECLARATION

EXHIBIT B

CLAIMS

**SECOND SUPPLEMENTAL DECLARATION
EXHIBIT B**

**CLAIMS
APPLICATION SERIAL NO. 09/836,750**

236. A method of growing a new portion of a pre-existing heart comprising the steps of placing a growth factor in a body of a human patient and growing new cardiac muscle and growing a new artery in said heart.
238. The method of claim 236, further comprising repairing a dead portion of said heart.
239. The method of claim 236, further comprising repairing a damaged portion of said heart.
240. The method of claim 236, wherein said growth factor comprises genetic material selected from the group consisting of a portion of a gene, a gene, a gene product, and an extracellular matrix.
241. The method of claim 240, wherein said genetic material comprises a gene.
242. The method of claim 241, wherein said gene comprises VEGF.
243. The method of claim 236, wherein said growth factor comprises a member selected from the group consisting of cells, cellular products, and derivatives of cellular products.
244. The method of claim 243, wherein said growth factor comprises a cell
245. The method of claim 244, wherein said cell is multifactorial and non-specific.
246. The method of claim 245, wherein said cell comprises a stem cell.

- 247. The method of claim 236, wherein said growth factor is placed in said patient by injection.
- 248. The method of claim 247, wherein said injection is intravenous.
- 249. The method of claim 247, wherein said injection is intraluminal.
- 250. The method of claim 247, wherein said injection is intramuscular.
- 251. The method of claim 236, wherein said growth factor is placed in said patient by a carrier.
- 252. The method of claim 251, wherein said carrier comprises an angioplasty balloon.
- 253. The method of claim 236, wherein said growth factor comprises a gene and a cell.
- 254. A method of growing a new portion of a pre-existing organ comprising placing a growth factor in a body of a patient to grow new muscle in said organ.
- 255. The method of claim 254, wherein said organ comprises a heart.
- 256. The method of claim 255, wherein said new muscle comprises cardiac muscle and said growth factor comprises a stem cell.

EVIDENCE APPENDIX

Item 17

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

APPLICANT: James P. Elia)	
)	
SERIAL NO.: 09/836,750)	EXAMINER: E.C. Kemmerer, Ph.D.
)	
FILED: April 17, 2001)	
)	GROUP ART UNIT: 1646
FOR: METHOD FOR GROWING)	
MUSCLE IN A HUMAN HEART)	

**THIRD SUPPLEMENTAL DECLARATION
OF RICHARD HEUSER, M.D., F.A.C.C., F.A.C.P.**

I, Richard Heuser, declare as follows:

1. I have offices at 500 West Thomas Road, Suite 900, Phoenix, Arizona 85013.
2. This Third Supplemental Declaration is submitted in addition to my previous Declaration, dated June 5, 2003; my Supplemental Declaration dated February 4, 2004; and my Second Supplemental Declaration dated July 18, 2004. No changes are made to any of such previous Declarations.
3. My Curriculum Vitae is attached as Exhibit A to my Declaration of June 5, 2003.
4. It is my understanding that the Examiner in charge of the above-identified patent application is also in the Examiner in charge of co-pending patent application Serial No. 09/794,456. In an Advisory Action dated November 26, 2004, for aforesaid Serial No. 09/794456, the Examiner further questioned my qualification to render my opinions in the three previous Declarations mentioned in above Paragraph 2. It is my further

understanding that the Examiner reviewed my U.S. Patent No. 6,190,379 and did not find mention of delivery of any substance to the myocardium nor the word "cell." Also, the Examiner questioned my role in the cell delivery portion of Bioheart's laboratory and clinical trials using skeletal muscle cultured and modified. I provide the following information to respond to the Examiner's newly raised questions.

5. Regarding, U.S. Patent No. 6,190,379, the following is stated in my Second Supplemental Declaration:

In my U.S. Patent No. 6,190,379 entitled "Hot Tip Catheter," I developed a technique to deliver radiofrequency (PMR). In the full embodiment of the patent, I discuss delivery of protein and/or muscle cells in the myocardium using the inventive technique.

By the above statement, I meant that the device shown in the patent has been used for the delivery of protein and/or muscle cells to the myocardium. At a presentation at the Angiogenesis Meeting in 1999 in Washington, D.C., we described this use of growth factors in a pig model with the development of neo vascularization. Moreover, I have had discussions with Bioheart regarding the use of my U.S. Patent No. 6,190,379 for delivery of cells.

Regarding my work at Bioheart, the following is stated in my Second Supplemental Declaration:

I have been involved as a member of the scientific advisory board with the world leader in cardiomyocyte regeneration, Bioheart, Miami Lakes, Florida. This company has been involved with laboratory and clinical trials using skeletal muscle cultured and modified. The sample is then delivered into the myocardium via a surgical or catheter approach.

To provide further information regarding the Examiner's questioning my involvement with Bioheart, I am a Scientific Advisory Board Member and in such role advise Bioheart throughout its pre-clinical and clinical work involving the delivery of skeletal muscle

cells into the myocardium. I am also an investigator with Bioheart's Phase 3 clinical trials in the United States. Such trials have not yet commenced.


6. Declarant states that the above opinion was reached independently.

Declarant understands that (1) any willful false statements and the like made herein are punishable by fine or imprisonment, or both (18 U.S.C. 1001) and may jeopardize the validity of the application or any patent issuing thereon, and (2) that all statements made of Declarant's own knowledge are true and that all statements made on information and belief are believed to be true.

Further Declarant sayeth not.

Date:

2/15/05


Richard Heuser, M.D., F.A.C.O., F.A.C.P.

EVIDENCE APPENDIX

Item 18

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

APPLICANT: James P. Elia)	
)	
SERIAL NO.: 09/836,750)	EXAMINER: Elizabeth C. Kemmerer
)	
FILED: April 17, 2001)	
)	
FOR: METHOD FOR GROWING)	GROUP ART UNIT: 1646
MUSCLE IN A HUMAN HEART)	

**FOURTH SUPPLEMENTAL DECLARATION
OF RICHARD HEUSER, M.D., F.A.C.C., F.A.C.P.**

I, Richard Heuser, declare as follows:

1. I have offices at 500 West Thomas Road, Suite 900, Phoenix, Arizona 85013.
2. This Fourth Supplemental Declaration is submitted in addition to my previous Declaration, dated June 5, 2003, my Supplemental Declaration dated February 4, 2004, my Second Supplemental Declaration dated July 18, 2004, and my Third Supplemental Declaration dated February 15, 2005. No changes are made to any of such previous Declarations.
3. My Curriculum Vitae (hereinafter "CV") is attached as Exhibit A to my Declaration of June 5, 2003, and my background is further amplified by materials submitted in my Second and Third Supplemental Declarations.

4. I have read and understood the disclosures of the above-referenced patent application at page 20, line 10 through page 21, line 15; and page 44, line 19 through page 46, line 16. Such disclosures are the same as I read and understood in my previous Declaration and Supplemental Declaration. A copy of such disclosures is attached hereto as Fourth Supplemental Declaration Exhibit A.

I have also read and understood additional disclosures of the above-referenced patent application at page 33, lines 8-10; page 37, lines 19-25; page 40, line 20 through page 43 line 3; page 44, lines 12 and 13; page 48, lines 13-15; page 53, line 1 through page 56, line 25; and page 62, lines 1-10. A copy of such additional disclosures is attached hereto as Fourth Supplemental Declaration Exhibit B.

5. The disclosures in Fourth Supplemental Declaration Exhibit A, also contained in my previous Declaration and Supplemental Declaration, relate to using growth factors, including cells, for promoting the growth of soft tissue and, more specifically, to a method which may use such growth factors for growing a new portion of a human heart by growing new cardiac muscle. Such disclosures are also directed to the growth of new arteries in the heart.

I understand that the additional disclosures in Fourth Supplemental Declaration Exhibit B relate to using cellular growth factors, including bone marrow stem cells, to grow soft tissue, including an artery. Stem cells harvested from bone marrow, peripheral blood and from culture banks are described as being implanted for promoting morphogenesis and growth of all three-germ tissue layers, i.e. mesoderm, ectoderm and endoderm tissues. It would be understood by one skilled in the art that morphogenesis includes the growth of an artery, which comprises mesodermal tissue.

6. I have read and understood the claims set forth in Fourth Supplemental Declaration Exhibit C and have been informed that such claims will be concurrently presented in this application with this Fourth Supplemental Declaration.
7. Based upon above Paragraphs 4-6, it is my opinion that one skilled in the medical arts, armed with the knowledge in the disclosures referenced therein, would be enabled to practice the method set forth in Fourth Supplemental Declaration Exhibit C and to predictably anticipate the results defined therein without need for resorting to undue experimentation. It is my further opinion that one skilled in the art reading such disclosures would understand that all of the well known administration procedures described at page 45 of the patent application, including intravenous, intraluminal, intramuscular, and with an angioplasty balloon, would be applicable for use in growing an artery in a human patient regardless of whether the genetic material was a gene; cell, including stem cells such as bone marrow stem cells; or another type of growth factor.

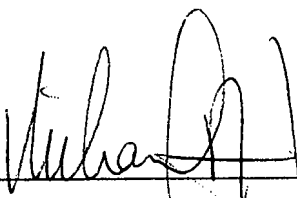
Declarant states that the above opinion was reached independently.

Declarant understands that (1) any willful false statements and the like made herein are punishable by fine or imprisonment, or both (18 U.S.C. 1001) and may jeopardize the validity of the application or any patent issuing thereon, and (2) that all statements made of Declarant's own knowledge are true and that all statements made on information and belief are believed to be true.

Further Declarant sayeth not.

Date: _____

6/16/00



Richard Heuser, M.D., F.A.C.C., F.A.C.P.

**FOURTH
SUPPLEMENTAL
DECLARATION**

EXHIBIT A

DISCLOSURES

EXHIBIT A
DISCLOSURES
APPLICATION SERIAL NO. 09/836,750

PAGE 20, LINE 10 – PAGE 21, LINE 15

Growth factors can be utilized to induce the growth of “hard tissue” or bone and “soft tissues” like ectodermal and mesodermal tissues. As used herein, the term growth factor encompasses compositions and living organisms which promote the growth of hard tissue, such as bone, or soft tissue, in the body of a patient. The compositions include organic and inorganic matter. The compositions can be genetically produced or manipulated. The living organisms can be bacteria, viruses, or any other living organism which promote tissue growth. By way of example and not limitation, growth factors can include platelet-derived growth factor (PDGF), epidermal growth factor (EGF), fibroblast growth factor (acidic/basic (FGF a,b), interleukins (IL’s), tumor necrosis factor (TNF), transforming growth factor (TGF-B), colony-stimulating factor (CSF), osteopontin (Eta-1 OPN), platelet-derived growth factor (PDGF), interferon (INF), bone morphogenic protein 1 (BMP-1), and insulin growth factor (IGF). Recombinant and non-recombinant growth factors can be utilized as desired. Bacteria or viruses can, when appropriate, be utilized as growth factors. For example, there is a bacterial hydrophilic polypeptide that self-assembles into a nanometer internal diameter pore to build a selective lipid body. Various enzymes can be utilized for the synthesis of peptides which contain amino acids that control three-dimensional protein structure and growth. Growth factors can be applied in gels or other carriers which regulate the rate of release of the growth factors and help maintain the growth factors and the carrier, at a desired location in the body. Time release capsules, granules, or other carriers containing growth factor can be activated by tissue pH, by enzymes, by ultrasound,

by electricity, by heat, by selected *in vivo* chemicals or by any other selected means to release the growth factor. The carrier can be resorbable or non-resorbable. Or, the growth factor itself can be activated by similar means. Either the carrier or the growth factor can mimic extracellular fluid to control cell growth, migration, and function. The growth factor can be administered orally, systemically, in a carrier, by hypodermic needle, through the respiratory tract, or by any other desired method. The growth factor can also be administered into a capsule or other man-made composition or structure placed in the body. While administration of the growth factor is presently usually localized in the patient's body, circumstances may arise where it is advantageous to distribute a growth factor throughout the patient's body in uniform or non-uniform concentrations. An advantage to growth factors is that they can often, especially when in capsule form or in some other containment system, be inserted to a desired site in the body by simply making a small incision and inserting the growth factor. The making of such small incision comprises minor surgery which can often be accomplished on an out-patient basis. The growth factors can be multifactorial and nonspecific.

PAGE 44, LINE 19 – PAGE 46, LINE 16

Genetic material comprising a portion of a gene, a gene, genes, a gene product (i.e., a composition a gene causes to be produced like, for example, an organ-producing growth factor), growth factor, or an ECM (extracellular matrix) can be used in or on the body to grow an organ to tissue. For example, the vascular epithelial growth factor gene (VEGF) or its growth factor equivalent can be inserted into the body to cause an artery to grow. When insertion of a gene, portion of a gene, gene product, growth factor, or ECM *in vivo* or *ex vivo* is referred to herein in connection with any of the implant techniques of the invention, it is understood that a cell

nutrient culture(s), physiological nutrient culture(s), carrier (s), enhancer(s), promoter(s), or any other desired auxiliary component(s) can be inserted with the gene or at the same location as the gene, growth factor, ECM, etc.

An artery is an organ from the circulatory system. An artery can be grown in the heart, legs, or other areas by injecting a gene or other genetic material into muscle at a desired site. Size, vascularity, simplicity of access, ease of exploitation, and any other desired factors can be utilized in selecting a desired site. The gene is one of several known VEGF genes which cause the production of vascular endothelial growth factors. Several VEGF genes which produce vascular endothelial growth factors are believed to exist because nature intends for there to be several pathways (i.e., genes) which enable the production of necessary growth factors. The existence of several pathways is believed important because if one of the genes is damaged or inoperative, other similar genes can still orchestrate the production of necessary growth factors. VEGF genes are used by the body to promote blood vessel growth. VEGF genes are assimilated (taken in) by muscle cells. The genes cause the muscle cells to make a VEGF protein which promotes the growth of new arteries. VEGF proteins can be made in a lab and injected into a patient intravenously, intraluminally, or intramuscularly to promote the growth of an artery. Or, the genes (or other genetic material) can be applied with an angioplasty balloon, with the assistance of a vector, or by any other method.

It is not always desirable to grow a completely new organ. Sometimes growing a portion of an organ is desirable. For example, in some heart attacks or strokes, a portion of the heart or brain remains viable and a portion dies. An injection of a gene to form cardiac muscle and/or an injection of a gene to form an artery can be utilized to revive or replace the dead portion of the heart. The dead portion of the heart may (or may not) be used as a matrix while the new muscles

and vessels grow. Thus, in this example, a partial new organ is grown in a pre-existing organ. A pacemaker may (or may not) be necessary. A second injection of a gene may (or may not) be necessary to stop cardiac muscle growth once it is completed. Portions of organs throughout the body can similarly be repaired or replaced. It may be necessary to provide gene(s) or growth factor(s) sequentially. For instance, one or more blood vessels are grown by inserting an appropriate gene or other genetic material into a selected area. Second, an appropriate gene or other genetic material is inserted in the selected area to grow a bone or other organ.

The size and shape limitation of the desired structure can come from a containment and boundary contact inhibition phenomenon or by a chemical inhibition.

A variation on the theme of growing a portion of an organ is as follows: a portion of a heart dies. The pericardium is utilized as a scaffold and seeded with cells and/or genes to grow new muscle, and genes (or other genetic material) to grow new arteries. Immediately adjacent the dead cardiac muscle, onto or into the pericardium, the appropriate cells, genes, and/or growth factors (or other genetic material) are placed. Once the new muscle and blood vessels have grown, the function specific tissue can be applied to the damaged portion of the heart and paced, if necessary, to augment cardiac action. If the surgeon desires, the dead muscle can be removed and the new muscle and blood vessels can be surgically rotated into the excised region and secured. This probably can be done endoscopically. In essence, the pericardium is utilized to allow the new muscle wall to grow. The new muscle wall is then transplanted into the damaged heart wall. This procedure utilizes the body as a factor to grow an organ and/or tissue, after which the organ and/or tissue is transplanted to a desired region. On the other hand, the new muscle wall may integrate itself into the old wall and not require transplantation.

**FOURTH
SUPPLEMENTAL
DECLARATION**

EXHIBIT B

DISCLOSURES

EXHIBIT B
DISCLOSURES
APPLICATION SERIAL NO. 09/836,750

PAGE 33, LINES 8-10

Morphogenesis or morphogenetics is the origin and evolution of morphological characters and is the growth and differentiation of cells and tissues during development.

PAGE 37, LINES 19-25

Multifactorial and nonspecific cells (such as stem cells and germinal cells) can provide the necessary in vivo and in vitro cascade of genetic material once an implanted master control gene's transcription has been activated. Likewise, any host cell, clone cell, cultured cell, or cell would work. Genetic switches (such as the insect hormone ecdysone) can be used to control genes inserted into humans and animals. These gene switches can also be used in cultured cells or other cells. Gene switches govern whether a gene is on or off making possible precise time of gene activity.

PAGE 40, LINE 20 – PAGE 43, LINE 3

EXAMPLE 11

MSX-1 and MSX-2 are the homeobox genes that control the generation and growth of a tooth. A sample of skin tissue is removed from the patient and the MSX-1 and MXS-2 homeobox gene(s) are removed from skin tissue cells. The genes are stored in an appropriate nutrient culture medium.

BMP-2 and BMP-4 growth factors are obtained by recombinant or natural extraction from bone.

Living stem cells are harvested from the bone marrow, the blood of the patient, or from cell culture techniques. The stem cells are placed in a nutrient culture medium at 98.6 degrees. The temperature of the culture medium can be varied as desired but ordinarily is between 40 to 102 degrees F.

MXS-1 and MXS-2 transcription factors are obtained which will initiate the expression of the MXS-1 and MXS-2 homeobox genes.

The MXS-1 and MXS-2 transcription factors, BMP-2 and BMP-4 bone morphogenic proteins, and MXS-1 and MXS-2 genes are added to the nutrient culture medium along with the living stem cells.

EXAMPLE 12

Example 11 is repeated except that the transcription factors bind to a receptor complex in the stem cell nucleus.

EXAMPLE 13

Example 11 is repeated except that the MXS-1 and MXS-2 transcription factors are not utilized. The transcription of the MXS-1 and MXS-2 homeobox genes is activated by applying an electric spark to the nutrient culture medium.

EXAMPLE 14

Example 13 is repeated except that the stem cells are starved and the transcription of the MXS-1 and MXS-2 homeobox genes is activated by applying an electric spark to the nutrient culture medium.

EXAMPLE 15

WT-1 and PAX genes are obtained from a sample of skin tissue is removed from the patient. The genes are stored in an appropriate nutrient culture medium. PAX genes produce PAX-2 and other transcription factors.

BMP-7 and other kidney related BMP growth factors are obtained by recombinant or natural extraction from bone.

Living stem cells are harvested from the bone marrow, the blood of the patient, or from cell culture techniques. The stem cells are placed in a nutrient culture medium at 98.6 degrees. The temperature of the culture medium can be varied as desired but ordinarily is between 40 and 102 degrees F.

The WT-1 and PAX genes, and BMP-7 and other kidney BMPS are added to the nutrient culture medium along with the living stem cells.

A primitive kidney germ is produced. The kidney germ is transplanted in the patient's body near a large artery. As the kidney grows, its blood supply will be derived from the artery.

EXAMPLE 16

The Aniridia gene is obtained from a sample of skin tissue is removed from the patient. The gene(s) is stored in an appropriate nutrient culture medium.

Aniridia transcription factor (activates expression of the Aniridia gene) and growth factors (function to help stem cells differentiate during morphogenesis to form an eye) are obtained.

Living stem cells are harvested from the bone marrow, the blood of the patient, or from cell culture techniques. The stem cells are placed in a nutrient culture medium at 98.6 degrees.

The temperature of the culture medium can be varied as desired but ordinarily is between 40 to 102 degrees F.

The Aniridia transcription factor and growth factors and the Aniridia gene are added to the nutrient culture medium along with the living stem cells.

A primitive eye germ is produced. The kidney germ is transplanted in the patient's body near the optic nerve. As the kidney grows, its blood supply will be derived from nearby arteries.

EXAMPLE 17

The Aniridia gene is obtained from a sample of skin tissue is removed from the patient. The gene(s) is stored in an appropriate nutrient culture medium.

Aniridia transcription factor (activates expression of the Aniridia gene) and growth factors (function to help stem cells differentiate during morphogenesis to form an eye) are obtained and added to the nutrient culture medium.

An eye germ develops. A branch of the nearby maxillary artery is translocated to a position adjacent the eye germ to promote the development of the eye germ. The eye germ matures into an eye which receives its blood supply from the maxillary artery.

The term "cell nutrient culture" as used herein can include any or any combination of the following: the extracellular matrix; conventional cell culture nutrients; and/or, a cell nutrient such as a vitamin. As such, the cell nutrient culture can be two-dimensional, three dimensional, or simply a nutrient, and is useful in promoting the processes of cellular dedifferentiation, redifferentiation, differentiation, growth, and development.

PAGE 44, LINES 12– 13

An organ, as used herein, consists of two or more kinds of tissues joined into one structure that has a certain task.

PAGE 48, LINES 13– 15

In the example above, if germinal cells (and in some cases, stem cells) are utilized a direct differentiation and morphogenesis into an organ can occur in vivo, ex vivo, or in vitro.

PAGE 53, LINE 1 – PAGE 56, LINE 25

EXAMPLE 18

A 36 year old Caucasian male experiences pain in his left leg. A medical examination reveals a damaged one inch long section of a large artery in his left leg. The examination also reveals that this damaged section of the artery is nearly completely clogged with plaque and that the wall of the artery is weakened. The weakening in the arterial wall makes attempting to clean out the artery risky and also makes it risky to attempt to insert a stent in the artery.

Recombinant cDNA encoded to combine with a cell ribosome to produce the human growth factor VEGF is assembled into a eukaryotic expression plasmid. The recombinant cDNA is from cDNA libraries prepared from HL60 leukemia cells and is known to cause the growth of arteries. The plasmid is maintained at a room temperature of 76 degrees F.

The clones are placed in 1.0 milliliters of a normal saline carrier solution at a room temperature of 76 degrees F to produce an genetic carrier solution. The genetic carrier solution contains about 250 ug of the cDNA clones. A nutrient culture can, if desired, be utilized in conjunction with or in place of the saline carrier. Each clone is identical. If desired, only a

single clone can be inserted in the normal saline carrier solution. The saline carrier solution comprises 0.09% by weight sodium chloride in water. A saline carrier solution is selected because it will not harm the DNA clone.

Two sites are selected for injection of the genetic carrier solution. While the selection of sites can vary as desired, the sites are selected at the lower end (the end nearest the left foot of the patient) of the damaged section of the artery so that the new arterial section grown can, if necessary, be used to take the place of the damaged section of the artery in the event the damaged section is removed.

The first site is on the exterior wall of the artery on one side of the lower end of the damaged section of the artery. A containment system is placed at the first site.

The second site is inside the wall of the artery on the other side of the lower end of the artery.

The genetic carrier solution is heated to a temperature of 98.6 degrees F. 0.25 milliliters of the genetic carrier solution is injected into the containment system at the first site. 0.25 milliliters of the genetic carrier solution is injected at the second site inside the wall of the artery. Care is taken to slowly inject the genetic carrier solution to avoid entry of the solution into the artery such that blood stream will carry away the cDNA in the solution.

After two weeks, an MRI is taken which shows the patient's leg artery. The MRI reveals new growth at the first and second sites.

After four weeks, another MRI is taken which shows the patient's leg artery. The MRI shows that (1) at the first site a new artery is growing adjacent the patient's original leg artery, and (2) at the second site a new section of artery is growing integral with the original artery, i.e., at the second site the new section of artery is lengthening the original artery, much like inserting

a new section of hose in a garden hose concentric with the longitudinal axis of the garden hose lengthens the garden hose.

After about eight to twelve weeks, another MRI is taken which shows that the new artery growing adjacent the patient's original artery has grown to a length of about one inch and has integrated itself at each of its ends with the original artery such that blood flows through the new section of artery. The MRI also shows that the new artery at the second site has grown to a length of one-half inch.

In any of the examples of the practice of the invention included herein, cell nutrient culture can be included with the gene, the growth factor, the extracellular matrix, or the environmental factors.

In any of the examples of the practice of the invention included herein, the concept of gene redundancy can be applied. For example, the Examples 1 to 14 concerning a tooth list the genes MSX-1 and MSX-2. These genes differ by only two base pairs. Either gene alone may be sufficient. A further example of redundancy occurs in growth factors. Looking at the Examples 10 to 14, BMP4 or BMP2 alone may be sufficient. Redundancy can also be utilized in connection with transcription factors, extracellular matrices, environmental factors, cell nutrient cultures, physiological nutrient cultures, vectors, promoters, etc.

One embodiment of the invention inserts genetic material (gene, growth factor, ECM, etc.) into the body to induce the formation of an organ. Similar inducing materials inserted ex vivo into or onto a living cell in an appropriate physiological nurturing environment will also induce the growth of an organ. The VCSEL laser allows early detection in a living cell of a morphogenic change indicating that organ formation has been initiated. With properly timed transplantation, organ growth completes itself.

During the ex vivo application of the invention, a gene and/or growth factor is inserted into a cell or a group of cells; an ECM or environmental factor(s) are placed around and in contact with a cell or group of cells; or, genetic material is inserted into a subunit of a cell to induce organ growth. An example of a subunit of a cell is an enucleated cell or a comparable artificially produced environment. In in vivo or ex vivo embodiments of the invention to induce the growth of an organ, the genes, growth factors, or other genetic material, as well as the environmental factors or cells utilized, can come from any desired source.

EXAMPLE 19

Genetically produced materials are inserted in the body to cause the body to grow, reproduce, and replace in vivo a clogged artery in the heart. This is an example of site-specific gene expression. A plasmid expression vector containing an enhancer/promoter is utilized to aid in the transfer of the gene into muscle cells. The enhancer is utilized to drive the specific expression of the transcriptional activator. After the enhancer drives the expression of the transcriptional activator, the transcriptional activator transactivates the muscle/artery genes. Saline is used as a carrier. Cardiac muscle can take up naked DNA injected intramuscularly. Injecting plasmid DNA into cardiac (or skeletal) muscle results in expression of the transgene in cardiac myocytes for several weeks or longer.

Readily available off-the-shelf (RAOTS) cDNA clones for recombinant human VEGF₁₆₅, isolated from cDNA libraries prepared from HL60 leukemia cells, are assembled in a RAOTS expression plasmid utilizing 736 bp CMV promoter/enhancer to drive VEGF expression. Other RAOTS promoters can be utilized to drive VEGF expression for longer periods of time. Other RAOTS recombinant clones of angiogenic growth factors other than VEGF can be utilized, for example, fibroblast growth factor family, endothelial cell growth

factor, etc. Downstream from the VEGF cDNA is an SV40 polyadenylation sequence. These fragments occur in the RAOTS pUC118 vector, which includes an Escherichia coli origin of replication and the Beta lactamase gene for ampicillin resistance.

The RAOTS construct is placed into a RAOTS 3 ml syringe with neutral pH physiologic saline at room temperature (or body temperature of about 37 degrees C). The syringe has a RAOTS 27 gauge needle.

Access to the cardiac muscle is gained by open heart surgery, endoscopic surgery, direction injection of the needle without incision, or by any other desired means. The cardiac muscle immediately adjacent a clogged artery is slowly injected with the RAOTS construct during a five second time period. Injection is slow to avoid leakage through the external covering of muscle cells. About 0.5 ml to 1.0 ml (milliliter) of fluid is injected containing approximately 500 ug phVEGF165 in saline (N=18). The readily available off-the-shelf cDNA clones cause vascular growth which automatically integrates itself with the cardiac muscle. Anatomic evidence of collateral artery formation is observed by the 30th day following injection to the RAOTS construct. One end of the artery integrates itself in the heart wall to receive blood from the heart. The other end of the artery branches into increasing smaller blood vessels to distribute blood into the heart muscle. Once the growth of the new artery is completed, the new artery is left in place in the heart wall. Transplantation of the new artery is not required.

Blood flow through the new artery is calculated in a number of ways. For example, Doppler-derived flow can be determined by electromagnetic flowmeters (using for example, a Doppler Flowmeter sold by Parks Medical Electronic of Aloha, Oregon) both in vitro and in vivo. RAOTS external ultrasound gives a semiquantitative analysis of arterial flow. Also, RAOTS angiograms or any other readily available commercial devices can be utilized.

VEGF gene expression can be evaluated by readily available off-the-shelf polymerase chain reaction (PCR) techniques.

If controls are desired, the plasmid pGSVLacZ containing a nuclear targeted Beta-galactosidase sequence coupled to the simian virus 40 early promoter can be used. To evaluate efficiency, a promoter-matched reporter plasmid, pCMV Beta (available from Clontech of Palo Alto, California), which encodes Beta-galactosidase under control of CMV promoter/enhancer can be utilized. Other RAOTS products can be utilized if desired.

EXAMPLE 20

A patient, a forty year old African-American female in good health, has been missing tooth number 24 for ten years. The space in her mouth in which her number 24 tooth originally resided is empty. All other teeth except tooth number 24 are present in the patient's mouth. The patient desires a new tooth in the empty "number 24" space in her mouth.

A full thickness mucoperiosteal flap surgery is utilized to expose the bone in the number 24 space. A slight tissue reflection into the number 23 tooth and number 25 tooth areas is carried out to insure adequate working conditions.

A Midwest Quietair handpiece (or other off-the-shelf handpiece) utilizing a #701XXL bur (Dentsply Midwest of Des Plaines, Illinois) (a #700, #557, #558, etc. bur can be utilized if desired) is used to excavate an implant opening or site in the bone. The implant opening is placed midway between the roots of the number 23 and number 25 teeth. The opening ends at a depth which is about fifteen millimeters and which approximates the depth of the apices of the roots of the number 23 and number 25 teeth. Care is taken not to perforate either the buccal or lingual wall of the bone. In addition, care is taken not to perforate or invade the periodontal ligament space of teeth numbers 23 and 25.

An interrupted drilling technique is utilized to avoid overheating the bone when the #701XXL bur is utilized to form the implant opening. During a drilling sequence, the drill is operated in five second increments and the handpiece is permitted to stall. Light pressure and a gentle downward stroke are utilized.

PAGE 62, LINES 1-10

EXAMPLE 36

Example 18 is repeated except that the patient is a 55 year old Caucasian male, and the genetic carrier solution is injected into two sites in the coronary artery of the patient. The first site is on the exterior wall on one side of the artery. The second site is inside the wall of the artery on the other side of the artery. A section of the artery is damaged, is partially blocked, and has a weakened wall. The first and second sites are each below the damaged section of the artery. Similar results are obtained, i.e., a new section of artery grows integral with the original artery, and a new section of artery grows adjacent the original artery. The new section of artery has integrated itself at either end with the original artery so that blood flows through the new section of artery.

**FOURTH
SUPPLEMENTAL
DECLARATION**

EXHIBIT C

CLAIMS

EXHIBIT C

CLAIMS **APPLICATION SERIAL NO. 09/836,750**

- Claim 236 A method of growing a new portion of a pre-existing heart comprising the steps of placing a growth factor in a body of a human patient and growing new cardiac muscle and growing a new artery in said heart.
- Claim 238 The method of claim 236, further comprising repairing a dead portion of said heart.
- Claim 239 The method of claim 236, further comprising repairing a damaged portion of said heart.
- Claim 243 The method of claim 236, wherein said growth factor comprises a member selected from the group consisting of cells, cellular products, and derivatives of cellular products.
- Claim 244 The method of claim 243, wherein said growth factor comprises a cell.
- Claim 245 The method of claim 244, wherein said cell is multifactorial and non-specific.
- Claim 246 The method of claim 245, wherein said cell comprises a stem cell.
- Claim 247 The method of claim 236, wherein said growth factor is placed in said patient by injection.
- Claim 248 The method of claim 247, wherein said injection is intravenous.
- Claim 249 The method of claim 247, wherein said injection is intraluminal.
- Claim 250 The method of claim 247, wherein said injection is intramuscular.

- Claim 251 The method of claim 236, wherein said growth factor is placed in said patient by a carrier.
- Claim 252 The method of claim 251, wherein said carrier comprises an angioplasty balloon.
- Claim 253 The method of claim 236, wherein said growth factor comprises a gene and a cell.
- Claim 257 The method of claim 236, wherein said growth factor is locally placed in said body.
- Claim 258 The method of claim 238, wherein said growth factor is locally placed in said body.
- Claim 259 The method of claim 239, wherein said growth factor is locally placed in said body.
- Claim 260 The method of claim 243, wherein said growth factor is locally placed in said body.
- Claim 261 The method of claim 236, wherein said growth factor comprises living stem cells harvested from bone marrow.
- Claim 262 The method of claim 238, wherein said growth factor comprises living stem cells harvested from bone marrow.
- Claim 263 The method of claim 239, wherein said growth factor comprises living stem cells harvested from bone marrow.
- Claim 264 A method of growing a new portion of a pre-existing heart comprising locally placing a growth factor comprising a stem cell in a body of a human patient to grow new cardiac muscle in said heart.

- Claim 265 The method of claim 264, wherein said stem cell is placed in said patient by injection.
- Claim 266 The method of claim 264, wherein said stem cell comprises living stem cells harvested from bone marrow.
- Claim 267 The method of claim 266, wherein said stem cell is placed in said patient by injection.
- Claim 268 The method of claim 262, wherein said stem cell is placed in said patient by injection.
- Claim 269 The method of claim 263, wherein said stem cell is placed in said patient by injection.
- Claim 270 The method of claim 258, wherein said growth factor comprises a cell and said cell is placed adjacent to said dead portion of said heart.
- Claim 271 The method of claim 259, wherein said growth factor comprises a cell and said cell is placed adjacent to said damaged portion of said heart.
- Claim 272 The method of claim 265, wherein said stem cell is injected into said heart.
- Claim 273 The method of claim 267, wherein said stem cell is injected into said heart.
- Claim 274 The method of claim 238, wherein said growth factor comprises a cell and said cell is placed in said body by intravenous injection.
- Claim 275 The method of claim 239, wherein said growth factor comprises a cell and said cell is placed in said body by intravenous injection.

- Claim 276 The method of claim 238, wherein said growth factor comprises a cell and said cell is placed in said body by intraluminal injection.
- Claim 277 The method of claim 239, wherein said growth factor comprises a cell and said cell is placed in said body by intraluminal injection.
- Claim 278 The method of claim 238, wherein said growth factor comprises a cell and said cell is placed in said body by an angioplasty balloon.
- Claim 279 The method of claim 239, wherein said growth factor comprises a cell and said cell is placed in said body by an angioplasty balloon.
- Claim 280 The method of claim 236 further comprising determining blood flow through said newly grown artery.
- Claim 281 The method of claim 238 further comprising determining blood flow through said newly grown artery.
- Claim 282 The method of claim 239 further comprising determining blood flow through said newly grown artery.
- Claim 283 The method of claim 236 further comprising observing said newly grown artery.
- Claim 284 The method of claim 238 further comprising observing said newly grown artery.
- Claim 285 The method of claim 239 further comprising observing said newly grown artery.

- Claim 286 A method of repairing a dead portion of a pre-existing heart comprising the steps of placing stem cells adjacent said dead portion; forming a new artery in said heart, thereby causing said dead portion of said heart to be repaired.
- Claim 287 The method of claim 286, wherein said stem cells are placed by injection.
- Claim 288 The method of claim 286, wherein said stem cells are placed by intraluminal administration.
- Claim 289 The method of claim 286, wherein said stem cells are placed by an angioplasty balloon.
- Claim 290 A method of repairing a damaged portion of a pre-existing heart comprising the steps of placing stem cells adjacent said damaged portion; forming a new artery in said heart, thereby causing said damaged portion of said heart to be repaired.
- Claim 291 The method of claim 290, wherein said stem cells are placed by injection.
- Claim 292 The method of claim 290, wherein said stem cells are placed by intraluminal administration.
- Claim 293 The method of claim 290, wherein said stem cells are placed by an angioplasty balloon.

EVIDENCE APPENDIX

Item 19

CERTIFICATE OF MAILING

I hereby certify that the attached DECLARATION OF ANDREW E. LORINCZ, M.D.
was delivered to the Assistant Commissioner for Patents by the undersigned from Arrow
Intellectual Property Services, 2001, Jefferson Davis Highway, Suite 602, Arlington, Virginia
22202, by hand carrying said DECLARATION to Art Unit 1646, Crystal Plaza 1, Tenth Floor,
Attention: Examiner Elizabeth C. Kemmerer this 17th day of June, 2003.

Dated: June 17, 2003

Ann Rutledge
Printed Name: Ann Rutledge

Docket No. ~~XXXXXXXX~~ 1000-10-CO1
Serial No. ~~XXXXXX~~ 09/836,750
Filed ~~XXXXXXXX~~ 04/17/01
Due Date: _____

ARROW INTELLECTUAL PROPERTY SERVICE

The Patent Office acknowledges, and has stamped hereon, the date of receipt
of the items check below:

- ☐ Transmittal Letter
- ☐ Application - Trademark
- ☐ Application - Patent Specification Total Pgs _____
- ☐ Total Claims _____ Ind. Claims _____ Total Pgs _____
- ☐ _____ Fee: \$ _____
- ☐ Abstract Total Pgs _____
- ☐ Drawings: Formal _____ Informal _____ Total Pgs _____
- ☐ Declaration/Oath/Power of Attorney Total Pgs _____
- ☐ Assignment Fee: \$ _____
- ☐ Request for Non-Publication
- ☐ Information Disclosure Statement
- ☐ Form PTO-1449 References _____ Total No _____
- ☐ Request for Extension of Time Fee: \$ _____
- ☐ Amendment/Response
- ☒ ~~XXXXXXXX~~ Declaration of Andrew Lorincz
- ☐ Brief/Reply Brief/Notice of Appeal
- ☐ Fee-Base/Maintenance Fee: \$ _____
- ☐ Check No. _____ Fee: \$ _____
- ☐ _____
- ☐ _____

COPY

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

APPLICANT: James P. Elia)	
)	
SERIAL NO.: 09/836,750)	EXAMINER: E.C. Kemmerer, Ph.D.
)	
FILED: April 17, 2001)	
)	GROUP ART UNIT: 1646
FOR: METHOD AND APPARATUS)	
FOR INSTALLATION OF)	
DENTAL IMPLANT)	

DECLARATION OF ANDREW E. LORINCZ, M.D.

I, Andrew E. Lorincz, declare as follows:

1. I reside at 3628 Belle Meade Way, Mountain Brook, Alabama 35223.
2. My Curriculum Vitae is attached hereto as Exhibit A.
3. I have read and understood the disclosures of the above-referenced patent application at page 20, line 10 through page 21, line 15; and page 44, line 19 through page 46, line 16. A copy of such disclosures is attached hereto as Exhibit B.
4. I note that the disclosures referenced in above Paragraph 3 relate to using a growth factor for promoting the growth of soft tissue and, more specifically, to a method of using a growth factor for growing muscle in a human heart.

5. I am aware of and have considered the definition of *growth factor* in the specification of the above-referenced patent application at page 20, line 10 through page 21, line 15. Such definition is set forth in Exhibit C along with a definition from the medical dictionary, MEDLINE plus: Merriam-Webster Medical Dictionary. A service of the U.S. NATIONAL LIBRARY OF MEDICINE and the NATIONAL INSTITUTES OF HEALTH. I find that the dictionary definition is consistent with that contained at page 20, line 10 through page 21, line 15 of the above-referenced patent application. I believe that both definitions are appropriate for use in the field of tissue growth and would be understood by one skilled in the medical arts. Accordingly, I am adopting and utilizing the definition contained in the patent application throughout this declaration.
6. I have read and understood the claims set forth in Exhibit D and have been informed that such claims will be presented to the Patent and Trademark Office in the near future.
7. The materials included in attached Exhibit E illustrate that placement of a growth factor in a human patient causes muscle growth in a heart. These materials report work performed by reputable, skilled scientists and reputable organizations in the medical arts. Consequently, I believe that these reports would be recognized as clearly valid by one of ordinary skill in the medical arts because they report the results of scientific tests conducted by competent, disinterested third parties with use of proper scientific controls.
8. Based upon above Paragraphs 3-7, it is my opinion that introducing a growth factor into a human patient will predictably cause new muscle growth in the heart of the patient.

9. Based upon above Paragraphs 3-6, it is my opinion that one skilled in the medical arts, armed with the knowledge in such paragraphs, would be able to practice the method set forth in Exhibit D without need for resorting to undue experimentation.
10. Declarant states that the above opinion was reached independently.

Declarant understands that (1) any willful false statements and the like made herein are punishable by fine or imprisonment, or both (18 U.S.C. 1001) and may jeopardize the validity of the application or any patent issuing thereon, and (2) that all statements made of Declarant's own knowledge are true and that all statements made on information and belief are believed to be true.

Further Declarant sayeth not.

Date: 6-9-03

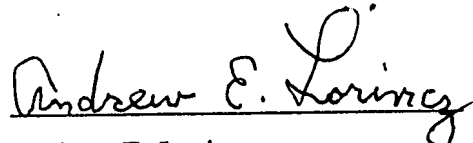

Andrew E. Lorincz

EXHIBIT A

CURRICULUM VITAE

NAME: Andrew E. Lorincz, M.D.
TITLE: Professor of Pediatrics
University of Alabama at Birmingham
SOCIAL SECURITY: 361-16-4853
ADDRESSES: University of Alabama at Birmingham
Mortimer Jordan Hall B-70
1825 University Boulevard
Birmingham, Alabama 35294-2010
Telephone: (205) 934-7038
FAX: (205) 975-9147
Email: aelorincz@vprua.vprua.UAB.edu

Home:

3628 Belle Meade Way
Mountain Brook, Alabama 35223
Telephone: (205) 967-4678

BIRTH: 5/17/26 Chicago, Illinois

MARITAL STATUS: Married, 12/14/65 - Diane DeNyse Lorincz

EDUCATION:

1948-1952 University of Chicago, School of Medicine, M.D. Degree
1948-1950 University of Chicago, B.S. Degree (Anatomy & Biochemistry)
1946-1948 University of Chicago, Ph.B. Degree

POSTDOCTORAL EDUCATION:

Jan-Mar 1980 Lysosomal Storage Disease Laboratory, Eunice Kennedy Shriver Center,
Waltham, MA (Harvard), Visiting Scientist
1955-1956 LaRabida Jackson Park Sanitarium, University of Chicago,
Junior Staff Physician Department of Pediatrics, University of Chicago
Clinics Bob Roberts Memorial Hospital
1955-1958 Arthritis and Rheumatism Foundation Fellow
1954-1955 Benjamin J. Rosenthal Clinical and Research Fellow
1953-1954 Junior Assistant Resident
1952-1953 Intern

ACADEMIC APPOINTMENTS:

1996-present	Professor Emeritus, Department of Pediatrics
1984-1996	School of Public Health, University of Alabama at Birmingham, Professor
1971-1996	University of Alabama at Birmingham, Member of Graduate Faculty
1968-1996	University of Alabama at Birmingham, Professor of Pediatrics
1971-1984	Division of Engineering Biophysics, University of Alabama at Birmingham, Associate Professor
1968-1982	University of Alabama at Birmingham, Associate Professor of Biochemistry
1976- 1980	School of Optometry, University of Alabama at Birmingham, Professor Optometry
1971-1980	School of Nursing, University of Alabama at Birmingham, Clinical Associate Professor
1970-1980	Center for Developmental and Learning Disorders, University of Alabama at Birmingham, (A University Affiliated Facility for Developmental Disability), Director
1970-1976	School of Optometry, University of Alabama at Birmingham, Associate Professor of Pediatric Optometry
1966-1968	Medical Teaching and Research, Unit of the University of Florida at the Sunland Training Center, Gainesville, Florida, Director
1963-1968	Department of Surgery (Orthopaedics), University of Florida College of Medicine, Gainesville, Florida, Research Associate Professor
1962-1968	Department of Pediatrics, University of Florida College of Medicine, Gainesville, Florida, Associate Professor
1959-1962	Department of Pediatrics, University of Florida College of Medicine, Gainesville, Florida, Assistant Professor
1956-1959	Department of Pediatrics, University of Chicago School of Medicine, Chicago, Illinois, Instructor

PROFESSIONAL LICENSES - PHYSICIAN AND SURGEON:

5/26/69	State of Alabama
8/10/59	State of Florida (inactive)
9/22/54	State of Illinois (inactive)

SPECIALTY CERTIFICATION:

May 1958 American Board of Pediatrics, Diplomate

BOARDS, COMMITTEES AND CONSULTANTSIPS:

1994-present	Board member of The Mental Retardation and Developmental Disabilities, Health Care Authority of Jefferson County, Inc.
1991-present	Editorial Board for the <u>Annals of Clinical and Laboratory Science</u> , Member
1988-present	Medical Advisory Board of the National MPS Society, Member
1980-1986	<u>Mental Retardation</u> , Consulting Editor
1979-present	National Tay-Sachs and Allied Diseases Association, Scientific Advisory Committee, Member
1978-present	Mayor's Council of Disability Issues
1979-1984	Osteogenesis Imperfecta Foundation, Inc., Board Member Alabama O.I. Chapter
1974-1981	Child Mental Health Services, Inc., Birmingham, Alabama, Board Member
1977-1978	Elizabethtown Committee on Planning and Evaluation, Legislative Committee, State of Pennsylvania
1973-1975	Human Rights Committee for the Partlow State School and Hospital, Tuscaloosa, Alabama, Member - Federal Court Appointed
1971-1974	American Academy of Pediatrics, Committee on Children With Handicaps
1971-1973	<u>American Journal of Mental Deficiency</u> , Consulting Editor
1965-1973	Head Start, Medical Consultant
1967-1972	<u>Journal of Investigative Dermatology</u> , Editorial Consultant
1961-1968	Sunland Hospital, Orlando, Florida, Medical and Research Consultant
1965-1966	State of Florida Interagency Committee on Mental Retardation Planning, Co-Chairman, Mental Retardation Research Committee <u>Alabama Developmental Disabilities Planning Council</u>
1982-1984	Maternal and Child Health, Member of Advisory Committee
1979-1984	Member (Secretary, 1980; Vice Chairman, 1984)
1973-1979	Consultant

American Association of University Affiliated Facilities

1975-1978 American Association of University Affiliated Programs for the Developmentally Disabled, Board Member

American Association on Mental Retardation

1980,84,85	Prevention Committee, Chairman
1980-1982	Member of Council
1978-1980	Medicine Division and Member Executive Committee, Vice President

BOARDS, COMMITTEES AND CONSULTANTSHIPS: (CONTD)

Association of Retarded Citizens of Jefferson County

1990-present	Board Member
1975-1985	Board Member
1977-1978	Second Vice President
1980	Recipient of Distinguished Service Award

PROFESSIONAL SOCIETIES:

American Academy for Cerebral Palsy and Developmental Medicine, (Fellow)
American Academy on Mental Retardation (President Elect, 1975-76; President, 1976-77)
Emeritus Member
American Academy of Pediatrics (Fellow)
American Association for the Advancement of Science
American Association for Clinical Chemistry, Inc.
American Association on Mental Retardation (Fellow)-Life Member
American Chemical Society
American Federation for Clinical Research
American Medical Association
American Society for Human Genetics
American Society for Investigative Pathology
Association of Clinical Scientists
International Society for Mycoplasmaology
Jefferson County Pediatric Society
Society for Complex Carbohydrates
Society for Investigative Dermatology
Society for Pediatric Research
Society for Sigma Xi
Southern Society for Pediatric Research (President, 1964)

BIBLIOGRAPHY

1. Dorfman, A.; Lorincz, A.E.: Occurrence of Urinary Acid Mucopolysaccharides in the Hurler Syndrome. Proceedings of the National Academy of Science, 43:443-46, 1957.
2. Dorfman, A., Gross, J.L.; Lorincz, A.E.: The Treatment of Acute Rheumatic Fever. Pediatrics, 27:692-705, May, 1961.
3. Lorincz, A.E.: Heritable Disorders of Acid Mucopolysaccharide Metabolism in Humans and in Snorter Dwarf Cattle. Annals of the New York Academy of Science, 91:644-58, 1961.
4. Shepard, T.H., Lorincz, A.E., Gartner, S.M.: Desulfuration of Thiourea by Saliva. Proceedings of the Society of Experimental Biology and Medicine, 112:38-42, July, 1963.

BIBLIOGRAPHY (CONT'D)

5. Lorincz, A.E.: A Heritable Disorder of Acid Mucopolysaccharide Metabolism in Cattle Which Resembles the Hurler Syndrome. Proceedings of the Second International Congress on Mental Retardation, Vienna, 1961, Part I: 70-74, 1963.
6. Gennaro, J.F., Jr., Callahan, W.P., III, Lorincz, A.E.: The Anatomy and Biochemistry of a Mucus-secreting Cell Type Present in the Poison Apparatus of the Pit Viper, Ancistrodon Piscivorus Piscivorus. Annals of the New York Academy of Sciences, 106:463-71, March, 1963.
7. Wolfson, S.L., Davidson, E., Harris, J.S., Kahana, L., Lorincz, A.E.: Long-term Corticosteroid Therapy in Hurler Syndrome. American Journal of Diseases in Children, 106:3-10, July, 1963.
8. Wainer, A., Lorincz, A.E.: Studies of Mercapturic Acid Synthesis by Humans. Life Sciences, No.7:504-08, 1963.
9. Lorincz, A.E.: Hurler's Syndrome in Man and Shorter Dwarfism in Cattle: Heritable Disorders of Connective Tissue Acid mucopolysaccharide Metabolism. Clinical Orthopaedics and Related Research, A.F. DePalma (Editor), J.B. Lippincott Co., 33:104-18, 1964.
10. Pearson, H.A., Lorincz, A.E.: A Characteristic Bone Marrow Finding in the Hurler Syndrome. Pediatrics, 34:281-82, August, 1964.
11. Brodgon, B.G., Bartley, T.D., Schiebler, G.L., Shanklin, D.R., Krovetz, L.J., Lorincz, E.E.: Cardiovascular Radiology in Calves. Angiology, 15:496-504, November, 1964.
12. Lorincz, A.E.: Hurler's Syndrome. Medical Aspects of Mental Retardation, C.H. Carter (Editor), Chapter 19, pp. 628-650, 1965.
13. Krovetz, L.J., Lorincz, A.E., Schiebler, G.L.: Cardiovascular Manifestations in the Hurler Syndrome. Hemodynamic and Angiocardiographic Observations in 15 Patients. Circulation, 31:132-141, 1965.
14. Gennaro, J.F., Jr., Lorincz, A.E., Brewster, H.B.: The Anterior Salivary Gland of the Octopus (Octopus Vulgaris) and its Mucous Secretion. Annals of the New York Academy of Science, 118:1021-25, November, 1965.
15. Gessner, I.H., Lorincz, A.E., Bostrom, H.: Acid Mucopolysaccharide Content of the Cardiac Jelly of the Chick Embryo. Journal of Experimental Zoology, 160:291-98, No. 3, December, 1965.
16. Callahan, W.P., Lorincz, A.E.: Hepatic Ultrastructure in the Hurler Syndrome. American Journal of Pathology, 48:277-98, No. 2, February, 1966.
17. Callahan, W.P., Hackett, R.L., Lorincz, A.E.: New Observations by Light Microscopy on Liver Histology in the Hurler's Syndrome. Archives of Pathology, 83:507-12, June, 1967.
18. Lorincz, A.E.: Screening Measurements of Urinary Acid Mucopolysaccharides for Detection of the Hurler Syndrome. The Clinical Pathology of Infancy, Sunderman, F.W. & Sunderman, F.W., Jr. (Editors), pp. 68-71, 1967.
19. Rhoades, R., Lorincz, A.E., Gennaro, J.F., Jr.: Polysaccharide Content of the Poison Apparatus of the Cottonmouth Moccasin, Agkistrodon Piscivorus Piscivorus. Toxicon, 5:125-31, 1967.

BIBLIOGRAPHY (CONT'D)

20. Krovetz, L.J., Lorincz, A.E., Schiebler, G.L.: Cardiovascular Manifestations of the Hunter-Hurler Syndrome. Heart Diseases in Infants, Children and Adolescents, A.J. Moss & F.H. Adams (Editors), The Williams and Wilkins Co. (Publisher), 1968.
21. Lorincz, A.E.: Hurler's Syndrome - Disorders in Systematic Disease - Disorders of the Mucopolysaccharides. The Growth Plate and Its Disorders, M. Rang (Editor), E & S Livingston, Ltd., London, pp. 49-56, 1969.
22. Lorincz, A.E.: Pediatric Dermatology. Dermatology in General Medicine, Fitzpatrick, Thomas, B., arndt, Kenneth, A., Clark, Wallace, H., Eisen, Authur, Z., Van Scott, Eugens, J., Vaughn (Editors), John H. McGraw-Hill, Blakiston Division, N.Y., pp. 1999-2006, 1971.
23. West, S.S., Lorincz, A.E.: Fluorescent Molecular Probes in Fluorescence Microspectrophotometry and Microspectropolarimetry. Fluorescence Techniques in Cell Biology, A.A. Thacr & M. Sernetz (Editors), Springer-Verlag, Berlin-New York-Heidelberg, pp. 395-407, 1973.
24. McMahon, J., Kugel, R.B., Bartram, J.H., Berenberg, W., Cavanaugh, J.J., Hanson, V., Kennell, J.H., Lorincz, A.E., Pearson, P.H., Scott, R.B., Scurletis, T.D., Taft, L.T. (Committee on Children With Handicaps, american Academy of Pediatrics): The Physician and the Deaf child. Pediatrics, 51:1100-01, No. 6, June, 1973.
25. Thompson, J.N., Finley, S.C., Lorincz, A.E., Finley, W.H.: Absence of -L-Iduronidase Activity in Various Tissues from Two Sibs Affected Presumably With Hurler-Scheie Compound Syndrome. Birth Defects: Original Article Series, Disorders of Connective Tissues, XI: 341-46, No. 6, 1975.
26. Hurst, R.E., Cezayirli, R.C., Lorincz, A.E.: Nature of the Glycosaminoglycanuria (Mucopolysacchariduria) in Brachycephalic "Shorter" Dwarf Cattle. Journal of Comparative Pathology, 85:481-86, 1975.
27. Lorincz, A.E., Montes, L.F.: Mucopolysaccharidosis (Type I Hurler-Scheie Compound). Journal of Cutaneous Pathology, 2:214-15, 1975.
28. Hurst, R.E., Settine, J.M., Lorincz, A.E.: A Method for the Quantitative Determination of Urinary Glycosaminoglycans. Clinica Chimica Acta, 70:427-32, 1976.
29. Hurst, R.E., Jennings, G.C., Lorincz, A.E.: Partition Techniques for Isolation and Fractionation of Urinary Glycosaminoglycans. Analytical Biochemistry, 79:502-12, 1977.
30. Lorincz, A.E.: The Mucopolysaccharidoses: Advances in Understanding and Treatment. Pediatric Annals, 7:64-98, 1978.
31. Lorincz, A.E.: Mucopolysaccharidoses. Medical Aspects of Mental Retardation, Charles H. Carter (Editor), Charles C. Thomas (Publisher), 2nd edition, 1978.
32. Lorincz, A.E.: Perspectives on Planning for Prevention of Mental Retardation. Planning for Services to Handicapped Persons: Community, Education, & Health, Magrab, P.R. & Elder, J.O. (Editors), P.H. Brooks (Publisher), Chapter 3, pp. 77-90, 1979.
33. Hurst, R.E., Settine, J.M., Floyd, W.M., Lorincz, A.E.: Glycosaminoglycan Excretion in Osteogenesis Imperfecta. Clinica Chimica Acta, 100:307-11, 1980.

BIBLIOGRAPHY (CONT'D)

34. Lorincz, A.E.: Learning Disabilities - A Pediatrician's Perspective. The Role of Vision in the Multidisciplinary Approach to Children with Learning Disabilities, Morton Davis & John C. Whitener (Editors), Charles C. Thomas (Publisher), Springfield, Illinois, Chapter 2, pp. 18-40, 1982.
35. Lorincz, A.E., Hurst, R.E., Kolodny, E.H.: The Early Laboratory Diagnosis of Mucopolysaccharidoses. Annals of Clinical and Laboratory Science, 12:258-66, 1982.
36. Lorincz, A.E.: Glycosaminoglycans in Heritable and Developmental Disorders of Bone. Glycosaminoglycans and Proteoglycans in Physiological and Pathological Processes of Body Systems, R.S. Varma and R. Varma (Editors), S. Karger AG (Publisher), Switzerland, pp. 264-75, 1982.
37. Lorincz, A.E., Kolodny, E.H.: The Early Diagnosis of Mucopolysaccharidoses. Proceedings of the 6th Congress of the International Association for the Scientific Study of Mental Deficiency, J.M. Berg (Editor), Volume II, 1983.
38. Lorincz, A.E.: Rapid Method for the Identification of Mycoplasma Organisms. Manual of Procedures for Applications of Nucleic Acid Probes and Monoclonal Antibodies in Human Disease, F. William Sunderman (Editor), Institute for Clinical Science, Inc. (Publisher), pp. 163-165, November 13-16, 1986.
39. Lorincz, A.E.: Rapid Fluorescence Technique for the Detection of Toxic Pulmonary Microorganisms, e.g. Legionella Pneumophila. Manual of Procedures for Clinical and Analytical Toxicology, F. William Sunderman (Editor), Institute for Clinical Science, Inc. (Publisher), pp. 129-131, November 12-15, 1987.
40. Reque, P.G., Lorincz, A.E.: Supravital Microscopic Fluorescent Technique for the Detection of Tinea capitis. Cutis, 42:111-114, August 1988.
41. Petcharuttana, Y., Cutter, G.R., Meeks, R.G., & Lorincz, A.E.: Fluorescence Microscopy of DFS-induced Morphologic Transformation in Unfixed, Cultured Cells. Journal of Oral Pathology and Medicine, 18:451-456, 1989.
42. Lorincz, A.E.: Supravital Microscopic Fluorescence Technique for the Detection of Malaria and Other Parasitic Organisms. Manual of Procedures on Laboratory Diagnosis of Diseases of the Liver, F. William Sunderman (Editor), Institute for Clinical Science, Inc. (Publisher), pp. 89-92, November 16-19, 1989.
43. Lorincz, A.E.: Rapid Screening for Fungal Organisms Utilizing Supravital Fluorescence Microscopy. Manual of Procedures on Laboratory Diagnosis of Metabolic Disorders With Emphasis on Diabetes Mellitus, F. William Sunderman (Editor), Institute for Clinical Science, Inc. (Publisher), pp. 191-193, November 12-15, 1992.
44. Lorincz, A.E., Kelly, D.R., Dobbins, G.C., Cardone, V.S., Fuchs, S.A., Schilleci, J.L.: Urinalysis: Current Status and Prospects for the Future. Annals of Clinical and Laboratory Science, Vol 29, No 23, pp.169-175

ABSTRACTS

1. Lorincz, A.E., Dorfman, A.: Occurrence of Urinary Acid Mucopolysaccharides in the Hurler Syndrome. American Journal of Diseases of Children, 94:552, June, 1957.
2. Lorincz, A.E.: Acid Mucopolysaccharides in the Hurler Syndrome. Federation Proceedings, 17:226, March, 1958.
3. Kroger, M., Warnick, A.C., Hentges, J.F., Lorincz, A.E.: Mucopolysaccharidosis in Snorter Dwarf Cattle. Journal of Animal Science, 19:1221, 1960.
4. Lorincz, A.E.: "Snorter" Dwarf Cattle: A Naturally Occurring Heretable Disorder of Acid Mucopolysaccharide Metabolism Which Resembles Hurler Syndrome. American Journal of Diseases of Children, 100:488, 1960.
5. Lorincz, A.E.: Urinary Acid Mucopolysaccharides in Hereditary Deforming Chondrodysplasia (Diaphysical Aclais). Federation Proceedings, 19:148, December, 1960.
6. Lorincz, A.E.: Urinary Acid Mucopolysaccharides in Hereditary Arthrodysplasia. Southern Medical Journal, 53:1588, March, 1960.
7. Wolfson, S.L., Kahana, L., Harris, J.S., Lorincz, A.E.: Biochemical and Roctgenographic Response to Long-term Corticosteroid Therapy on the Hurler Syndrome. American Journal of Diseases of Children, 102:638, 1961.
8. Lorincz, A.E.: Mucopolysacchariduria in Children With Hereditary Arthro-osteo-onychodysplasia. Federation Proceedings, 21:173, March/April, 1962.
9. Schiebler, G.L., Lorincz, A.E., Brodgon, B.G., Shanklin, D.R., Bartley, T.D., Krovetz, L.J.: Cardiovascular Manifestations of Hurler's Syndrome. Circulation, 26:782, 1962.
10. Gessner, L.H., Lorincz, A.E.: Acid Mucopolysaccharide Content of the Three-day Old Chick Embryo Heart. Federation Proceedings, 22:2, December, 1963.
11. Krovetz, L.J., Lorincz, A.E., Schiebler, G.L.: Hemodynamic Studies in the Hunter-Hurler Syndrome (Gargoylism). Circulation, 28:753., 1963.
12. Batey, R.L., Lorincz, A.E.: A New Technique for the Identification of Acid Mucopolysaccharide Hydrolysates. Southern Medical Journal, 56:1437, 1963.
13. Callahan, W.P., Lorincz, A.E.: Changes in Liver Ultrastructure Associated With Hurler Syndrome. Anatomical Record, 148:267, 1964.
14. Renn, W.H., Lorincz, A.E.: A New Urinary Mucopolysaccharide Associated With the Nail-Patella Syndrome (Hereditary arthro-osteo-onychodydysplasia). Southern Medical Journal, 58:1584, December, 1965.
15. Lorincz, A.E., Callahan, W.P.: Electron Microscopic Observations of Liver in the Hurler Syndrome. Southern Medical Journal, 58:1584, December, 1965.
16. Renn, W.H., Enneking, W.F., Hase, M.F., Lorincz, A.E.: Acid Mucopolysaccharide Content of Human Chondrosarcomatous Tumors. Journal of Bone and Joint Surgery, 50-A:1072, 1968.
17. Lorincz, A.E., Tiller, R.E., West, S.S.: A New Biophysical Cytochemical Finding in Viable Leukocytes Obtained from Patients with Cystic Fibrosis. Cystic Fibrosis Club Abstracts, 13th Annual Meeting, May 20, 1972.
18. Lorincz, A.E., Finley, S.C., Finley, W.H., West, S.S.: Biophysical Cytochemical Study of Glycosaminoglycans in Fibroblasts and Other Cells Derived From Marteaux-Lamy Mucopolysaccharidosis. Federation Proceedings, Vol. 31, No. 2, March-April, 1973.

ABSTRACTS (CONT'D)

19. Warren, R.J., Condrón, C.J., Hollister, D., Huifing, F., Neufeld, E.F., Hall, C.W., Mcleod, A.G.W., Lorincz, A.E.: Antenatal Diagnosis of Mucopoliddosis II (I-Cell Disease). Pediatric Research, Vol. 7, No. 343, April, 1973.
20. Lorincz, A.E., West, S.S.: The Biophysical Cytochemistry of Living Cells Derived From Patients With Mucopolysaccharidoses. Journal of Bone and Joint Surgery: Proceedings of the 19th Annual Meeting of the Orthopedic Research Society, Las Vegas, Nevada, 55-A:655, January 30, 31 & February 1, 1973.
21. Hurst, R.E., Cezayirli, R.C., Lorincz, A.E.: The Nature of the Glycosaminoglycans (Mucoploysacchariduria) in Brachycephalic "Snorter" Dwarf Cattle. Federation Proceedings, 34:861, March, 1975.
22. Lorincz, A.E., Hurst, R.E., Cezayirli, R.C.: Definitive Characterization of the Glycosaminoglycanuria in "Snorter" Dwarf Cattle. Pediatric Research, 9:314, 1975.
23. Lorincz, A.E., Hurst, R.E., Floyd, W.M.: Glycosaminoglycan excretion of Osteogenesis Imperfecta. Transactions of the 25th Annual Meeting, Orthopedic Research Society: Journal of Bone and Joint Surgery, 4:108, 1978.
24. Lorincz, A.E.: Clinical Chemistry in the Profoundly Retarded. Annals of Clinical and Laboratory Science, 14:316-17, 1984.
25. Lorincz, A.E.: Direct Visualization of Mycoplasma via Supravital Staining and Fluorescence Microscopy. Abstracts of the 6th International Congress of the International Organization for Mycoplasma, p. 272, 1986.
26. Lorincz, A.E., Thompson, J.N.: Use of Microscopic Fluorescence Amniotic Fluid to Diagnose Intrauterine Mucopolysaccharidosis. 1st International Congress on Mucopolysaccharidosis and Related Diseases, p. 33, 1988.
27. Lorincz, A.E., Reque, P.G.: Comparison of Supravital Microscopic Fluorescence Technique (SMFT) to Darkfield Microscopy for Detection of Spirochetes. Annals of Clinical and Laboratory Science, 20:281-82, 1990.
28. Lorincz, A.E.: One Step On-site Epi-fluorescence Detection of Fungi: A Possible Alternate to KOH Screening. Annals of Clinical and Laboratory Science, 23: 307, 1993.
29. Lorincz, A.E. and Bueschen, A.J.: Mycoplasma-like Mollicutes in Human Prostate Tumor Tissue. Annals of Clinical and Laboratory Science, 24: 196, 1994.
30. Lorincz, A.E., Baltaro, R.J. and Adamson, D.M.: Detection of Significant Bacteruria Using Supravital Fluorescence Microscopy. Annals of Clinical and Laboratory Science, 25: 363, 1995.
31. Lorincz, A.E., Bueschen, A.J., and Urban, D.A.: Presence of Mycoplasma-Like Mollicutes In Prostate Needle Biopsy Specimens, Annals of Clinical and Laboratory Science, 26:364, 1996.

EDITORIALS AND COMMENTARIES

1. Lorincz, A.E.: Biochemical Genetic Defects. Journal of the Florida Medical Association, 66:852, 1960.
2. Lorincz, A.E., Smith, R.T.: Research at the University of Florida College of Medicine, Department of Pediatrics. Journal of the Florida Medical Association, 51:808-11, 1965.
3. Lorincz, A.E.: Birth Defects - A Continuing Challenge. Journal of the Florida Medical Association, 55:126-27, 1968.

BOOK REVIEWS

1. "Modern Topics in Paediatric Dermatology", Verbov, J. (Editor), J.B. Lippincott, Philadelphia (Publisher), JAMA, 244:1269, 1980.
2. "Dermatologic Disorders in Black Children and Adolescents", Laude, T.A. & Russo, R.U. (Editors), Medical Examination Publishing Company, New Hyde Park, New York (Publisher), JAMA, 252:1769-70, 1984.
3. "Practical Pediatric Dermatology", Weston, W.L. (Editor), Little Brown and Company, Boston (Publisher), JAMA, 277:549, 1987.

**EXHIBIT
B**

DISCLOSURES

**APPLICATION
SERIAL NO. 09/836,750**

EXHIBIT B
DISCLOSURES
APPLICATION SERIAL NO. 09/836,750

PAGE 20, LINE 10 – PAGE 21, LINE 15

Growth factors can be utilized to induce the growth of “hard tissue” or bone and “soft tissues” like ectodermal and mesodermal tissues. As used herein, the term growth factor encompasses compositions and living organisms which promote the growth of hard tissue, such as bone, or soft tissue, in the body of a patient. The compositions include organic and inorganic matter. The compositions can be genetically produced or manipulated. The living organisms can be bacteria, viruses, or any other living organism which promote tissue growth. By way of example and not limitation, growth factors can include platelet-derived growth factor (PDGF), epidermal growth factor (EGF), fibroblast growth factor (acidic/basic (FGF a,b), interleukins (IL's), tumor necrosis factor (TNF), transforming growth factor (TGF-B), colony-stimulating factor (CSF), osteopontin (Eta-1 OPN), platelet-derived growth factor (PDGF), interferon (INF), bone morphogenic protein 1 (BMP-1), and insulin growth factor (IGF). Recombinant and non-recombinant growth factors can be utilized as desired. Bacteria or viruses can, when appropriate, be utilized as growth factors. For example, there is a bacterial hydrophilic polypeptide that self-assembles into a nanometer internal diameter pore to build a selective lipid body. Various enzymes can be utilized for the synthesis of peptides which contain amino acids that control three-dimensional protein structure and growth. Growth factors can be applied in gels or other carriers which regulate the rate of release of the growth factors and help maintain the growth factors and the carrier, at a desired location in the body. Time release capsules, granules, or other carriers containing growth factor can be activated by tissue pH, by enzymes, by ultrasound,

by electricity, by heat, by selected *in vivo* chemicals or by any other selected means to release the growth factor. The carrier can be resorbable or non-resorbable. Or, the growth factor itself can be activated by similar means. Either the carrier or the growth factor can mimic extracellular fluid to control cell growth, migration, and function. The growth factor can be administered orally, systemically, in a carrier, by hypodermic needle, through the respiratory tract, or by any other desired method. The growth factor can also be administered into a capsule or other man-made composition or structure placed in the body. While administration of the growth factor is presently usually localized in the patient's body, circumstances may arise where it is advantageous to distribute a growth factor throughout the patient's body in uniform or non-uniform concentrations. An advantage to growth factors is that they can often, especially when in capsule form or in some other containment system, be inserted to a desired site in the body by simply making a small incision and inserting the growth factor. The making of such small incision comprises minor surgery which can often be accomplished on an out-patient basis. The growth factors can be multifactorial and nonspecific.

PAGE 44, LINE 19 – PAGE 46, LINE 16

Genetic material comprising a portion of a gene, a gene, genes, a gene product (i.e., a composition a gene causes to be produced like, for example, an organ-producing growth factor), growth factor, or an ECM (extracellular matrix) can be used in or on the body to grow an organ to tissue. For example, the vascular epithelial growth factor gene (VEGF) or its growth factor equivalent can be inserted into the body to cause an artery to grow. When insertion of a gene, portion of a gene, gene product, growth factor, or ECM *in vivo* or *ex vivo* is referred to herein in connection with any of the implant techniques of the invention, it is understood that a cell

nutrient culture(s), physiological nutrient culture(s), carrier (s), enhancer(s), promoter(s), or any other desired auxiliary component(s) can be inserted with the gene or at the same location as the gene, growth factor, ECM, etc.

An artery is an organ from the circulatory system. An artery can be grown in the heart, legs, or other areas by injecting a gene or other genetic material into muscle at a desired site. Size, vascularity, simplicity of access, ease of exploitation, and any other desired factors can be utilized in selecting a desired site. The gene is one of several known VEGF genes which cause the production of vascular endothelial growth factors. Several VEGF genes which produce vascular endothelial growth factors are believed to exist because nature intends for there to be several pathways (i.e., genes) which enable the production of necessary growth factors. The existence of several pathways is believed important because if one of the genes is damaged or inoperative, other similar genes can still orchestrate the production of necessary growth factors. VEGF genes are used by the body to promote blood vessel growth. VEGF genes are assimilated (taken in) by muscle cells. The genes cause the muscle cells to make a VEGF protein which promotes the growth of new arteries. VEGF proteins can be made in a lab and injected into a patient intravenously, intraluminally, or intramuscularly to promote the growth of an artery. Or, the genes (or other genetic material) can be applied with an angioplasty balloon, with the assistance of a vector, or by any other method.

It is not always desirable to grow a completely new organ. Sometimes growing a portion of an organ is desirable. For example, in some heart attacks or strokes, a portion of the heart or brain remains viable and a portion dies. An injection of a gene to form cardiac muscle and/or an injection of a gene to form an artery can be utilized to revive or replace the dead portion of the heart. The dead portion of the heart may (or may not) be used as a matrix while the new muscles

and vessels grow. Thus, in this example, a partial new organ is grown in a pre-existing organ. A pacemaker may (or may not) be necessary. A second injection of a gene may (or may not) be necessary to stop cardiac muscle growth once it is completed. Portions of organs throughout the body can similarly be repaired or replaced. It may be necessary to provide gene(s) or growth factor(s) sequentially. For instance, one or more blood vessels are grown by inserting an appropriate gene or other genetic material into a selected area. Second, an appropriate gene or other genetic material is inserted in the selected area to grow a bone or other organ.

The size and shape limitation of the desired structure can come from a containment and boundary contact inhibition phenomenon or by a chemical inhibition.

A variation on the theme of growing a portion of an organ is as follows: a portion of a heart dies. The pericardium is utilized as a scaffold and seeded with cells and/or genes to grow new muscle, and genes (or other genetic material) to grow new arteries. Immediately adjacent the dead cardiac muscle, onto or into the pericardium, the appropriate cells, genes, and/or growth factors (or other genetic material) are placed. Once the new muscle and blood vessels have grown, the function specific tissue can be applied to the damaged portion of the heart and paced, if necessary, to augment cardiac action. If the surgeon desires, the dead muscle can be removed and the new muscle and blood vessels can be surgically rotated into the excised region and secured. This probably can be done endoscopically. In essence, the pericardium is utilized to allow the new muscle wall to grow. The new muscle wall is then transplanted into the damaged heart wall. This procedure utilizes the body as a factor to grow an organ and/or tissue, after which the organ and/or tissue is transplanted to a desired region. On the other hand, the new muscle wall may integrate itself into the old wall and not require transplantation.

EXHIBIT C

DEFINITIONS

EXHIBIT C

DEFINITIONS

PAGE 20, LINE 10 – PAGE 21, LINE 15

Growth factors can be utilized to induce the growth of “hard tissue” or bone and “soft tissues” like ectodermal and mesodermal tissues. As used herein, the term growth factor encompasses compositions and living organisms which promote the growth of hard tissue, such as bone, or soft tissue, in the body of a patient. The compositions include organic and inorganic matter. The compositions can be genetically produced or manipulated. The living organisms can be bacteria, viruses, or any other living organism which promote tissue growth. By way of example and not limitation, growth factors can include platelet-derived growth factor (PDGF), epidermal growth factor (EGF), fibroblast growth factor (acidic/basic (FGF a,b), interleukins (IL's), tumor necrosis factor (TNF), transforming growth factor (TGF-B), colony-stimulating factor (CSF), osteopontin (Eta-1 OPN), platelet-derived growth factor (PDGF), interferon (INF), bone morphogenic protein 1 (BMP-1), and insulin growth factor (IGF). Recombinant and non-recombinant growth factors can be utilized as desired. Bacteria or viruses can, when appropriate, be utilized as growth factors. For example, there is a bacterial hydrophilic polypeptide that self-assembles into a nanometer internal diameter pore to build a selective lipid body. Various enzymes can be utilized for the synthesis of peptides which contain amino acids that control three-dimensional protein structure and growth. Growth factors can be applied in gels or other carriers which regulate the rate of release of the growth factors and help maintain the growth factors and the carrier, at a desired location in the body. Time release capsules, granules, or other carriers containing growth factor can be activated by tissue pH, by enzymes, by ultrasound,

by electricity, by heat, by selected *in vivo* chemicals or by any other selected means to release the growth factor. The carrier can be resorbable or non-resorbable. Or, the growth factor itself can be activated by similar means. Either the carrier or the growth factor can mimic extracellular fluid to control cell growth, migration, and function. The growth factor can be administered orally, systemically, in a carrier, by hypodermic needle, through the respiratory tract, or by any other desired method. The growth factor can also be administered into a capsule or other man-made composition or structure placed in the body. While administration of the growth factor is presently usually localized in the patient's body, circumstances may arise where it is advantageous to distribute a growth factor throughout the patient's body in uniform or non-uniform concentrations. An advantage to growth factors is that they can often, especially when in capsule form or in some other containment system, be inserted to a desired site in the body by simply making a small incision and inserting the growth factor. The making of such small incision comprises minor surgery which can often be accomplished on an out-patient basis. The growth factors can be multifactorial and nonspecific.

MEDLINE PLUS: MERRIAM-WEBSTER MEDICAL DICTIONARY
A SERVICE OF THE U.S. NATIONAL LIBRARY OF MEDICINE AND THE
NATIONAL INSTITUTES OF HEALTH

Growth factor: a substance (as a vitamin B₁₂ or an interleukin)
that promotes growth and especially cellular growth

EXHIBIT D

CLAIMS

EXHIBIT D

CLAIMS

Claim X: A method for growing a new portion of a pre-existing heart comprising the steps of: placing a growth factor in a body of a human patient and growing new muscle in said heart.

EXHIBIT E

PUBLICATIONS

EXHIBIT E

PUBLICATION INFORMATION SUMMARY

TITLE	AUTHOR	CITATION	DATE	AUTHOR COUNTRY	ROUTE OF ADMINISTRATION	GROWTH FACTOR ADMINISTERED	RESULT
Left Ventricular Electromechanical Mapping to Assess Efficacy of phVEGF165 Gene Transfer for Therapeutic Angiogenesis in Chronic Myocardial Ischemia	Vale	Circulation. 2000; 102:965-974	08/29/00	U.S.	Small incision (minithoracotomy) with syringe injection	VEGF (Gene form)	Repair of damaged portion of heart -- Also pertains to new muscle growth
Repair of Infarcted Myocardium by Autologous Intracoronary Mononuclear Bone Marrow Cell Transplantation in Humans	Strauer	Circulation. 2002; 106:1913-1918	10/08/02	Germany	Balloon catheter with injection	Bone Marrow Cells	Repair of dead portion of heart -- also pertains to new muscle growth

TITLE	AUTHOR	CITATION	DATE	AUTHOR COUNTRY	ROUTE OF ADMINISTRATION	GROWTH FACTOR ADMINISTERED	RESULT
Viability and differentiation of autologous skeletal myoblast grafts in ischemic cardiomyopathy	Hagege	Lancet 2003 Feb 8; 361 (9356):491-492	2003	France	Injection	Skeletal Muscle Cells	Repair of dead portion of heart; Histological Proof (muscle)
Autologous Cell Transplant Helpful in Ischemic Heart or Legs	Barclay	Medscape Medical News 2000 – Abstract from American Heart Association's 75 th Scientific Sessions on 11/18/02, Chicago	11/18/02	U.S.	Surgery with syringe injection	Bone Marrow Cells	Repair of damaged portion of heart – also pertains to new muscle growth
Autologous skeletal myoblasts transplanted to ischemia-damaged myocardium in humans. Histological analysis of cell survival and differentiation	Pagani	J Am Coll Cardiol 2003 Mar 5; 41(5): 879-888	2003	U.S.	Surgery with syringe injection	Skeletal Muscle Cells	Repair of dead portion of heart; Histological Proof (muscle and blood vessels)

Repair of Infarcted Myocardium by Autologous Intracoronary Mononuclear Bone Marrow Cell Transplantation in Humans

Bodo E. Strauer, MD; Michael Brehm, MD; Tobias Zeus, MD; Matthias Köstering, MD; Anna Hernandez, PhD; Rüdiger V. Sorg, PhD; Gesine Kögler, PhD; Peter Wernet, MD

Background—Experimental data suggest that bone marrow–derived cells may contribute to the healing of myocardial infarction (MI). For this reason, we analyzed 10 patients who were treated by intracoronary transplantation of autologous, mononuclear bone marrow cells (BMCs) in addition to standard therapy after MI.

Methods and Results—After standard therapy for acute MI, 10 patients were transplanted with autologous mononuclear BMCs via a balloon catheter placed into the infarct-related artery during balloon dilatation (percutaneous transluminal coronary angioplasty). Another 10 patients with acute MI were treated by standard therapy alone. After 3 months of follow-up, the infarct region (determined by left ventriculography) had decreased significantly within the cell therapy group (from 30 ± 13 to $12 \pm 7\%$, $P=0.005$) and was also significantly smaller compared with the standard therapy group ($P=0.04$). Likewise, infarction wall movement velocity increased significantly only in the cell therapy group (from 2.0 ± 1.1 to 4.0 ± 2.6 cm/s, $P=0.028$). Further cardiac examinations (dobutamine stress echocardiography, radionuclide ventriculography, and catheterization of the right heart) were performed for the cell therapy group and showed significant improvement in stroke volume index, left ventricular end-systolic volume and contractility (ratio of systolic pressure and end-systolic volume), and myocardial perfusion of the infarct region.

Conclusions—These results demonstrate for the first time that selective intracoronary transplantation of autologous, mononuclear BMCs is safe and seems to be effective under clinical conditions. The marked therapeutic effect may be attributed to BMC-associated myocardial regeneration and neovascularization. (*Circulation*. 2002;106:1913-1918.)

Key Words: myocardial infarction ■ cell transplantation, intracoronary ■ angiogenesis ■ bone marrow ■ myogenesis

Remodeling of the left ventricle after myocardial infarction (MI) represents a major cause of infarct-related heart failure and death. This process depends on acute and chronic transformation of both the necrotic infarct region and the non-necrotic, peri-infarct tissue.^{1,2} Despite application of pharmacotherapeutics and mechanical interventions, the cardiomyocytes lost during MI cannot be regenerated. The recent finding that a small population of cardiac muscle cells is able to replicate itself is encouraging but is still consistent with the concept that such regeneration is restricted to viable myocardium.³

In animal experiments, attempts to replace the necrotic zone by transplanting other cells (eg, fetal cardiomyocytes or skeletal myoblasts) have invariably succeeded in reconstituting heart muscle structures, ie, myocardium and coronary vessels. However, these cells fail to integrate structurally and do not display characteristic physiological functions.⁴⁻⁷ Another approach to reverse myocardial remodeling is to repair myocardial tissue by using bone marrow–derived cells. Bone

marrow contains multipotent adult stem cells that show a high capacity for differentiation.⁸⁻¹⁰ Experimental studies have shown that bone marrow cells (BMCs) are capable of regenerating infarcted myocardium and inducing myogenesis and angiogenesis; this leads in turn to amelioration of cardiac function in mice and pigs.¹¹⁻¹⁴ However, procedures based on this phenomenon remain largely uninvestigated in a human clinical setting.

An investigation of one patient receiving autologous skeletal myoblasts into a postinfarction scar during coronary artery bypass grafting revealed improvement of contraction and viability 5 months afterward.¹⁵ Autologous mononuclear BMCs transplanted in a similar surgical setting showed long-term improvement of myocardial perfusion in 3 of 5 patients and no change in 2 patients.¹⁶ However, such studies entail a surgical approach and are therefore associated with well-known perioperative risks. Moreover, this surgical procedure cannot be used with MI. We therefore looked for a nonsurgical, safer mode for transplanting autologous cells

Received August 2, 2002; accepted August 2, 2002.

From the Department of Medicine, Division of Cardiology (B.E.S., M.B., T.Z., M.K.) and Institute for Transplantation Diagnostics and Cell Therapeutics (A.H., R.V.S., G.K., P.W.), Heinrich-Heine-University of Düsseldorf, Germany.

Correspondence to Professor Dr Bodo E. Strauer, Department of Medicine, Division of Cardiology, Heinrich-Heine-University, Moorenstr 5, 40225 Düsseldorf, Germany. E-mail: Strauer@med.uni-duesseldorf.de

© 2002 American Heart Association, Inc.

Circulation is available at <http://www.circulationaha.org>

DOI: 10.1161/01.CIR.0000034046.87607.1C

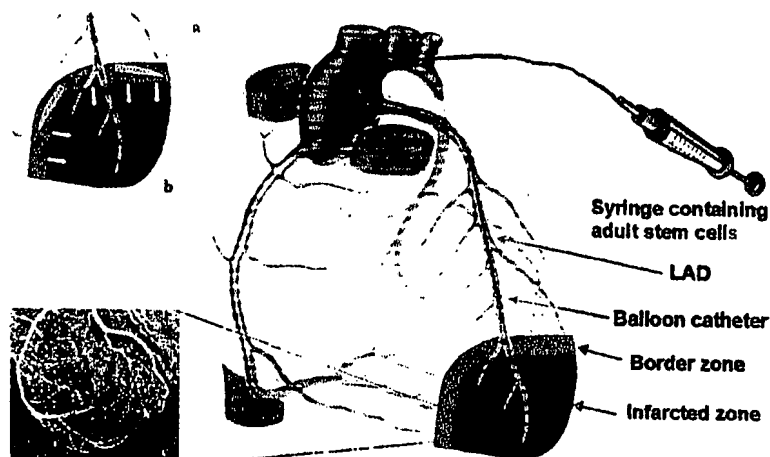


Figure 1. Procedure of cell transplantation into infarcted myocardium in humans. *a*, The balloon catheter enters the infarct-related artery and is placed above the border zone of the infarction. It is then inflated and the cell suspension is infused at high pressure under stop-flow conditions. *b*, In this way, cells are transplanted into the infarcted zone via the infarct-related vasculature (red dots). Cells infiltrate the infarcted zone. Blue and white arrows suggest the possible route of migration. *c*, A supply of blood flow exists within the infarcted zone.³⁵ The cells are therefore able to reach both the border and the infarcted zone.

into postinfarction tissue. A pilot study from our group demonstrated that intracoronary transplantation of autologous mononuclear BMCs 6 days after MI was associated with a marked decrease in infarct area and an increase in left ventricular (LV) function after 3 and 6 months of follow-up.¹⁷ To confirm these results and validate this promising new therapy for MI, we established a clinical trial involving 20 patients for comparing the safety and bioefficacy of autologous BMC transplantation. All 20 patients underwent standard therapy, and 10 patients received additional intracoronary cell transplantation. All 20 patients were followed up for 3 months.

Methods

Patient Population

All 20 patients had suffered transmural infarction according to World Health Organization criteria with the involvement of the left anterior descending coronary artery ($n=4$), left circumflex coronary artery ($n=3$), or right coronary artery ($n=13$). Mean duration of infarct pain was 12 ± 10 hours before invasive diagnostics and therapy. Patients had to be <70 years old and were excluded if one of the following criteria were met: screening >72 hours after infarction, cardiac shock, severe comorbidity, alcohol or drug dependency, or excessive travel distance to the study center.

After right and left heart catheterization, coronary angiography, and left ventriculography, mechanical treatment was initiated with recanalization of the infarct-related artery by balloon angioplasty ($n=20$) and subsequent stent implantation ($n=19$). All patients were monitored in our intensive care unit, and no arrhythmogenic events or hemodynamic impairments were recorded in either patient group.

All 20 patients were briefed in detail about the procedure of BMC transplantation. Informed consent was obtained from 10 patients, who formed the cell therapy group, whereas 10 patients who refused additional cell therapy served as controls. The local ethics committee of the Heinrich-Heine-University, Düsseldorf, approved the study protocol. All procedures conformed to institutional guidelines.

Before taking part in rehabilitation programs, all patients left the hospital with standard medication consisting of acetylsalicylic acid, an ACE inhibitor, a β -blocker, and a statin.

Bone Marrow Aspiration, Isolation, and Cultivation

Seven (± 2) days after acute coronary angiography, bone marrow (~ 40 mL) was aspirated under local anesthesia from ilium of cell therapy patients ($n=10$). Mononuclear BMCs were isolated by Ficoll density separation on Lymphocyte Separation Medium (BioWhittaker) before the erythrocytes were lysed with H_2O . For overnight

cultivation, 1×10^6 BMCs/mL were placed in Teflon bags (Vuelife, Cell Genix) and cultivated in X-Vivo 15 Medium (BioWhittaker) supplemented with 2% heat-inactivated autologous plasma. The next day, BMCs were harvested and washed 3 times with heparinized saline before final resuspension in heparinized saline. Viability was $93 \pm 3\%$. Heparinization and filtration (cell strainer, FALCON) was carried out to prevent cell clotting and microembolization during intracoronary transplantation. The mean number of mononuclear cells harvested after overnight culture was 2.8×10^7 ; this consisted of $0.65 \pm 0.4\%$ AC133-positive cells and $2.1 \pm 0.28\%$ CD34-positive cells. All microbiological tests of the clinically used cell preparations proved negative. As a viability and quality ex vivo control, 1×10^5 cells grown in H5100 medium (Stem Cell Technology) were found to be able to generate mesenchymal cells in culture.

Intracoronary Transplantation of BMCs

Five to nine days after onset of acute infarction, cells were directly transplanted into the infarcted zone (Figure 1). This was accomplished with the use of a balloon catheter, which was placed within the infarct-related artery. After exact positioning of the balloon at the site of the former infarct-vessel occlusion, percutaneous transluminal coronary angioplasty (PTCA) was performed 6 to 7 times for 2 to 4 minutes each. During this time, intracoronary cell transplantation via the balloon catheter was performed, using 6 to 7 fractional high-pressure infusions of 2 to 3 mL cell suspension, each of which contained 1.5 to 4×10^6 mononuclear cells. PTCA thoroughly prevented the backflow of cells and at the same time produced a stop-flow beyond the site of the balloon inflation to facilitate high-pressure infusion of cells into the infarcted zone. Thus, prolonged contact time for cellular migration was allowed.¹⁸

Functional Assessment of Hemodynamics

After 3 months, all 20 patients were followed up by left heart catheterization, left ventriculography, and coronary angiography. Ejection fraction, infarct region, and regional wall movement of the infarcted zone during ejection were determined by left ventriculography. Ejection fraction was measured with Quantcor software (Siemens). To quantify infarction wall movement velocity, 5 axes were placed perpendicular to the long axis in the main akinetic or dyskinetic segment of the ventricular wall. Relative systolic and diastolic lengths were measured, and the mean difference was divided by the systolic duration (in seconds). To quantify the infarct region, the centerline method according to Sheehan was used.¹⁹ All hemodynamic investigations were obtained by two independent observers.

In the cell therapy group before and 3 months after cell transplantation, additional examinations for measuring hemodynamics and myocardial perfusion included dobutamine stress echocardiography, radionuclide ventriculography, catheterization of the right heart, and

TABLE 1. Baseline Characteristics of the Patients

Clinical Data	Cell Therapy	Standard Therapy	P
Characteristics			
No. of patients	10	10	...
Age, y	49±10	50±6	NS
Sex	Male	Male	...
Onset of infarction before angioplasty, h	10±8	13±11	NS
Coronary angiography			
No. of diseased vessels	1.7±0.9	2.1±0.7	NS
No. of patients with LAD/LCX/RCA as the affected vessel	4/1/5	0/2/8	...
No. of patients with stent implantation	9	10	...
Laboratory parameters			
Creatinine kinase, U/L	1138±1170	1308±1187	NS
Creatinine kinase-MB, U/L	106±72	124±92	NS
Bone marrow puncture after angioplasty, d	7±2
Mononuclear bone marrow cells, n (×10 ⁷)	2.8±2.2

Values are mean±SD or number of patients. NS indicates not significant; LAD, left anterior descending coronary artery; LCX, left circumflex coronary artery; and RCA, right coronary artery.

stress-redistribution-reinjection ²⁰¹thallium scintigraphy. The contractility index P_{mv}/ESV was calculated by dividing LV systolic pressure (P_{mv}) by end-systolic volume (ESV). Perfusion defect was calculated by scintigraphic bull's-eye technique. Each examination was performed according to standard protocols.

There were no complications or side effects determined in any patient throughout the diagnostic or therapeutic procedure or within the 3-month follow-up period.

Statistical Analysis

All data are presented as mean±SD. Statistical significance was accepted when P was <0.05. Discrete variables were compared as rates, and comparisons were made by χ^2 analysis. Intra-individual comparison of baseline versus follow-up continuous variables was performed with a paired t test. Comparison of nonparametric data between the two groups was performed with Wilcoxon test and Mann-Whitney test. Statistical analysis was performed with SPSS for Windows (version 10.1).

Results

Clinical data between the two groups did not differ significantly. The range of creatinine kinase levels was slightly but not significantly higher in the standard therapy group than it was in the cell therapy group (Table 1).

Comparison of the 2 groups 3 months after cell or standard therapy showed several significant differences in LV dynamics, according to the global and regional analysis of left ventriculogram. The infarct region as a percentage of hypokinetic, akinetic, or dyskinetic segments of the circumference of the left ventricle decreased significantly in the cell therapy group (from 30±13 to 12±7%, $P=0.005$). It was also significantly smaller compared with the standard therapy group after 3 months ($P=0.04$). Within the standard therapy group, only a statistically nonsignificant decrease from 25±8 to 20±11% could be seen. Wall movement velocity over the infarct region rose significantly in the cell therapy group (from 2.0±1.1 to 4.0±2.6 cm/s, $P=0.028$) but not in the standard therapy group (from 1.8±1.3 to 2.3±1.6 cm/s, $P=NS$). No significant difference was observed between the

two groups. Ejection fraction increased in both groups, albeit nonsignificantly (from 57±8 to 62±10% in the cell therapy group and from 60±7 to 64±7% in the standard therapy group) (Table 2).

Further significant improvement could also be seen on additional analysis of the cell therapy group alone. Perfusion defect was considerably decreased by 26% in the cell therapy group (from 174±99 to 128±71 cm², $P=0.016$, assessed by ²⁰¹thallium scintigraphy) (Figure 2). Parallel to the reduction in perfusion defect, improvement (Table 3) could also be seen in:

- (1) Cardiac function, as revealed by increase in stroke volume index (from 49±7 to 56±7 mL/m², $P=0.010$) and ejection fraction (from 51±14 to 53±13%, $P=NS$).
- (2) Cardiac geometry, as shown by decreases in both end-diastolic (from 158±20 to 143±30 mL, $P=NS$) and end-systolic volume (from 82±26 to 67±21 mL, $P=0.011$). Radionuclide ventriculography was used to acquire the data.
- (3) Contractility as evaluated by an increase in the velocity of circumferential fiber shortening (from 20.5±4.2 to 24.4±7.7 mm/s, $P=NS$, assessed by stress echocardiography) and by a marked increase in the ratio of systolic pressure to end-systolic volume (from 1.81±1.44 to 2.27±1.72 mm Hg/mL, $P=0.005$).

Discussion

The present report describes the first clinical trial of intracoronary, autologous, mononuclear BMC transplantation for improving heart function and myocardial perfusion in patients after acute MI. The results demonstrate that transplanted autologous BMCs may lead to repair of infarcted tissue when applied during the immediate postinfarction period. These results also show that the intracoronary approach of BMC transplantation seems to represent a novel

TABLE 2. Comparison of Cell Therapy and Standard Therapy Groups

	Cell Therapy	Standard Therapy	P
No. of patients	10	10	...
Infarct region as functional defect			
Hypokinetic, akinetic, or dyskinetic region at 0 mo, %	30±13	25±8	NS
Hypokinetic, akinetic, or dyskinetic region at 3 mo, %	12±7	20±11	0.04
P	0.005	NS	...
Contractility indices			
Infarction wall movement velocity at 0 mo, cm/s	2.0±1.1	1.8±1.3	NS
Infarction wall movement velocity at 3 mo, cm/s	4.0±2.6	2.3±1.6	NS
P	0.028	NS	...
Hemodynamic data			
LV ejection fraction at 0 mo, %	57±8	60±7	NS
LV ejection fraction at 3 mo, %	62±10	64±7	NS
P	NS	NS	...

NS indicates not significant; 0 mo, zero months, which means the time of infarction; 3 mo, 3 months, which means the time of the follow-up examinations. All data were obtained according to analysis of left ventriculogram.

and effective therapeutic procedure for concentrating and/or depositing infused cells within the region of interest.

Neogenesis of both cardiomyocytes and coronary capillaries with some functional improvement has been shown recently by several investigators using bone marrow-derived cells in experimental infarction.^{11–14,18,20–23} Moreover, trans-endothelial migration from the coronary capillaries and incorporation of cells into heart muscle has been observed experimentally.^{3,12,24–26} Until now, clinical data only existed for the cell therapy of surgically treated chronic ischemic heart disease.^{15,16} Our aim was to transform the encouraging results from animal models to a safe clinical setting. The most crucial questions we had to address while designing and

realizing this trial were: (1) What cell population should we deliver? (2) Which application method is the most efficient? (3) When should the cells be transplanted?

In recent years, several laboratories have shown that environmentally dictated changes of fate (transdetermination) are not restricted to stem cells but may also involve progenitor cells at different steps of a given differentiation pathway (transdifferentiation). Moreover, mesenchymal stem cells may represent an ideal cell source for treating different diseases.²⁷ Adult, mononuclear BMCs contain such stem and progenitor cells (≤1%), eg, mesodermal progenitor cells, hematopoietic progenitor cells, and endothelial progenitor cells. In several animal infarction models it has been shown that: (1) Bone marrow hemangioblasts contribute to the formation of new vessels; (2) bone marrow hematopoietic stem cells differentiate into cardiomyocytes, endothelium,

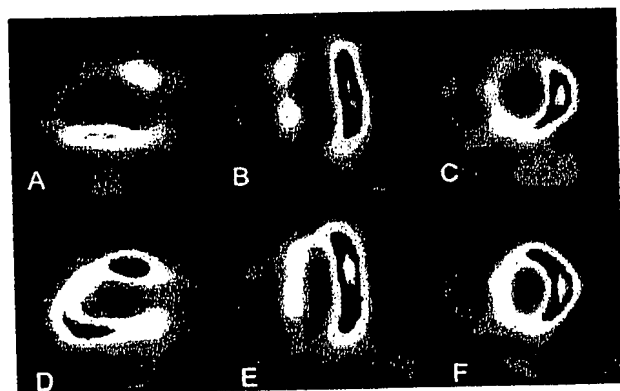


Figure 2. Improved myocardial perfusion of infarcted anterior wall 3 months after intracoronary cell transplantation subsequent to an acute anterior wall infarction detected by ²⁰¹thallium scintigraphy. The images on the left (A, D, sagittal) and in the middle (B, E) show the long axis, whereas those on the right (C, F, frontal) show the short axis of the heart. Initially the anterior wall, with green-colored apical and anterior regions, had reduced myocardial perfusion (A, B, C). Three months after cell transplantation the same anterior wall, now yellow in color, revealed a significant improvement in myocardial perfusion (D, E, F). All illustrations depict the exercise phase.

TABLE 3. Cardiac Function Analysis at 3-Month Follow-Up

	Before Cell Therapy	3 Months After Cell Therapy	P
No. of patients	10	10	...
Hemodynamic data			
LV ejection fraction, %	51±14	53±13	NS
Stroke volume index, mL/m ²	49±7	56±7	0.010
Cardiac geometry			
LV end-diastolic volume, mL	158±20	143±30	NS
LV end-systolic volume, mL	82±26	67±21	0.011
Contractility indices			
Circumferential fiber shortening, mm/s	20.5±4.2	24.4±7.7	NS
P _{2ys} /ESV, mm Hg/mL	1.81±1.44	2.27±1.72	0.005
Infarct region as perfusion defect			
²⁰¹ Thallium scintigraphy, cm ²	174±99	128±71	0.016

NS indicates not significant.

and smooth muscle cells⁸⁻¹³; (3) BMCs give rise to mesodermal progenitor cells that differentiate to endothelial cells²⁸; and (4) endothelial progenitors can transdifferentiate into beating cardiomyocytes.²⁹ Thus, several different fractions of mononuclear BMCs may contribute to the regeneration of necrotic myocardium and vessels. In order to utilize this large and perhaps heterogeneous regenerative potential, we decided to use all mononuclear cells from the bone marrow aspirate as a whole, rather than a subpopulation. No further expansion was performed because experimental data have revealed a dramatic decline in the homing capacity of *in vitro* amplified hematopoietic stem or progenitor cells.³⁰

The second question was how to deliver the cells most efficiently. When given intravenously, only a very small fraction of infused cells can reach the infarct region after the following injection: assuming a normal coronary blood flow of 80 mL/min per 100 g of LV weight, a quantity of 160 mL per left ventricle (assuming a regular LV mass of ~200 g) will flow per minute.^{31,32} This corresponds to only about 3% of cardiac output (assuming a cardiac output of 5000 mL/min).³¹ Therefore, intravenous application would require many circulation passages to enable infused cells to come into contact with the infarct-related artery. Throughout this long circulation and recirculation time, homing of cells to other organs could considerably reduce the numbers of cells dedicated to cell repair in the infarcted zone. Thus, supplying the entire complement of cells by intracoronary administration obviously seems to be advantageous for the tissue repair of infarcted heart muscle and may also be superior to intraventricular injection,³³ because all cells are able to flow through the infarcted and peri-infarcted tissue during the immediate first passage. Accordingly, by this intracoronary procedure the infarct tissue and the peri-infarct zone can be enriched with the maximum available amount of cells at all times.

As stem cells differentiate into more mature types of progenitor cells, it is thought that a special microenvironment in so-called niches regulates cell activity by providing specific combinations of cytokines and by establishing direct cellular contact. For successful long-term engraftment, at least some stem cells have to reach their niches, a process referred to as homing. Mouse experiments have shown that significant numbers of BMCs appear in liver, spleen, and bone marrow after intravenous injection.³⁴ To offer the BMCs the best chance of finding their niche within the myocardium, a selective intracoronary delivery route was chosen. Presumably, therefore, fewer cells were lost by extraction toward organs of secondary interest by this first pass-like effect. To facilitate transendothelial passage and migration into the infarcted zone, cells were infused by high-pressure injection directly into the necrotic area, and the balloon was kept inflated for 2 to 3 minutes; the cells were not washed away immediately under these conditions.

The time point for delivery was chosen as 7 to 8 days after infarction onset for the following reasons:

- (1) In dogs, infarcted territory becomes rich in capillaries and contains enlarged, pericyte-poor "mother vessels" and endothelial bridges 7 days after myocardial ischemia and reperfusion. Twenty-eight days later, a significant muscular vessel wall has already formed.³⁵ Thus, with such timing, cells may be able to reach the worst

damaged parts and at the same time salvage tissue. Transendothelial cell migration may also be enhanced because an adequate muscular coat is not yet formed.

- (2) Until now, only one animal study has attempted to determine the optimum time for cardiomyocyte transplantation to maximize myocardial function after LV injury. Adult rat hearts were cryoinjured and fetal rat cardiomyocytes were transplanted immediately, 2 weeks later, and 4 weeks later. The authors discussed the inflammatory process, which is strongest in the first days after infarction, as being responsible for the negative results after immediate cell transplantation, and they assumed that the best results seen after 2 weeks may have been due to transplantation before scar expansion.³⁶ Until now, however, no systematic experiments have been performed with BMCs to correlate the results of transplantation with the length of such a time delay.
- (3) Another important variable is the inflammatory response in MI, which seems to be a superbly orchestrated interaction of cells, cytokines, growth factors and extracellular matrix proteins mediating myocardial repair. In the first 48 hours, debridement and formation of a fibrin-based provisional matrix predominates before a healing phase ensues.³⁷⁻⁴⁰ Moreover, vascular endothelial growth factor is at its peak concentration 7 days after MI, and the decline of adhesion molecules (intercellular adhesion molecules, vascular cell adhesion molecules) does not take place before days 3 to 4 after MI. We assumed that transplantation of mononuclear BMCs within the "hot" phase of post-MI inflammation might lead them to take part in the inflammation cascade rather than the formation of functional myocardium and vessels.

Taking all of this into account, we can conclude that cell transplantation within the first 5 days after acute infarction is not possible for logistical reasons and is not advisable because of the inflammatory process. On the other hand, transplantation 2 weeks after infarction scar formation seems to reduce the benefit of cell transplantation. Although the ideal time point for transplantation remains to be defined, it is most likely between days 7 and 14 after the onset of MI, as in the present study.

This trial was designed as a phase I safety and feasibility trial, meaning that no control group is necessarily required. However, to validate the results, we correlated them with those obtained from 10 patients who refused to get additional cell therapy and thus received standard therapy alone. We are aware of the fact that such a comparison does not reach the power of a randomly allocated, blinded control group. However, the significant improvement with regard to infarct region, hemodynamics (stroke volume index), cardiac geometry (LV end-systolic volume), and contractility (P_{max}/ESV and infarction wall movement velocity) did confirm a positive effect of the additional cell therapy because the changes observed in the standard therapy group failed to reach significance.

Another important factor for interpreting the results is time interval between onset of symptoms and revascularization of the infarct-related artery by angioplasty; this represents a crucial determinant of LV recovery. For patients with acute MI, it has

been shown that if the time interval is >4 hours, no significant changes in ejection fraction, regional wall motion, or ESV are observed after 6-month follow-up by echocardiography and angiography.⁴¹ None of our 20 patients was treated by angioplasty within 4 hours after onset of symptoms. Our average time interval was 12 ± 10 hours. Thus, PTCA-induced improvement of LV function can be nearly excluded; indeed, the only mild and nonsignificant changes within the standard therapy group are consistent with the above-mentioned data.⁴¹ In contrast, the cell therapy group showed considerable and significant improvement in the same parameters, which may be attributed to BMC-mediated coronary angiogenesis and cardiomyogenesis.

These results show that transplantation of autologous BMCs, as well as the intracoronary approach, represent a novel and effective therapeutic procedure for the repair of infarcted myocardium. For this method of therapy, no ethical problems exist, and no side effects were observed at any point of time. The therapeutic benefit for the patient's heart seems to prevail. However, further experimental studies, controlled prospective clinical trials, and variations of cell preparations are required to define the role of this new approach for the therapy of acute MI in humans.

References

- Pfeffer MA, Braunwald E. Ventricular remodeling after myocardial infarction: experimental observations and clinical implications. *Circulation*. 1990;81:1161-1172.
- Ertl G, Gaudron P, Hu K. Ventricular remodeling after myocardial infarction: experimental and clinical studies. *Basic Res Cardiol*. 1993;88:125-137.
- Quaini F, Urbanek K, Beltrami AP, et al. Chimerism of the transplanted heart. *N Engl J Med*. 2002;346:5-15.
- Leor J, Patterson M, Quinones MJ, et al. Transplantation of fetal myocardial tissue into infarcted myocardium of rat: a potential method for repair of infarcted myocardium? *Circulation*. 1996;94(suppl II):332-336.
- Murry CE, Wiseman RW, Schwartz SM, et al. Skeletal myoblast transplantation for repair of myocardial necrosis. *J Clin Invest*. 1996;98:2512-2523.
- Taylor DA, Atkins BZ, Hungspreugs P, et al. Regenerating functional myocardium: improved performance after skeletal myoblast transplantation. *Nat Med*. 1998;4:929-933.
- Tomita S, Li RK, Weisel RD, et al. Autologous transplantation of bone marrow cells improves damaged heart function. *Circulation*. 1999;100(suppl II):247-256.
- Blau HM, Brazelton TR, Weimann JM. The evolving concept of a stem cell: entity or function? *Cell*. 2001;105:829-841.
- Krause DS, Theise ND, Collector MI, et al. Multi-organ, multi-lineage engraftment by a single bone marrow-derived stem cell. *Cell*. 2001;105:369-377.
- Goodell MA, Jackson KA, Majka SM, et al. Stem cell plasticity in muscle and bone marrow. *Ann NY Acad Sci*. 2001;938:208-218.
- Orlic D, Kajstura J, Chimenti S, et al. Bone marrow cells regenerate infarcted myocardium. *Nature*. 2001;410:701-705.
- Orlic D, Kajstura J, Chimenti S, et al. Mobilized bone marrow cells repair the infarcted heart, improving function and survival. *Proc Natl Acad Sci U S A*. 2001;98:10344-10349.
- Kocher AA, Schuster MD, Szabolcs MJ, et al. Neovascularization of ischemic myocardium by human bone-marrow-derived angioblasts prevents cardiomyocyte apoptosis, reduces remodeling and improves cardiac function. *Nat Med*. 2001;7:430-436.
- Tomita S, Mickle DA, Weisel RD, et al. Improved heart function with myogenesis and angiogenesis after autologous porcine bone marrow stromal cell transplantation. *J Thorac Cardiovasc Surg*. 2002;123:1132-1135.
- Menasche P, Hagege AA, Scorsin M, et al. Myoblast transplantation for heart failure. *Lancet*. 2001;357:279-280.
- Hamano K, Nishida M, Hirata K, et al. Local implantation of autologous bone marrow cells for therapeutic angiogenesis in patients with ischemic heart disease: clinical trial and preliminary results. *Jpn Circ J*. 2001;65:845-847.
- Strauer BE, Brehm M, Zeus T, et al. Myocardial regeneration after intracoronary transplantation of human autologous stem cells following acute myocardial infarction. *Disch med Wschr*. 2001;126:932-938.
- Wang JS, Shum-Tim D, Chedrawy E, et al. The coronary delivery of marrow stromal cells for myocardial regeneration: pathophysiologic and therapeutic implications. *J Thorac Cardiovasc Surg*. 2001;122:699-705.
- Sheehan FH, Bolson EL, Dodge HT, et al. Advantages and applications of the Centerline method for characterizing regional ventricular function. *Circulation*. 1986;74:293-305.
- Sussman M. Cardiovascular biology: hearts and bones. *Nature*. 2001;410:640-641.
- Toma C, Pittenger MF, Cahill KS, et al. Human mesenchymal stem cells differentiate to a cardiomyocyte phenotype in the adult murine heart. *Circulation*. 2002;105:93-98.
- Kamihata H, Matsubara H, Nishiue T, et al. Implantation of bone marrow mononuclear cells into ischemic myocardium enhances collateral perfusion and regional function via side supply of angioblasts, angiogenic ligands, and cytokines. *Circulation*. 2001;104:1046-1052.
- Ferrari G, Cusella-De Angelis G, Coletta M, et al. Muscle regeneration by bone marrow-derived myogenic progenitors. *Science*. 1998;279:1528-1530.
- Kawamoto A, Gwon HC, Iwaguro H, et al. Therapeutic potential of ex vivo expanded endothelial progenitor cells for myocardial ischemia. *Circulation*. 2001;103:634-637.
- Robinson SW, Cho PW, Levitsky HI, et al. Arterial delivery of genetically labelled skeletal myoblasts to the murine heart: long-term survival and phenotypic modification of implanted myoblasts. *Cell Transplant*. 1996;5:77-91.
- Bitner RE, Schofer C, Weipolshammer K, et al. Recruitment of bone-marrow-derived cells by skeletal and cardiac muscle in adult dystrophic mdx mice. *Anat Embryol (Berl)*. 1999;199:391-396.
- Jiang Y, Jahagirdar B, Reinhardt RL, et al. Pluripotency of mesenchymal stem cells derived from adult marrow. *Nature*. 2002;20:1-12.
- Reyes M, Lund T, Lenvik T, et al. Purification and ex vivo expansion of postnatal human marrow mesodermal progenitor cells. *Blood*. 2001;98:2615-2625.
- Condorelli G, Borello U, De Angelis L, et al. Cardiomyocytes induce endothelial cells to trans-differentiate into cardiac muscle: implications for myocardium regeneration. *Proc Natl Acad Sci U S A*. 2001;98:10733-10738.
- Szilvassy SJ, Bass MJ, Van Zant G, et al. Organ-selective homing defines engraftment kinetics of murine hematopoietic stem cells and is compromised by ex vivo expansion. *Blood*. 1999;93:1557-1566.
- Gregg DE, Fisher LC. Blood supply to the heart. In: *Handbook of Physiology*. Vol 2. Washington, DC: American Physiological Society; 1963:chap 44, 1517-1584.
- Strauer BE. Myocardial oxygen consumption in chronic heart disease: role of wall stress, hypertrophy and coronary reserve. *Am J Cardiol*. 1979;4:730-740.
- Toma C, Pittenger MF, Byrne BJ, et al. Adult human mesenchymal stem cells differentiate to a striated muscle phenotype following arterial delivery to the murine heart. *Circulation*. 2000;102(suppl II):II-683. Abstract.
- Hendriks PJ, Martens CM, Hagenbeek A, et al. Homing of fluorescently labeled murine hematopoietic stem cells. *Exp Hematol*. 1996;24:129-140.
- Ren G, Michael LH, Entman ML, et al. Morphological characteristics of the microvasculature in healing myocardial infarcts. *J Histochem Cytochem*. 2002;50:71-79.
- Li RK, Mickle DA, Weisel RD, et al. Optimal time for cardiomyocyte transplantation to maximize myocardial function after left ventricular injury. *Ann Thorac Surg*. 2001;72:1957-1963.
- Frangogiannis NG, Smith CW, Entman ML. The inflammatory response in myocardial infarction. *Cardiovasc Res*. 2002;53:31-47.
- Allgöwer M. *The Cellular Basis of Wound Repair*. Springfield, Ill: Charles C. Thomas; 1956.
- Xie Y, Zhou T, Shen W, et al. Soluble cell adhesion molecules in patients with acute coronary syndrome. *Chin Med J*. 2000;113:286-288.
- Soeki T, Tamura Y, Shinohara H, et al. Serial changes in serum VEGF and HGF in patients with acute myocardial infarction. *Cardiology*. 2000;93:168-174.
- Sheiban I, Frangasso G, Rosano GMC, et al. Time course and determinants of left ventricular function recovery after primary angioplasty in patients with acute myocardial infarction. *J Am Coll Cardiol*. 2001;38:464-471.



PubMed

Nucleotide

Protein

Genome

Structure

PMC

Taxonomy

OMIM

Books

Search for

☒ Limits

☐ Preview/Index

☐ History

☐ Clipboard

☐ Details

About Entrez

Text Version

Entrez PubMed

Overview

Help | FAQ

Tutorial

New/Noteworthy

E-Utilities

PubMed Services

Journals Database

MeSH Database

Single Citation Matcher

Batch Citation Matcher

Clinical Queries

LinkOut

Cubby

Related Resources

Order Documents

NLM Gateway

TOXNET

Consumer Health

Clinical Alerts

ClinicalTrials.gov

PubMed Central

Privacy Policy

1: Lancet 2003 Feb 8;361(9356):491-2

[ELSEVIER SCIENCE](#)
[FULL-TEXT/ARTICLE](#)

Viability and differentiation of autologous skeletal myoblast grafts in ischaemic cardiomyopathy.

Hagege AA, Carrion C, Menasche P, Vilquin JT, Duboc D, Marolleau JP, Desnos M, Bruneval P.

Assistance Publique-Hopitaux de Paris, Department of Cardiology, Hopital Europeen Georges Pompidou and INSERM EMI-16, Necker-Paris V University, Paris, France. hagege@club-internet.fr

Autologous skeletal myoblast transplantation might improve postinfarction ventricular function, but graft viability and differentiation (ie, proof of concept) has not been shown. A 72-year-old man had autologous cultured myoblasts from his vastus lateralis injected to an area of transmural inferior myocardial infarction in non-reperused scar tissue. He showed improvement in symptoms and left-ventricular ejection fraction. When he died 17.5 months after the procedure, the grafted post-infarction scar showed well developed skeletal myotubes with a preserved contractile apparatus. 65% of myotubes expressed the slow myosin isoform and 33% coexpressed the slow and fast isoforms (vs 44% and 0.6%, respectively, in skeletal muscle). Myoblast grafts can survive and show a switch to slow-twitch fibres, which might allow sustained improvement in cardiac function.

PMID: 12583951 [PubMed - indexed for MEDLINE]

Related Articles, Links

myocardial tissue. These results establish the feasibility of myoblast transplants for myocardial repair in humans.

Publication Types:

- Clinical Trial
- Clinical Trial, Phase I

PMID: 12628737 [PubMed - indexed for MEDLINE]

Display	Abstract	Show:	20	Sort	Send to	Text
---------	----------	-------	----	------	---------	------

Write to the Help Desk
[NCBI](#) | [NLM](#) | [NIH](#)
 Department of Health & Human Services
[Freedom of Information Act](#) | [Disclaimer](#)

May 2 2003 16:34:23

This is Google's cache of http://www.medscape.com/viewarticle/444727_print.
 Google's cache is the snapshot that we took of the page as we crawled the web.
 The page may have changed since that time. Click here for the current page without highlighting.
 To link to or bookmark this page, use the following url: http://www.google.com/search?q=cache:cxRoFu2EUxYC:www.medscape.com/viewarticle/444727_print+regenerate+dead+heart+tissue+after+myocardial+infarction&hl=en&ie=UTF-8

Google is not affiliated with the authors of this page nor responsible for its content.

These search terms have been

regenerate dead heart tissue after myocardial infarction



www.medscape.com

To Print: Click your browser's PRINT button.

NOTE: To view the article with Web enhancements, go to:
<http://www.medscape.com/viewarticle/444727>



Autologous Cell Transplant Helpful in Ischemic Heart or Legs

Laurie Barclay, MD

Medscape Medical News 2002. © 2002 Medscape

Nov. 18, 2002 — Autologous cell transplantation may benefit ischemic hearts and legs, according to three presentations on Nov. 18 at the American Heart Association's 75th Scientific Sessions held in Chicago, Illinois. Two studies focused on injecting autologous bone marrow cells or autologous skeletal myoblasts into the scarred area of an infarcted heart. In another study, injecting autologous bone marrow into ischemic limbs led to new vessel growth, reducing the need for amputation.

"Bone marrow not only can differentiate into heart cells, but also smooth muscle cells, connective tissue cells and other types of cells to reconstitute the entire structure of a tissue," presenter Manuel Galinanes, MD, from the University of Leicester in the U.K., says in a news release. "The benefit [of transplanting bone marrow into scar tissue of the heart] could be seen only six weeks after injection."

In 14 patients with low ejection fraction post-myocardial infarction (MI), autologous bone marrow from the sternum was injected into scarred myocardium during nonemergency coronary artery bypass surgery. Heart wall motion measured with echocardiography improved within weeks of treatment, and improvements persisted for at least 10 months after treatment.

The regional wall motion score decreased significantly, reflecting less movement abnormality, from a mean score of 2.41 at baseline to 2.16 six weeks after treatment and 2.09 ten months after treatment. The global wall motion score also decreased significantly from 1.96 before surgery to 1.64 at six weeks, and stabilized at 1.65 after 10 months.

Although it is still unproven that bone marrow creates a new cellular infrastructure in heart scar tissue, "that is the only possible explanation," Galinanes says. "The ability to confirm the presence of scar tissue with dobutamine stress echo before surgery, and then confirm it again during surgery, told us that the affected area was dysfunctional and the abnormality was irreversible. We wanted to make sure that we were injecting the marrow into dead tissue to help ensure that the injection would not pose any serious risk to the patient."

If additional studies confirm safety and efficacy, Galinanes says that this treatment would be a welcome addition to the post-MI arsenal, which also includes gene therapy, growth factor therapy, and laser treatments.

In a multicenter trial supervised by the U.S. Food and Drug Administration, investigators safely transplanted 16 patients with autologous skeletal myoblasts injected into hearts severely damaged by MI or heart failure. Baseline left-ventricular ejection fraction was less than 30%. Eleven patients were undergoing coronary artery bypass surgery and five were having implantation of a left ventricular assist device. Myoblasts extracted from thigh muscle were grown in large quantities in vitro using a controlled cell expansion manufacturing process, and were injected in doses ranging from 10 million to 300 million cells.

"We have been able to regenerate dead heart muscle, or scar tissue, in the area of heart attack without increasing risk of death. Our findings will allow us to move forward with testing if the procedure can improve the contractility of the heart," says lead author Nabil Dib, MD, from the Arizona Heart Institute in Phoenix. "We found that the transplanted myoblasts survived and thrived in patients. Areas damaged by heart attack and cardiovascular disease showed evidence of repair and viability."

Twelve weeks after transplant, mean ejection fraction rates improved from 22.7% to 35.8%, or a 58% increase. Echocardiogram, magnetic resonance imaging, and positron emission tomography showed evidence of regeneration in the area of the graft. There were no significant adverse events related to the cell transplant procedure at nine-month follow-up.

The third study showed that bone marrow cells implanted into ischemic legs in patients with peripheral arterial disease (PAD) formed new blood vessels, increased blood flow, and prevented amputation.

"This is the first multicenter and double-blind clinical study to prove the clinical efficacy of growing new blood vessels (angiogenesis) using bone marrow cell transplantation," says lead author Hiroya Masaki, MD, PhD, from Kansai Medical University in Osaka, Japan.

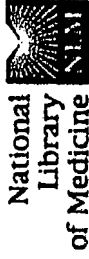
In this randomized trial, 45 patients with PAD received injections of autologous bone marrow mononuclear cells into the calf muscles. Compared with controls who received saline injections, patients who received bone marrow mononuclear cell transplants had a "striking" increase in new capillary formation and in newly visible collateral vessels.

Of 45 treated patients, 31 had an increase in ankle-brachial pressure index in the treated limbs, and 39 had decreased rest pain with improved treadmill endurance. Ischemic ulcers or gangrene healed in 21 of 28 treated limbs.

CD34-cells, which can develop into endothelial progenitor cells, expressed angiogenic growth factors including basic fibroblast growth factor, vascular endothelial growth factor, and angiopoietin-1. Although more research is needed to determine long-term efficacy and safety, "this new angiogenesis therapy using bone marrow cell transplantation may help many patients suffering with ischemic limbs," Masaki says.

AHA 75th Scientific Sessions: Abstracts 111623, 101758, 109801. Presented Nov. 18, 2002.

Reviewed by Gary D. Vagin, MD



PubMed

Nucleotide

Protein

Genome

Structure

PMC

Taxonomy

Books

Search PubMed for

☒ Limits

Preview/Index

History

Clipboard

Details

About Entrez

Text Version

Entrez PubMed

Overview

Help | FAQ

Tutorial

New/Noteworthy

E-Utilities

PubMed Services

Journals Database

MeSH Database

Single Citation Matcher

Batch Citation Matcher

Clinical Queries

LinkOut

Cubby

Related Resources

Order Documents

NLM Gateway

TOXNET

Consumer Health

Clinical Alerts

ClinicalTrials.gov

PubMed Central

Privacy Policy

☐ 1: J Am Coll Cardiol 2003 Mar 5;41(5):879-88

[PUBMED SCIENCE
FULL-TEXT ARTICLE](#)

Autologous skeletal myoblasts transplanted to ischemia-damaged myocardium in humans. Histological analysis of cell survival and differentiation.

Pagani FD, DerSimonian H, Zawadzka A, Wetzel K, Edge AS, Jacoby DB, Dinsmore JH, Wright S, Aretz TH, Eisen HJ, Aaronson KD.

Section of Cardiac Surgery, University of Michigan, Ann Arbor, MI 48109, USA. fpagani@umich.edu

Related Articles, Links

OBJECTIVES: We report histological analysis of hearts from patients with end-stage heart disease who were transplanted with autologous skeletal myoblasts concurrent with left ventricular assist device (LVAD) implantation. **BACKGROUND:** Autologous skeletal myoblast transplantation is under investigation as a means to repair infarcted myocardium. To date, there is only indirect evidence to suggest survival of skeletal muscle in humans. **METHODS:** Five patients (all male; median age 60 years) with ischemic cardiomyopathy, refractory heart failure, and listed for heart transplantation underwent muscle biopsy from the quadriceps muscle. The muscle specimen was shipped to a cell isolation facility where myoblasts were isolated and grown. Patients received a transplant of 300 million cells concomitant with LVAD implantation. Four patients underwent LVAD explant after 68, 91, 141, and 191 days of LVAD support (three transplant, one LVAD death), respectively. One patient remains alive on LVAD support awaiting heart transplantation. **RESULTS:** Skeletal muscle cell survival and differentiation into mature myofibers were directly demonstrated in scarred myocardium from three of the four explanted hearts using an antibody against skeletal muscle-specific myosin heavy chain. An increase in small vessel formation was observed in one of three patients at the site of surviving myotubes, but not in adjacent tissue devoid of engrafted cells. **CONCLUSIONS:** These findings represent demonstration of autologous myoblast cell survival in human heart. The implanted skeletal myoblasts formed viable grafts in heavily scarred human

EVIDENCE APPENDIX

Item 20

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

APPLICANT: James P. Elia)	
)	
SERIAL NO.: 09/836,750)	EXAMINER: E.C. Kemmerer, Ph.D.
)	
FILED: April 17, 2001)	
)	GROUP ART UNIT: 1646
FOR: METHOD FOR GROWING)	
MUSCLE IN A HUMAN HEART)	

SUPPLEMENTAL DECLARATION OF ANDREW E. LORINCZ, M.D.

I, Andrew E. Lorincz, declare as follows:

1. I reside at 13820 NW County Rd 235, Apt 8, Alachua, FL 32616-2098.
2. This Supplemental Declaration is submitted in addition to my previously submitted Declaration in this application, dated June 9, 2003, and makes no changes to such previous Declaration.
3. My Curriculum Vitae is attached as Exhibit A to my previous Declaration.
4. I have read and understood the disclosures of the above-referenced patent application at page 20, line 10 through page 21, line 15; and page 44, line 19 through page 46, line 16. Such disclosures are the same as read and understood by me in my previous Declaration. A copy of such disclosures is attached hereto as Supplemental Exhibit A.

5. I note that the disclosures referenced in above Paragraph 4 relate to using a growth factor for promoting the growth of soft tissue and, more specifically, to a method which may use such growth factors for growing a new portion of a human heart by growing new muscle in the heart.
6. I have read and understood the claims set forth in Supplemental Exhibit B and have been informed that such claims will be presented to the Patent and Trademark Office in the near future.
7. Based upon above Paragraphs 4-6 and Paragraph 7 of my previous Declaration, it is my opinion that introducing a growth factor into a human patient will predictably cause new muscle growth in the heart of the patient.
8. Based upon above Paragraphs 4-6, it is my opinion that one skilled in the medical arts, armed with the knowledge in such paragraphs, would be able to practice the method set forth in Supplemental Exhibit B without need for resorting to undue experimentation. I have been informed that the Examiner has questioned the fact that dosages are not recited in the specification of the above-identified application in connection with the administration of cell growth factors to a human patient with use of intravenous or intraluminal techniques. Such techniques are the subject of claims 248-249 in above-mentioned Supplemental Exhibit B. In my opinion, dosages of cellular growth factors to achieve the above-mentioned heart muscle growth are a matter of routine medical practice, requiring only a reasonable degree of experimentation, depending upon such factors as extent of prior heart condition, size of patient, age of patient, health of patient, etc. Consequently, it is my opinion that the disclosure mentioned in Supplemental Exhibit A would enable a person skilled in the medical arts to practice the invention of claims 248-249 and predictably anticipate the results defined therein without need for resorting to undue experimentation.

9. Declarant states that the above opinion was reached independently.

Declarant understands that (1) any willful false statements and the like made herein are punishable by fine or imprisonment, or both (18 U.S.C. 1001) and may jeopardize the validity of the application or any patent issuing thereon, and (2) that all statements made of Declarant's own knowledge are true and that all statements made on information and belief are believed to be true.

Further Declarant sayeth not.

Date: 2-3-04

Andrew E. Lorincz
Andrew E. Lorincz

SUPPLEMENTAL EXHIBIT A
DISCLOSURES
APPLICATION SERIAL NO. 09/836,750

PAGE 20, LINE 10 – PAGE 21, LINE 15

Growth factors can be utilized to induce the growth of “hard tissue” or bone and “soft tissues” like ectodermal and mesodermal tissues. As used herein, the term growth factor encompasses compositions and living organisms which promote the growth of hard tissue, such as bone, or soft tissue, in the body of a patient. The compositions include organic and inorganic matter. The compositions can be genetically produced or manipulated. The living organisms can be bacteria, viruses, or any other living organism which promote tissue growth. By way of example and not limitation, growth factors can include platelet-derived growth factor (PDGF), epidermal growth factor (EGF), fibroblast growth factor (acidic/basic (FGF a,b), interleukins (IL's), tumor necrosis factor (TNF), transforming growth factor (TGF-B), colony-stimulating factor (CSF), osteopontin (Eta-1 OPN), platelet-derived growth factor (PDGF), interferon (INF), bone morphogenic protein 1 (BMP-1), and insulin growth factor (IGF). Recombinant and non-recombinant growth factors can be utilized as desired. Bacteria or viruses can, when appropriate, be utilized as growth factors. For example, there is a bacterial hydrophilic polypeptide that self-assembles into a nanometer internal diameter pore to build a selective lipid body. Various enzymes can be utilized for the synthesis of peptides which contain amino acids that control three-dimensional protein structure and growth. Growth factors can be applied in gels or other carriers which regulate the rate of release of the growth factors and help maintain the growth factors and the carrier, at a desired location in the body. Time release capsules, granules, or other carriers containing growth factor can be activated by tissue pH, by enzymes, by ultrasound,

by electricity, by heat, by selected *in vivo* chemicals or by any other selected means to release the growth factor. The carrier can be resorbable or non-resorbable. Or, the growth factor itself can be activated by similar means. Either the carrier or the growth factor can mimic extracellular fluid to control cell growth, migration, and function. The growth factor can be administered orally, systemically, in a carrier, by hypodermic needle, through the respiratory tract, or by any other desired method. The growth factor can also be administered into a capsule or other man-made composition or structure placed in the body. While administration of the growth factor is presently usually localized in the patient's body, circumstances may arise where it is advantageous to distribute a growth factor throughout the patient's body in uniform or non-uniform concentrations. An advantage to growth factors is that they can often, especially when in capsule form or in some other containment system, be inserted to a desired site in the body by simply making a small incision and inserting the growth factor. The making of such small incision comprises minor surgery which can often be accomplished on an out-patient basis. The growth factors can be multifactorial and nonspecific.

PAGE 44, LINE 19 – PAGE 46, LINE 16

Genetic material comprising a portion of a gene, a gene, genes, a gene product (i.e., a composition a gene causes to be produced like, for example, an organ-producing growth factor), growth factor, or an ECM (extracellular matrix) can be used in or on the body to grow an organ to tissue. For example, the vascular epithelial growth factor gene (VEGF) or its growth factor equivalent can be inserted into the body to cause an artery to grow. When insertion of a gene, portion of a gene, gene product, growth factor, or ECM *in vivo* or *ex vivo* is referred to herein in connection with any of the implant techniques of the invention, it is understood that a cell

nutrient culture(s), physiological nutrient culture(s), carrier (s), enhancer(s), promoter(s), or any other desired auxiliary component(s) can be inserted with the gene or at the same location as the gene, growth factor, ECM, etc.

An artery is an organ from the circulatory system. An artery can be grown in the heart, legs, or other areas by injecting a gene or other genetic material into muscle at a desired site. Size, vascularity, simplicity of access, ease of exploitation, and any other desired factors can be utilized in selecting a desired site. The gene is one of several known VEGF genes which cause the production of vascular endothelial growth factors. Several VEGF genes which produce vascular endothelial growth factors are believed to exist because nature intends for there to be several pathways (i.e., genes) which enable the production of necessary growth factors. The existence of several pathways is believed important because if one of the genes is damaged or inoperative, other similar genes can still orchestrate the production of necessary growth factors. VEGF genes are used by the body to promote blood vessel growth. VEGF genes are assimilated (taken in) by muscle cells. The genes cause the muscle cells to make a VEGF protein which promotes the growth of new arteries. VEGF proteins can be made in a lab and injected into a patient intravenously, intraluminally, or intramuscularly to promote the growth of an artery. Or, the genes (or other genetic material) can be applied with an angioplasty balloon, with the assistance of a vector, or by any other method.

It is not always desirable to grow a completely new organ. Sometimes growing a portion of an organ is desirable. For example, in some heart attacks or strokes, a portion of the heart or brain remains viable and a portion dies. An injection of a gene to form cardiac muscle and/or an injection of a gene to form an artery can be utilized to revive or replace the dead portion of the heart. The dead portion of the heart may (or may not) be used as a matrix while the new muscles

and vessels grow. Thus, in this example, a partial new organ is grown in a pre-existing organ. A pacemaker may (or may not) be necessary. A second injection of a gene may (or may not) be necessary to stop cardiac muscle growth once it is completed. Portions of organs throughout the body can similarly be repaired or replaced. It may be necessary to provide gene(s) or growth factor(s) sequentially. For instance, one or more blood vessels are grown by inserting an appropriate gene or other genetic material into a selected area. Second, an appropriate gene or other genetic material is inserted in the selected area to grow a bone or other organ.

The size and shape limitation of the desired structure can come from a containment and boundary contact inhibition phenomenon or by a chemical inhibition.

A variation on the theme of growing a portion of an organ is as follows: a portion of a heart dies. The pericardium is utilized as a scaffold and seeded with cells and/or genes to grow new muscle, and genes (or other genetic material) to grow new arteries. Immediately adjacent the dead cardiac muscle, onto or into the pericardium, the appropriate cells, genes, and/or growth factors (or other genetic material) are placed. Once the new muscle and blood vessels have grown, the function specific tissue can be applied to the damaged portion of the heart and paced, if necessary, to augment cardiac action. If the surgeon desires, the dead muscle can be removed and the new muscle and blood vessels can be surgically rotated into the excised region and secured. This probably can be done endoscopically. In essence, the pericardium is utilized to allow the new muscle wall to grow. The new muscle wall is then transplanted into the damaged heart wall. This procedure utilizes the body as a factor to grow an organ and/or tissue, after which the organ and/or tissue is transplanted to a desired region. On the other hand, the new muscle wall may integrate itself into the old wall and not require transplantation.

SUPPLEMENTAL EXHIBIT B

CLAIMS APPLICATION SERIAL NO. 09/836,750

- 236. A method of growing a new portion of a pre-existing heart comprising the steps of placing a growth factor in a body of a human patient and growing new muscle and growing a new artery in said heart.
- 238. The method of claim 236, further comprising repairing a dead portion of said heart.
- 239. The method of claim 236, further comprising repairing a damaged portion of said heart.
- 240. The method of claim 236, wherein said growth factor comprises genetic material selected from the group consisting of a portion of a gene, a gene, a gene product, and an extracellular matrix.
- 241. The method of claim 240, wherein said genetic material comprises a gene.
- 242. The method of claim 241, wherein said gene comprises VEGF.
- 243. The method of claim 236, wherein said growth factor comprises a member selected from the group consisting of cells, cellular products, and derivatives of cellular products.
- 244. The method of claim 243, wherein said growth factor comprises a cell
- 245. The method of claim 244, wherein said cell is multifactorial and non-specific.
- 246. The method of claim 245, wherein said cell comprises a stem cell.

- 247. The method of claim 236, wherein said growth factor is placed in said patient by injection.
- 248. The method of claim 247, wherein said injection is intravenous.
- 249. The method of claim 247, wherein said injection is intraluminal.
- 250. The method of claim 247, wherein said injection is intramuscular.
- 251. The method of claim 236, wherein said growth factor is placed in said patient by a carrier.
- 252. The method of claim 251, wherein said carrier comprises an angioplasty balloon.
- 253. The method of claim 236, wherein said growth factor comprises a gene and a cell.

EVIDENCE APPENDIX

Item 21

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

APPLICANT: James P. Elia)	
)	
SERIAL NO.: 09/836,750)	EXAMINER: E.C. Kemmerer, Ph.D.
)	
FILED: April 17, 2001)	
)	GROUP ART UNIT: 1646
FOR: METHOD FOR GROWING)	
MUSCLE IN A HUMAN HEART)	

**SECOND SUPPLEMENTAL DECLARATION
OF ANDREW E. LORINCZ, M.D.**

I, Andrew E. Lorincz, declare as follows:

1. I reside at 13820 NW County Rd 235, Apt 8, Alachua, FL 32616-2098.
2. This Second Supplemental Declaration is submitted in addition to my previous Declaration dated June 5, 2003 and my Supplemental Declaration dated February 3, 2004. No changes are made to either of such previous Declarations.
3. My Curriculum Vitae (hereinafter "CV") is attached as Exhibit A to my previous Declaration.
4. It is my understanding that the Examiner in charge of the above-identified patent application, in an Office Action dated June 1, 2004 for related patent application Serial No. 09/794,456, questioned my qualification, for the first time, to render my previous opinions mentioned in above Paragraph 2. It is my further understanding that the basis for such questioning was that the Examiner noted that I did not report experience with cellular therapy. I desire to provide the

information contained in following paragraph 5 so that the Examiner can consider such information in this application, as well.

5. In addition to the qualifications set forth in my CV, I am familiar with stem cell technology, including bone marrow preparation.
6. I have read and understood the disclosures of the above-referenced patent application at page 20, line 10 through page 21, line 15; and page 44, line 19 through page 46, line 16. Such disclosures are the same as I read and understood in my previous Declaration and Supplemental Declaration. A copy of such disclosures is attached hereto as Second Supplement Declaration Exhibit A.
7. I note that the disclosures referenced in above Paragraph 6 relate to using a growth factor for promoting the growth of soft tissue and, more specifically, to a method which may use such growth factors for growing a new portion of a human heart by growing new cardiac muscle in the heart.
8. I have read and understood the claims set forth in Second Supplemental Declaration Exhibit B and have been informed that such claims are currently presented in this application.
9. Based upon above Paragraphs 6-8, it is my opinion that one skilled in the medical arts, armed with the knowledge in such paragraphs, would be enabled to practice the method set forth in Second Supplemental Declaration Exhibit B and to predictably anticipate the results defined therein without need for resorting to undue experimentation.
10. I believe that one skilled in the medical arts, upon reading the disclosures in above Paragraph 6, would understand that cellular growth factors, such as multifactorial and non-specific cells, are included in such disclosures. Moreover,

such skilled person would understand the disclosure on page 45 to be authored as an illustration of various modes of delivery of growth factors, whether they are genes or other genetic material; and that such skilled person would further understand that the disclosures on pages 45 and 46 describe genetic material to include appropriate cells and genes.

11. Declarant states that the above opinion was reached independently.

Declarant understands that (1) any willful false statements and the like made herein are punishable by fine or imprisonment, or both (18 U.S.C. 1001) and may jeopardize the validity of the application or any patent issuing thereon, and (2) that all statements made of Declarant's own knowledge are true and that all statements made on information and belief are believed to be true.

Further Declarant sayeth not.

Date: 7-19-04

Andrew E. Lorincz, M.D.
Andrew E. Lorincz, M.D.

**SECOND
SUPPLEMENTAL
DECLARATION**

EXHIBIT A

DISCLOSURES

**SECOND SUPPLEMENTAL DECLARATION
EXHIBIT A**

**DISCLOSURES
APPLICATION SERIAL NO. 09/836,750**

PAGE 20, LINE 10 – PAGE 21, LINE 15

Growth factors can be utilized to induce the growth of “hard tissue” or bone and “soft tissues” like ectodermal and mesodermal tissues. As used herein, the term growth factor encompasses compositions and living organisms which promote the growth of hard tissue, such as bone, or soft tissue, in the body of a patient. The compositions include organic and inorganic matter. The compositions can be genetically produced or manipulated. The living organisms can be bacteria, viruses, or any other living organism which promote tissue growth. By way of example and not limitation, growth factors can include platelet-derived growth factor (PDGF), epidermal growth factor (EGF), fibroblast growth factor (acidic/basic (FGF a,b), interleukins (IL’s), tumor necrosis factor (TNF), transforming growth factor (TGF-B), colony-stimulating factor (CSF), osteopontin (Eta-1 OPN), platelet-derived growth factor (PDGF), interferon (INF), bone morphogenic protein 1 (BMP-1), and insulin growth factor (IGF). Recombinant and non-recombinant growth factors can be utilized as desired. Bacteria or viruses can, when appropriate, be utilized as growth factors. For example, there is a bacterial hydrophilic polypeptide that self-assembles into a nanometer internal diameter pore to build a selective lipid body. Various enzymes can be utilized for the synthesis of peptides which contain amino acids that control three-dimensional protein structure and growth. Growth factors can be applied in gels or other carriers which regulate the rate of release of the growth factors and help maintain the growth factors and the carrier, at a desired location in the body. Time release capsules, granules, or

other carriers containing growth factor can be activated by tissue pH, by enzymes, by ultrasound, by electricity, by heat, by selected *in vivo* chemicals or by any other selected means to release the growth factor. The carrier can be resorbable or non-resorbable. Or, the growth factor itself can be activated by similar means. Either the carrier or the growth factor can mimic extracellular fluid to control cell growth, migration, and function. The growth factor can be administered orally, systemically, in a carrier, by hypodermic needle, through the respiratory tract, or by any other desired method. The growth factor can also be administered into a capsule or other man-made composition or structure placed in the body. While administration of the growth factor is presently usually localized in the patient's body, circumstances may arise where it is advantageous to distribute a growth factor throughout the patient's body in uniform or non-uniform concentrations. An advantage to growth factors is that they can often, especially when in capsule form or in some other containment system, be inserted to a desired site in the body by simply making a small incision and inserting the growth factor. The making of such small incision comprises minor surgery which can often be accomplished on an out-patient basis. The growth factors can be multifactorial and nonspecific.

PAGE 44, LINE 19 – PAGE 46, LINE 16

Genetic material comprising a portion of a gene, a gene, genes, a gene product (i.e., a composition a gene causes to be produced like, for example, an organ-producing growth factor), growth factor, or an ECM (extracellular matrix) can be used in or on the body to grow an organ to tissue. For example, the vascular epithelial growth factor gene (VEGF) or its growth factor equivalent can be inserted into the body to cause an artery to grow. When insertion of a gene, portion of a gene, gene product, growth factor, or ECM *in vivo* or *ex vivo* is referred to herein in

connection with any of the implant techniques of the invention, it is understood that a cell nutrient culture(s), physiological nutrient culture(s), carrier (s), enhancer(s), promoter(s), or any other desired auxiliary component(s) can be inserted with the gene or at the same location as the gene, growth factor, ECM, etc.

An artery is an organ from the circulatory system. An artery can be grown in the heart, legs, or other areas by injecting a gene or other genetic material into muscle at a desired site. Size, vascularity, simplicity of access, ease of exploitation, and any other desired factors can be utilized in selecting a desired site. The gene is one of several known VEGF genes which cause the production of vascular endothelial growth factors. Several VEGF genes which produce vascular endothelial growth factors are believed to exist because nature intends for there to be several pathways (i.e., genes) which enable the production of necessary growth factors. The existence of several pathways is believed important because if one of the genes is damaged or inoperative, other similar genes can still orchestrate the production of necessary growth factors. VEGF genes are used by the body to promote blood vessel growth. VEGF genes are assimilated (taken in) by muscle cells. The genes cause the muscle cells to make a VEGF protein which promotes the growth of new arteries. VEGF proteins can be made in a lab and injected into a patient intravenously, intraluminally, or intramuscularly to promote the growth of an artery. Or, the genes (or other genetic material) can be applied with an angioplasty balloon, with the assistance of a vector, or by any other method.

It is not always desirable to grow a completely new organ. Sometimes growing a portion of an organ is desirable. For example, in some heart attacks or strokes, a portion of the heart or brain remains viable and a portion dies. An injection of a gene to form cardiac muscle and/or an injection of a gene to form an artery can be utilized to revive or replace the dead portion of the

heart. The dead portion of the heart may (or may not) be used as a matrix while the new muscles and vessels grow. Thus, in this example, a partial new organ is grown in a pre-existing organ. A pacemaker may (or may not) be necessary. A second injection of a gene may (or may not) be necessary to stop cardiac muscle growth once it is completed. Portions of organs throughout the body can similarly be repaired or replaced. It may be necessary to provide gene(s) or growth factor(s) sequentially. For instance, one or more blood vessels are grown by inserting an appropriate gene or other genetic material into a selected area. Second, an appropriate gene or other genetic material is inserted in the selected area to grow a bone or other organ.

The size and shape limitation of the desired structure can come from a containment and boundary contact inhibition phenomenon or by a chemical inhibition.

A variation on the theme of growing a portion of an organ is as follows: a portion of a heart dies. The pericardium is utilized as a scaffold and seeded with cells and/or genes to grow new muscle, and genes (or other genetic material) to grow new arteries. Immediately adjacent the dead cardiac muscle, onto or into the pericardium, the appropriate cells, genes, and/or growth factors (or other genetic material) are placed. Once the new muscle and blood vessels have grown, the function specific tissue can be applied to the damaged portion of the heart and paced, if necessary, to augment cardiac action. If the surgeon desires, the dead muscle can be removed and the new muscle and blood vessels can be surgically rotated into the excised region and secured. This probably can be done endoscopically. In essence, the pericardium is utilized to allow the new muscle wall to grow. The new muscle wall is then transplanted into the damaged heart wall. This procedure utilizes the body as a factor to grow an organ and/or tissue, after which the organ and/or tissue is transplanted to a desired region. On the other hand, the new muscle wall may integrate itself into the old wall and not require transplantation.

**SECOND SUPPLEMENTAL
DECLARATION**

EXHIBIT B

CLAIMS

**SECOND SUPPLEMENTAL DECLARATION
EXHIBIT B**

**CLAIMS
APPLICATION SERIAL NO. 09/836,750**

236. A method of growing a new portion of a pre-existing heart comprising the steps of placing a growth factor in a body of a human patient and growing new cardiac muscle and growing a new artery in said heart.
238. The method of claim 236, further comprising repairing a dead portion of said heart.
239. The method of claim 236, further comprising repairing a damaged portion of said heart.
240. The method of claim 236, wherein said growth factor comprises genetic material selected from the group consisting of a portion of a gene, a gene, a gene product, and an extracellular matrix.
241. The method of claim 240, wherein said genetic material comprises a gene.
242. The method of claim 241, wherein said gene comprises VEGF.
243. The method of claim 236, wherein said growth factor comprises a member selected from the group consisting of cells, cellular products, and derivatives of cellular products.
244. The method of claim 243, wherein said growth factor comprises a cell
245. The method of claim 244, wherein said cell is multifactorial and non-specific.
246. The method of claim 245, wherein said cell comprises a stem cell.

- 247. The method of claim 236, wherein said growth factor is placed in said patient by injection.
- 248. The method of claim 247, wherein said injection is intravenous.
- 249. The method of claim 247, wherein said injection is intraluminal.
- 250. The method of claim 247, wherein said injection is intramuscular.
- 251. The method of claim 236, wherein said growth factor is placed in said patient by a carrier.
- 252. The method of claim 251, wherein said carrier comprises an angioplasty balloon.
- 253. The method of claim 236, wherein said growth factor comprises a gene and a cell.
- 254. A method of growing a new portion of a pre-existing organ comprising placing a growth factor in a body of a patient to grow new muscle in said organ.
- 255. The method of claim 254, wherein said organ comprises a heart.
- 256. The method of claim 255, wherein said new muscle comprises cardiac muscle and said growth factor comprises a stem cell.

EVIDENCE APPENDIX

Item 22

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

APPLICANT: James P. Elia)	
)	
SERIAL NO.: 09/836,750)	EXAMINER: Elizabeth C. Kemmerer
)	
FILED: April 17, 2001)	
)	
FOR: METHOD FOR GROWING)	GROUP ART UNIT: 1646
MUSCLE IN A HUMAN HEART)	

**THIRD SUPPLEMENTAL DECLARATION
OF ANDREW E. LORINCZ, M.D.**

I, Andrew E. Lorincz, declare as follows:

1. I reside at 16135 NW 243rd Way, High Springs, Florida 32643-3813.
2. This Third Supplemental Declaration is submitted in addition to my previous Declaration, dated June 9, 2003, my Supplemental Declaration dated February 3, 2004, and my Second Supplemental Declaration dated July 19, 2004. No changes are made to any of such previous Declarations.
3. My Curriculum Vitae (hereinafter "CV") is attached as Exhibit A to my Declaration of June 9, 2003, and my background is further amplified by materials submitted in my Second Supplemental Declaration.
4. I have read and understood the disclosures of the above-referenced patent application at page 20, line 10 through page 21, line 15; and page 44, line 19 through page 46, line 16. Such disclosures are the same as I read and understood

in my previous Declaration and Supplemental Declaration. A copy of such disclosures is attached hereto as Third Supplemental Declaration Exhibit A.

I have also read and understood additional disclosures of the above-referenced patent application at page 33, lines 8-10; page 37, lines 19-25; page 40, line 20 through page 43 line 3; page 44, lines 12 and 13; page 48, lines 13-15; page 53, line 1 through page 56, line 25; and page 62, lines 1-10. A copy of such additional disclosures is attached hereto as Third Supplemental Declaration Exhibit B.

5. The disclosures in Third Supplemental Declaration Exhibit A, also contained in my previous Declaration and Supplemental Declaration, relate to using growth factors, including cells, for promoting the growth of soft tissue and, more specifically, to a method which may use such growth factors for growing a new portion of a human heart by growing new cardiac muscle. Such disclosures are also directed to the growth of new arteries in the heart.

I understand that the additional disclosures in Third Supplemental Declaration Exhibit B relate to using cellular growth factors, including bone marrow stem cells, to grow soft tissue, including an artery. Stem cells harvested from bone marrow, peripheral blood and from culture banks are described as being implanted for promoting morphogenesis and growth of all three-germ tissue layers, i.e. mesoderm, ectoderm and endoderm tissues. It would be understood by one skilled in the art that morphogenesis includes the growth of an artery, which comprises mesodermal tissue.

6. I have read and understood the claims set forth in Third Supplemental Declaration Exhibit C and have been informed that such claims will be concurrently presented in this application with this Third Supplemental Declaration.

7. Based upon above Paragraphs 4-6, it is my opinion that one skilled in the medical arts, armed with the knowledge in the disclosures referenced therein, would be enabled to practice the method set forth in Third Supplemental Declaration Exhibit C and to predictably anticipate the results defined therein without need for resorting to undue experimentation. It is my further opinion that one skilled in the art reading such disclosures would understand that all of the well known administration procedures described at page 45 of the patent application, including intravenous, intraluminal, intramuscular, and with an angioplasty balloon, would be applicable for use in growing an artery in a human patient regardless of whether the genetic material was a gene; cell, including stem cells such as bone marrow stem cells; or another type of growth factor.

Declarant states that the above opinion was reached independently.

Declarant understands that (1) any willful false statements and the like made herein are punishable by fine or imprisonment, or both (18 U.S.C. 1001) and may jeopardize the validity of the application or any patent issuing thereon, and (2) that all statements made of Declarant's own knowledge are true and that all statements made on information and belief are believed to be true.

Further Declarant sayeth not.

Date: 5 June 2006

Andrew E. Lorincz, M.D.
Andrew E. Lorincz, M.D.

**THIRD
SUPPLEMENTAL
DECLARATION**

EXHIBIT A

DISCLOSURES

EXHIBIT A
DISCLOSURES
APPLICATION SERIAL NO. 09/836,750

PAGE 20, LINE 10 – PAGE 21, LINE 15

Growth factors can be utilized to induce the growth of “hard tissue” or bone and “soft tissues” like ectodermal and mesodermal tissues. As used herein, the term growth factor encompasses compositions and living organisms which promote the growth of hard tissue, such as bone, or soft tissue, in the body of a patient. The compositions include organic and inorganic matter. The compositions can be genetically produced or manipulated. The living organisms can be bacteria, viruses, or any other living organism which promote tissue growth. By way of example and not limitation, growth factors can include platelet-derived growth factor (PDGF), epidermal growth factor (EGF), fibroblast growth factor (acidic/basic (FGF a,b), interleukins (IL’s), tumor necrosis factor (TNF), transforming growth factor (TGF-B), colony-stimulating factor (CSF), osteopontin (Eta-1 OPN), platelet-derived growth factor (PDGF), interferon (INF), bone morphogenic protein 1 (BMP-1), and insulin growth factor (IGF). Recombinant and non-recombinant growth factors can be utilized as desired. Bacteria or viruses can, when appropriate, be utilized as growth factors. For example, there is a bacterial hydrophilic polypeptide that self-assembles into a nanometer internal diameter pore to build a selective lipid body. Various enzymes can be utilized for the synthesis of peptides which contain amino acids that control three-dimensional protein structure and growth. Growth factors can be applied in gels or other carriers which regulate the rate of release of the growth factors and help maintain the growth factors and the carrier, at a desired location in the body. Time release capsules, granules, or other carriers containing growth factor can be activated by tissue pH, by enzymes, by ultrasound,

by electricity, by heat, by selected *in vivo* chemicals or by any other selected means to release the growth factor. The carrier can be resorbable or non-resorbable. Or, the growth factor itself can be activated by similar means. Either the carrier or the growth factor can mimic extracellular fluid to control cell growth, migration, and function. The growth factor can be administered orally, systemically, in a carrier, by hypodermic needle, through the respiratory tract, or by any other desired method. The growth factor can also be administered into a capsule or other man-made composition or structure placed in the body. While administration of the growth factor is presently usually localized in the patient's body, circumstances may arise where it is advantageous to distribute a growth factor throughout the patient's body in uniform or non-uniform concentrations. An advantage to growth factors is that they can often, especially when in capsule form or in some other containment system, be inserted to a desired site in the body by simply making a small incision and inserting the growth factor. The making of such small incision comprises minor surgery which can often be accomplished on an out-patient basis. The growth factors can be multifactorial and nonspecific.

PAGE 44, LINE 19 – PAGE 46, LINE 16

Genetic material comprising a portion of a gene, a gene, genes, a gene product (i.e., a composition a gene causes to be produced like, for example, an organ-producing growth factor), growth factor, or an ECM (extracellular matrix) can be used in or on the body to grow an organ to tissue. For example, the vascular epithelial growth factor gene (VEGF) or its growth factor equivalent can be inserted into the body to cause an artery to grow. When insertion of a gene, portion of a gene, gene product, growth factor, or ECM *in vivo* or *ex vivo* is referred to herein in connection with any of the implant techniques of the invention, it is understood that a cell

nutrient culture(s), physiological nutrient culture(s), carrier (s), enhancer(s), promoter(s), or any other desired auxiliary component(s) can be inserted with the gene or at the same location as the gene, growth factor, ECM, etc.

An artery is an organ from the circulatory system. An artery can be grown in the heart, legs, or other areas by injecting a gene or other genetic material into muscle at a desired site. Size, vascularity, simplicity of access, ease of exploitation, and any other desired factors can be utilized in selecting a desired site. The gene is one of several known VEGF genes which cause the production of vascular endothelial growth factors. Several VEGF genes which produce vascular endothelial growth factors are believed to exist because nature intends for there to be several pathways (i.e., genes) which enable the production of necessary growth factors. The existence of several pathways is believed important because if one of the genes is damaged or inoperative, other similar genes can still orchestrate the production of necessary growth factors. VEGF genes are used by the body to promote blood vessel growth. VEGF genes are assimilated (taken in) by muscle cells. The genes cause the muscle cells to make a VEGF protein which promotes the growth of new arteries. VEGF proteins can be made in a lab and injected into a patient intravenously, intraluminally, or intramuscularly to promote the growth of an artery. Or, the genes (or other genetic material) can be applied with an angioplasty balloon, with the assistance of a vector, or by any other method.

It is not always desirable to grow a completely new organ. Sometimes growing a portion of an organ is desirable. For example, in some heart attacks or strokes, a portion of the heart or brain remains viable and a portion dies. An injection of a gene to form cardiac muscle and/or an injection of a gene to form an artery can be utilized to revive or replace the dead portion of the heart. The dead portion of the heart may (or may not) be used as a matrix while the new muscles

and vessels grow. Thus, in this example, a partial new organ is grown in a pre-existing organ. A pacemaker may (or may not) be necessary. A second injection of a gene may (or may not) be necessary to stop cardiac muscle growth once it is completed. Portions of organs throughout the body can similarly be repaired or replaced. It may be necessary to provide gene(s) or growth factor(s) sequentially. For instance, one or more blood vessels are grown by inserting an appropriate gene or other genetic material into a selected area. Second, an appropriate gene or other genetic material is inserted in the selected area to grow a bone or other organ.

The size and shape limitation of the desired structure can come from a containment and boundary contact inhibition phenomenon or by a chemical inhibition.

A variation on the theme of growing a portion of an organ is as follows: a portion of a heart dies. The pericardium is utilized as a scaffold and seeded with cells and/or genes to grow new muscle, and genes (or other genetic material) to grow new arteries. Immediately adjacent the dead cardiac muscle, onto or into the pericardium, the appropriate cells, genes, and/or growth factors (or other genetic material) are placed. Once the new muscle and blood vessels have grown, the function specific tissue can be applied to the damaged portion of the heart and paced, if necessary, to augment cardiac action. If the surgeon desires, the dead muscle can be removed and the new muscle and blood vessels can be surgically rotated into the excised region and secured. This probably can be done endoscopically. In essence, the pericardium is utilized to allow the new muscle wall to grow. The new muscle wall is then transplanted into the damaged heart wall. This procedure utilizes the body as a factor to grow an organ and/or tissue, after which the organ and/or tissue is transplanted to a desired region. On the other hand, the new muscle wall may integrate itself into the old wall and not require transplantation.

**THIRD
SUPPLEMENTAL
DECLARATION**

EXHIBIT B

DISCLOSURES

EXHIBIT B
DISCLOSURES
APPLICATION SERIAL NO. 09/836,750

PAGE 33, LINES 8-10

Morphogenesis or morphogenetics is the origin and evolution of morphological characters and is the growth and differentiation of cells and tissues during development.

PAGE 37, LINES 19-25

Multifactorial and nonspecific cells (such as stem cells and germinal cells) can provide the necessary in vivo and in vitro cascade of genetic material once an implanted master control gene's transcription has been activated. Likewise, any host cell, clone cell, cultured cell, or cell would work. Genetic switches (such as the insect hormone ecdysone) can be used to control genes inserted into humans and animals. These gene switches can also be used in cultured cells or other cells. Gene switches govern whether a gene is on or off making possible precise time of gene activity.

PAGE 40, LINE 20 – PAGE 43, LINE 3

EXAMPLE 11

MSX-1 and MSX-2 are the homeobox genes that control the generation and growth of a tooth. A sample of skin tissue is removed from the patient and the MSX-1 and MXS-2 homeobox gene(s) are removed from skin tissue cells. The genes are stored in an appropriate nutrient culture medium.

BMP-2 and BMP-4 growth factors are obtained by recombinant or natural extraction from bone.

Living stem cells are harvested from the bone marrow, the blood of the patient, or from cell culture techniques. The stem cells are placed in a nutrient culture medium at 98.6 degrees. The temperature of the culture medium can be varied as desired but ordinarily is between 40 to 102 degrees F.

MXS-1 and MXS-2 transcription factors are obtained which will initiate the expression of the MXS-1 and MXS-2 homeobox genes.

The MXS-1 and MXS-2 transcription factors, BMP-2 and BMP-4 bone morphogenic proteins, and MXS-1 and MXS-2 genes are added to the nutrient culture medium along with the living stem cells.

EXAMPLE 12

Example 11 is repeated except that the transcription factors bind to a receptor complex in the stem cell nucleus.

EXAMPLE 13

Example 11 is repeated except that the MXS-1 and MXS-2 transcription factors are not utilized. The transcription of the MXS-1 and MXS-2 homeobox genes is activated by applying an electric spark to the nutrient culture medium.

EXAMPLE 14

Example 13 is repeated except that the stem cells are starved and the transcription of the MXS-1 and MXS-2 homeobox genes is activated by applying an electric spark to the nutrient culture medium.

EXAMPLE 15

WT-1 and PAX genes are obtained from a sample of skin tissue is removed from the patient. The genes are stored in an appropriate nutrient culture medium. PAX genes produce PAX-2 and other transcription factors.

BMP-7 and other kidney related BMP growth factors are obtained by recombinant or natural extraction from bone.

Living stem cells are harvested from the bone marrow, the blood of the patient, or from cell culture techniques. The stem cells are placed in a nutrient culture medium at 98.6 degrees. The temperature of the culture medium can be varied as desired but ordinarily is between 40 and 102 degrees F.

The WT-1 and PAX genes, and BMP-7 and other kidney BMPS are added to the nutrient culture medium along with the living stem cells.

A primitive kidney germ is produced. The kidney germ is transplanted in the patient's body near a large artery. As the kidney grows, its blood supply will be derived from the artery.

EXAMPLE 16

The Aniridia gene is obtained from a sample of skin tissue is removed from the patient. The gene(s) is stored in an appropriate nutrient culture medium.

Aniridia transcription factor (activates expression of the Aniridia gene) and growth factors (function to help stem cells differentiate during morphogenesis to form an eye) are obtained.

Living stem cells are harvested from the bone marrow, the blood of the patient, or from cell culture techniques. The stem cells are placed in a nutrient culture medium at 98.6 degrees.

The temperature of the culture medium can be varied as desired but ordinarily is between 40 to 102 degrees F.

The Aniridia transcription factor and growth factors and the Aniridia gene are added to the nutrient culture medium along with the living stem cells.

A primitive eye germ is produced. The kidney germ is transplanted in the patient's body near the optic nerve. As the kidney grows, its blood supply will be derived from nearby arteries.

EXAMPLE 17

The Aniridia gene is obtained from a sample of skin tissue is removed from the patient. The gene(s) is stored in an appropriate nutrient culture medium.

Aniridia transcription factor (activates expression of the Aniridia gene) and growth factors (function to help stem cells differentiate during morphogenesis to form an eye) are obtained and added to the nutrient culture medium.

An eye germ develops. A branch of the nearby maxillary artery is translocated to a position adjacent the eye germ to promote the development of the eye germ. The eye germ matures into an eye which receives its blood supply from the maxillary artery.

The term "cell nutrient culture" as used herein can include any or any combination of the following: the extracellular matrix; conventional cell culture nutrients; and/or, a cell nutrient such as a vitamin. As such, the cell nutrient culture can be two-dimensional, three dimensional, or simply a nutrient, and is useful in promoting the processes of cellular dedifferentiation, redifferentiation, differentiation, growth, and development.

PAGE 44, LINES 12– 13

An organ, as used herein, consists of two or more kinds of tissues joined into one structure that has a certain task.

PAGE 48, LINES 13– 15

In the example above, if germinal cells (and in some cases, stem cells) are utilized a direct differentiation and morphogenesis into an organ can occur in vivo, ex vivo, or in vitro.

PAGE 53, LINE 1 – PAGE 56, LINE 25

EXAMPLE 18

A 36 year old Caucasian male experiences pain in his left leg. A medical examination reveals a damaged one inch long section of a large artery in his left leg. The examination also reveals that this damaged section of the artery is nearly completely clogged with plaque and that the wall of the artery is weakened. The weakening in the arterial wall makes attempting to clean out the artery risky and also makes it risky to attempt to insert a stent in the artery.

Recombinant cDNA encoded to combine with a cell ribosome to produce the human growth factor VEGF is assembled into a eukaryotic expression plasmid. The recombinant cDNA is from cDNA libraries prepared from HL60 leukemia cells and is known to cause the growth of arteries. The plasmid is maintained at a room temperature of 76 degrees F.

The clones are placed in 1.0 milliliters of a normal saline carrier solution at a room temperature of 76 degrees F to produce an genetic carrier solution. The genetic carrier solution contains about 250 ug of the cDNA clones. A nutrient culture can, if desired, be utilized in conjunction with or in place of the saline carrier. Each clone is identical. If desired, only a

single clone can be inserted in the normal saline carrier solution. The saline carrier solution comprises 0.09% by weight sodium chloride in water. A saline carrier solution is selected because it will not harm the DNA clone.

Two sites are selected for injection of the genetic carrier solution. While the selection of sites can vary as desired, the sites are selected at the lower end (the end nearest the left foot of the patient) of the damaged section of the artery so that the new arterial section grown can, if necessary, be used to take the place of the damaged section of the artery in the event the damaged section is removed.

The first site is on the exterior wall of the artery on one side of the lower end of the damaged section of the artery. A containment system is placed at the first site.

The second site is inside the wall of the artery on the other side of the lower end of the artery.

The genetic carrier solution is heated to a temperature of 98.6 degrees F. 0.25 milliliters of the genetic carrier solution is injected into the containment system at the first site. 0.25 milliliters of the genetic carrier solution is injected at the second site inside the wall of the artery. Care is taken to slowly inject the genetic carrier solution to avoid entry of the solution into the artery such that blood stream will carry away the cDNA in the solution.

After two weeks, an MRI is taken which shows the patient's leg artery. The MRI reveals new growth at the first and second sites.

After four weeks, another MRI is taken which shows the patient's leg artery. The MRI shows that (1) at the first site a new artery is growing adjacent the patient's original leg artery, and (2) at the second site a new section of artery is growing integral with the original artery, i.e., at the second site the new section of artery is lengthening the original artery, much like inserting

a new section of hose in a garden hose concentric with the longitudinal axis of the garden hose lengthens the garden hose.

After about eight to twelve weeks, another MRI is taken which shows that the new artery growing adjacent the patient's original artery has grown to a length of about one inch and has integrated itself at each of its ends with the original artery such that blood flows through the new section of artery. The MRI also shows that the new artery at the second site has grown to a length of one-half inch.

In any of the examples of the practice of the invention included herein, cell nutrient culture can be included with the gene, the growth factor, the extracellular matrix, or the environmental factors.

In any of the examples of the practice of the invention included herein, the concept of gene redundancy can be applied. For example, the Examples 1 to 14 concerning a tooth list the genes MSX-1 and MSX-2. These genes differ by only two base pairs. Either gene alone may be sufficient. A further example of redundancy occurs in growth factors. Looking at the Examples 10 to 14, BMP4 or BMP2 alone may be sufficient. Redundancy can also be utilized in connection with transcription factors, extracellular matrices, environmental factors, cell nutrient cultures, physiological nutrient cultures, vectors, promoters, etc.

One embodiment of the invention inserts genetic material (gene, growth factor, ECM, etc.) into the body to induce the formation of an organ. Similar inducing materials inserted ex vivo into or onto a living cell in an appropriate physiological nurturing environment will also induce the growth of an organ. The VCSEL laser allows early detection in a living cell of a morphogenic change indicating that organ formation has been initiated. With properly timed transplantation, organ growth completes itself.

During the ex vivo application of the invention, a gene and/or growth factor is inserted into a cell or a group of cells; an ECM or environmental factor(s) are placed around and in contact with a cell or group of cells; or, genetic material is inserted into a subunit of a cell to induce organ growth. An example of a subunit of a cell is an enucleated cell or a comparable artificially produced environment. In in vivo or ex vivo embodiments of the invention to induce the growth of an organ, the genes, growth factors, or other genetic material, as well as the environmental factors or cells utilized, can come from any desired source.

EXAMPLE 19

Genetically produced materials are inserted in the body to cause the body to grow, reproduce, and replace in vivo a clogged artery in the heart. This is an example of site-specific gene expression. A plasmid expression vector containing an enhancer/promoter is utilized to aid in the transfer of the gene into muscle cells. The enhancer is utilized to drive the specific expression of the transcriptional activator. After the enhancer drives the expression of the transcriptional activator, the transcriptional activator transactivates the muscle/artery genes. Saline is used as a carrier. Cardiac muscle can take up naked DNA injected intramuscularly. Injecting plasmid DNA into cardiac (or skeletal) muscle results in expression of the transgene in cardiac myocytes for several weeks or longer.

Readily available off-the-shelf (RAOTS) cDNA clones for recombinant human VEGF165, isolated from cDNA libraries prepared from HL60 leukemia cells, are assembled in a RAOTS expression plasmid utilizing 736 bp CMV promoter/enhancer to drive VEGF expression. Other RAOTS promoters can be utilized to drive VEGF expression for longer periods of time. Other RAOTS recombinant clones of angiogenic growth factors other than VEGF can be utilized, for example, fibroblast growth factor family, endothelial cell growth

factor, etc. Downstream from the VEGF cDNA is an SV40 polyadenylation sequence. These fragments occur in the RAOTS pUC118 vector, which includes an Escherichia coli origin of replication and the Beta lactamase gene for ampicillin resistance.

The RAOTS construct is placed into a RAOTS 3 ml syringe with neutral pH physiologic saline at room temperature (or body temperature of about 37 degrees C). The syringe has a RAOTS 27 gauge needle.

Access to the cardiac muscle is gained by open heart surgery, endoscopic surgery, direction injection of the needle without incision, or by any other desired means. The cardiac muscle immediately adjacent a clogged artery is slowly injected with the RAOTS construct during a five second time period. Injection is slow to avoid leakage through the external covering of muscle cells. About 0.5 ml to 1.0 ml (milliliter) of fluid is injected containing approximately 500 ug phVEGF165 in saline (N=18). The readily available off-the-shelf cDNA clones cause vascular growth which automatically integrates itself with the cardiac muscle. Anatomic evidence of collateral artery formation is observed by the 30th day following injection to the RAOTS construct. One end of the artery integrates itself in the heart wall to receive blood from the heart. The other end of the artery branches into increasing smaller blood vessels to distribute blood into the heart muscle. Once the growth of the new artery is completed, the new artery is left in place in the heart wall. Transplantation of the new artery is not required.

Blood flow through the new artery is calculated in a number of ways. For example, Doppler-derived flow can be determined by electromagnetic flowmeters (using for example, a Doppler Flowmeter sold by Parks Medical Electronic of Aloha, Oregon) both in vitro and in vivo. RAOTS external ultrasound gives a semiquantitative analysis of arterial flow. Also, RAOTS angiograms or any other readily available commercial devices can be utilized.

VEGF gene expression can be evaluated by readily available off-the-shelf polymerase chain reaction (PCR) techniques.

If controls are desired, the plasmid pGSVLacZ containing a nuclear targeted Beta-galactosidase sequence coupled to the simian virus 40 early promoter can be used. To evaluate efficiency, a promoter-matched reporter plasmid, pCMV Beta (available from Clontech of Palo Alto, California), which encodes Beta-galactosidase under control of CMV promoter/enhancer can be utilized. Other RAOTS products can be utilized if desired.

EXAMPLE 20

A patient, a forty year old African-American female in good health, has been missing tooth number 24 for ten years. The space in her mouth in which her number 24 tooth originally resided is empty. All other teeth except tooth number 24 are present in the patient's mouth. The patient desires a new tooth in the empty "number 24" space in her mouth.

A full thickness mucoperiosteal flap surgery is utilized to expose the bone in the number 24 space. A slight tissue reflection into the number 23 tooth and number 25 tooth areas is carried out to insure adequate working conditions.

A Midwest Quietair handpiece (or other off-the-shelf handpiece) utilizing a #701XXL bur (Dentsply Midwest of Des Plaines, Illinois) (a #700, #557, #558, etc. bur can be utilized if desired) is used to excavate an implant opening or site in the bone. The implant opening is placed midway between the roots of the number 23 and number 25 teeth. The opening ends at a depth which is about fifteen millimeters and which approximates the depth of the apices of the roots of the number 23 and number 25 teeth. Care is taken not to perforate either the buccal or lingual wall of the bone. In addition, care is taken not to perforate or invade the periodontal ligament space of teeth numbers 23 and 25.

An interrupted drilling technique is utilized to avoid overheating the bone when the #701XXL bur is utilized to form the implant opening. During a drilling sequence, the drill is operated in five second increments and the handpiece is permitted to stall. Light pressure and a gentle downward stroke are utilized.

PAGE 62, LINES 1-10

EXAMPLE 36

Example 18 is repeated except that the patient is a 55 year old Caucasian male, and the genetic carrier solution is injected into two sites in the coronary artery of the patient. The first site is on the exterior wall on one side of the artery. The second site is inside the wall of the artery on the other side of the artery. A section of the artery is damaged, is partially blocked, and has a weakened wall. The first and second sites are each below the damaged section of the artery. Similar results are obtained, i.e., a new section of artery grows integral with the original artery, and a new section of artery grows adjacent the original artery. The new section of artery has integrated itself at either end with the original artery so that blood flows through the new section of artery.

**THIRD
SUPPLEMENTAL
DECLARATION**

EXHIBIT C

CLAIMS

EXHIBIT C

CLAIMS

APPLICATION SERIAL NO. 09/836,750

- Claim 236 A method of growing a new portion of a pre-existing heart comprising the steps of placing a growth factor in a body of a human patient and growing new cardiac muscle and growing a new artery in said heart.
- Claim 238 The method of claim 236, further comprising repairing a dead portion of said heart.
- Claim 239 The method of claim 236, further comprising repairing a damaged portion of said heart.
- Claim 243 The method of claim 236, wherein said growth factor comprises a member selected from the group consisting of cells, cellular products, and derivatives of cellular products.
- Claim 244 The method of claim 243, wherein said growth factor comprises a cell.
- Claim 245 The method of claim 244, wherein said cell is multifactorial and non-specific.
- Claim 246 The method of claim 245, wherein said cell comprises a stem cell.
- Claim 247 The method of claim 236, wherein said growth factor is placed in said patient by injection.
- Claim 248 The method of claim 247, wherein said injection is intravenous.
- Claim 249 The method of claim 247, wherein said injection is intraluminal.
- Claim 250 The method of claim 247, wherein said injection is intramuscular.

- Claim 251 The method of claim 236, wherein said growth factor is placed in said patient by a carrier.
- Claim 252 The method of claim 251, wherein said carrier comprises an angioplasty balloon.
- Claim 253 The method of claim 236, wherein said growth factor comprises a gene and a cell.
- Claim 257 The method of claim 236, wherein said growth factor is locally placed in said body.
- Claim 258 The method of claim 238, wherein said growth factor is locally placed in said body.
- Claim 259 The method of claim 239, wherein said growth factor is locally placed in said body.
- Claim 260 The method of claim 243, wherein said growth factor is locally placed in said body.
- Claim 261 The method of claim 236, wherein said growth factor comprises living stem cells harvested from bone marrow.
- Claim 262 The method of claim 238, wherein said growth factor comprises living stem cells harvested from bone marrow.
- Claim 263 The method of claim 239, wherein said growth factor comprises living stem cells harvested from bone marrow.
- Claim 264 A method of growing a new portion of a pre-existing heart comprising locally placing a growth factor comprising a stem cell in a body of a human patient to grow new cardiac muscle in said heart.

- Claim 265 The method of claim 264, wherein said stem cell is placed in said patient by injection.
- Claim 266 The method of claim 264, wherein said stem cell comprises living stem cells harvested from bone marrow.
- Claim 267 The method of claim 266, wherein said stem cell is placed in said patient by injection.
- Claim 268 The method of claim 262, wherein said stem cell is placed in said patient by injection.
- Claim 269 The method of claim 263, wherein said stem cell is placed in said patient by injection.
- Claim 270 The method of claim 258, wherein said growth factor comprises a cell and said cell is placed adjacent to said dead portion of said heart.
- Claim 271 The method of claim 259, wherein said growth factor comprises a cell and said cell is placed adjacent to said damaged portion of said heart.
- Claim 272 The method of claim 265, wherein said stem cell is injected into said heart.
- Claim 273 The method of claim 267, wherein said stem cell is injected into said heart.
- Claim 274 The method of claim 238, wherein said growth factor comprises a cell and said cell is placed in said body by intravenous injection.
- Claim 275 The method of claim 239, wherein said growth factor comprises a cell and said cell is placed in said body by intravenous injection.

- Claim 276 The method of claim 238, wherein said growth factor comprises a cell and said cell is placed in said body by intraluminal injection.
- Claim 277 The method of claim 239, wherein said growth factor comprises a cell and said cell is placed in said body by intraluminal injection.
- Claim 278 The method of claim 238, wherein said growth factor comprises a cell and said cell is placed in said body by an angioplasty balloon.
- Claim 279 The method of claim 239, wherein said growth factor comprises a cell and said cell is placed in said body by an angioplasty balloon.
- Claim 280 The method of claim 236 further comprising determining blood flow through said newly grown artery.
- Claim 281 The method of claim 238 further comprising determining blood flow through said newly grown artery.
- Claim 282 The method of claim 239 further comprising determining blood flow through said newly grown artery.
- Claim 283 The method of claim 236 further comprising observing said newly grown artery.
- Claim 284 The method of claim 238 further comprising observing said newly grown artery.
- Claim 285 The method of claim 239 further comprising observing said newly grown artery.

- Claim 286 A method of repairing a dead portion of a pre-existing heart comprising the steps of placing stem cells adjacent said dead portion; forming a new artery in said heart, thereby causing said dead portion of said heart to be repaired.
- Claim 287 The method of claim 286, wherein said stem cells are placed by injection.
- Claim 288 The method of claim 286, wherein said stem cells are placed by intraluminal administration.
- Claim 289 The method of claim 286, wherein said stem cells are placed by an angioplasty balloon.
- Claim 290 A method of repairing a damaged portion of a pre-existing heart comprising the steps of placing stem cells adjacent said damaged portion; forming a new artery in said heart, thereby causing said damaged portion of said heart to be repaired.
- Claim 291 The method of claim 290, wherein said stem cells are placed by injection.
- Claim 292 The method of claim 290, wherein said stem cells are placed by intraluminal administration.
- Claim 293 The method of claim 290, wherein said stem cells are placed by an angioplasty balloon.